

# Product Information & ELISA Manual

Enzyme-linked Immunosorbent Assay  
for quantitative detection.

**Sample Insert for Reference Use Only**

## Contact

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Novus kits are  
guaranteed for 6 months  
from date of receipt.

**For research use only.  
Not for diagnostic or  
therapeutic procedures.**

FOR RESEARCH USE ONLY  
NOT FOR USE IN CLINICAL DIAGNOSTIC PROCEDURES

Version 8.0

## [ INTENDED USE ]

The Chemiluminescent Immunoassay kit is designed for the in vitro sensitive quantitative measurement of PDGF-B in porcine plasma..

## [ REAGENTS AND MATERIALS PROVIDED ]

| Reagents                                     | Quantity | Reagents                  | Quantity |
|----------------------------------------------|----------|---------------------------|----------|
| Pre-coated, ready to use 96-well strip plate | 1        | Plate sealer for 96 wells | 4        |
| Standard                                     | 2        | Standard Diluent          | 1×20mL   |
| Detection Reagent A                          | 1×120μL  | Assay Diluent A           | 1×12mL   |
| Detection Reagent B                          | 1×120μL  | Assay Diluent B           | 1×12mL   |
| Substrate A                                  | 1×10mL   | Substrate B               | 1×2mL    |
| Wash Buffer (30 × concentrate)               | 1×20mL   | Instruction manual        | 1        |

## [ MATERIALS REQUIRED BUT NOT SUPPLIED ]

1. Luminometer capable of reading 96-well microplates with the following parameters:  
lag time 30.0secs; read time 1.0 sec/well .
2. Single or multi-channel pipettes with high precision and disposable tips.
3. Microcentrifuge Tubes.
4. Deionized or distilled water.
5. Absorbent paper for blotting the microplate.
6. Container for Wash Solution.
7. 0.01mol/L (or 1×) Phosphate Buffered Saline(PBS), pH7.0-7.2.

## [ STORAGE OF THE KITS ]

1. **For unopened kit:** The whole kit could be stored at -20°C in shelf life, while up to one month at 4°C. For experiment convenience, reagents could also be stored seperately, **Standard**, **Detection Reagent A**, **Detection Reagent B** and **96-well strip plate** should be stored at -20°C while the others could be at 4 °C.
2. **For used kit:** When the kit is used, the remaining reagents need to be stored according to the above storage condition. Besides, please return the unused wells to the foil pouch containing the desiccant pack, and zip-seal the foil pouch.

### **Note:**

It is highly recommended to use the remaining reagents within 1 month provided this is prior to the expiration date of the kit. For the expiration date of the kit, please refer to the label on the kit box. All components are stable up to the expiration date.

## [ SAMPLE COLLECTION AND STORAGE ]

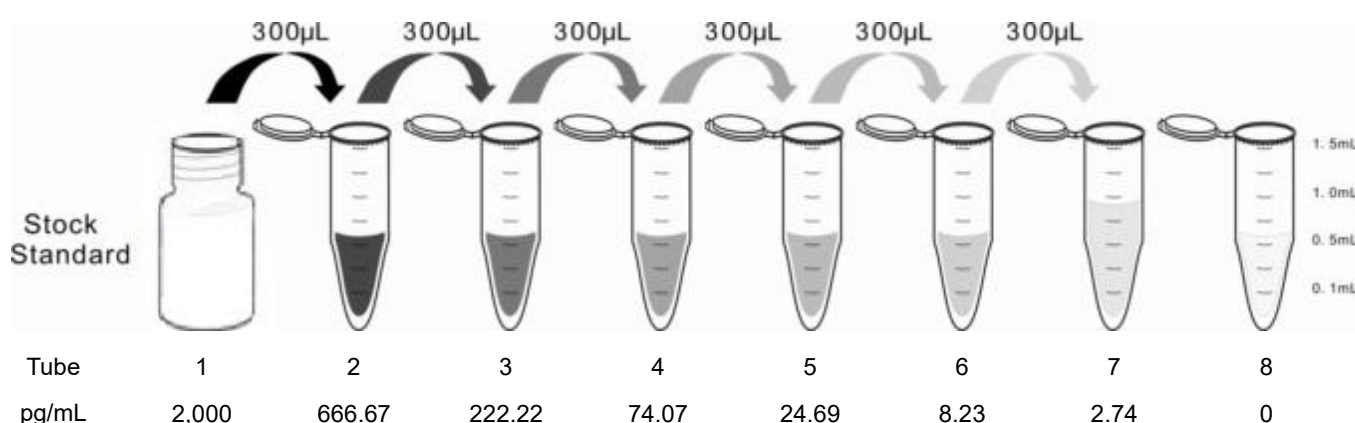
**Plasma** - Collect plasma using 3.8% sodium citrate(sodium citrate :blood = 1: 9) as an anticoagulant. Centrifuge samples for 15 minutes at 1,000×g at 2-8°C within 30 minutes of collection. Remove plasma and assay immediately or store samples in aliquot at -20°C or -80°C for later use. Avoid repeated freeze/thaw cycles.

**Note:**

1. Samples to be used within 5 days may be stored at 4°C, otherwise samples must be stored at -20°C ( $\leq 1$  month) or -80°C ( $\leq 2$  months) to avoid loss of bioactivity and contamination.
2. Sample hemolysis will influence the result, so hemolytic specimen should not be used.
3. When performing the assay, bring samples to room temperature.

## [ REAGENT PREPARATION ]

1. Bring all kit components and samples to room temperature (18-25°C) before use. If the kit will not be used up in one time, please only take out strips and reagents for present experiment, and leave the remaining strips and reagents in required condition.
2. **Standard** - Reconstitute the **Standard** with 1.0mL of **Standard Diluent**, kept for 10 minutes at room temperature, shake gently(not to foam). The concentration of the standard in the stock solution is 2,000pg/mL. Please prepare 7 tubes containing 0.6mL Standard Diluent and produce a triple dilution series according to the picture shown below. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 2,000pg/mL, 666.67pg/mL, 222.22pg/mL, 74.07pg/mL, 24.69pg/mL, 8.23pg/mL, 2.74pg/mL, and the last EP tubes with **Standard Diluent** is the blank as 0pg/mL.



3. **Detection Reagent A and Detection Reagent B** - Briefly spin or centrifuge the stock Detection A and Detection B before use. Dilute them to the working concentration 100-fold with **Assay Diluent A** and **B**, respectively.
4. **Wash Solution** - Dilute 20mL of Wash Solution concentrate (30×) with 580mL of deionized or distilled water to prepare 600mL of Wash Solution (1×).
5. **Substrate working Solution** - Mix the substrate A and B by the ratio of 99:1 to make the substrate working solution. Mix thoroughly. For example, prepare 1,000µL Substrate working Solution with 990µL Substrate A + 10µL Substrate B.

**Note:**

1. Making serial dilution in the wells directly is not permitted.
2. Prepare standard within 15 minutes before assay. Please do not dissolve the reagents at 37°C directly.
3. Please carefully reconstitute Standards or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals are completely dissolved. To minimize imprecision caused

by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10 $\mu$ L for once pipetting.

4. The reconstituted Standards, Detection Reagent A and Detection Reagent B can be **used only once**.
5. Prepare Substrate working Solution within 15 minutes before assay.
6. If crystals have formed in the Wash Solution concentrate (30 $\times$ ), warm to room temperature and mix gently until the crystals are completely dissolved.
7. Contaminated water or container for reagent preparation will influence the detection result.

## **[ SAMPLE PREPARATION ]**

1. We are only responsible for the kit itself, but not for the samples consumed during the assay. The user should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance.
2. Please predict the concentration before assaying. If values for these are not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments. Sample should be diluted by PBS.
3. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
4. Tissue or cell extraction samples prepared by chemical lysis buffer may cause unexpected CLIA results due to the impacts from certain chemicals.
5. Due to the possibility of mismatching between antigen from other origin and antibody used in our kits (e.g., antibody targets conformational epitope rather than linear epitope), some native or recombinant proteins from other manufacturers may not be recognized by our products.
6. Influenced by the factors including cell viability, cell number or sampling time, samples from cell culture supernates may not be detected by the kit.
7. Fresh samples without long time storage is recommended for the test. Otherwise, protein degradation and denaturalization may occur in those samples and finally lead to wrong results.

## **[ ASSAY PROCEDURE ]**

1. Determine wells for diluted standard, blank and sample. Prepare 7 wells for standard, 1 well for blank. Add 100 $\mu$ L each of dilutions of standard (read Reagent Preparation), blank and samples into the appropriate wells. Cover with the Plate sealer. Incubate for 1 hour at 37°C.
2. Remove the liquid of each well, don't wash.
3. Add 100 $\mu$ L of **Detection Reagent A** working solution to each well, cover the wells with the plate sealer and incubate for 1 hour at 37°C.
4. Aspirate the solution and wash with 350 $\mu$ L of 1 $\times$  Wash Solution to each well using a squirt bottle, multi-channel pipette, manifold dispenser or autowasher, and let it sit for 1~2 minutes. Remove the remaining liquid from all wells completely by snapping the plate onto absorbent paper. Totally wash 3 times. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against absorbent paper.
5. Add 100 $\mu$ L of **Detection Reagent B** working solution to each well, cover the wells with the plate sealer and incubate for 30 minutes at 37°C.
6. Repeat the aspiration/wash process for five times as conducted in step 4.

7. Add 100µL of **Substrate working Solution** to each well. Cover with a new Plate sealer. Incubate for 10 minutes at 37°C. Protect from light.
8. Measure the chemiluminescence signal in a microplate luminometer or as appropriate for the instrument used.

**Note:**

1. **Assay preparation:** Keep appropriate numbers of wells for each experiment and remove extra wells from microplate. Rest wells should be resealed and stored at -20°C.
2. **Samples or reagents addition: Please use the freshly prepared Standard.** Please carefully add samples to wells and mix gently to avoid foaming. Do not touch the well wall. For each step in the procedure, total dispensing time for addition of reagents or samples to the assay plate should not exceed 10 minutes. This will ensure equal elapsed time for each pipetting step, without interruption. Duplication of all standards and specimens, although not required, is recommended. To avoid cross-contamination, change pipette tips between additions of standards, samples, and reagents. Also, use separated reservoirs for each reagent.
3. **Incubation:** To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary. Do not allow wells to sit uncovered for extended periods between incubation steps. Once reagents are added to the well strips, DO NOT let the strips DRY at any time during the assay. Incubation time and temperature must be controlled.
4. **Washing:** The wash procedure is critical. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Solution by aspirating or decanting and remove any drop of water and fingerprint on the bottom of the plate. Insufficient washing will result in poor precision and false elevated absorbance reading.
5. **For Substrate A and B**, please protect it from light.
6. Relative light units (RLUs) may differ from different luminometers. The Immunoassay was optimized using a Beijing Hamamatsu luminometer. Other instruments may require settings to be adjusted.

## **[ TEST PRINCIPLE ]**

The microplate provided in this kit has been pre-coated with an antibody specific to PDGF-B. Standards or samples are then added to the appropriate microplate wells with a biotin-conjugated antibody specific to PDGF-B. Next, Avidin conjugated to Horseradish Peroxidase (HRP) is added to each microplate well and incubated. Then the mixture of substrate A and B is added to generate glow light emission kinetics. Upon plate development, the intensity of the emitted light is proportional to the PDGF-B level in the sample or standard.

## **[ ASSAY PROCEDURE SUMMARY ]**

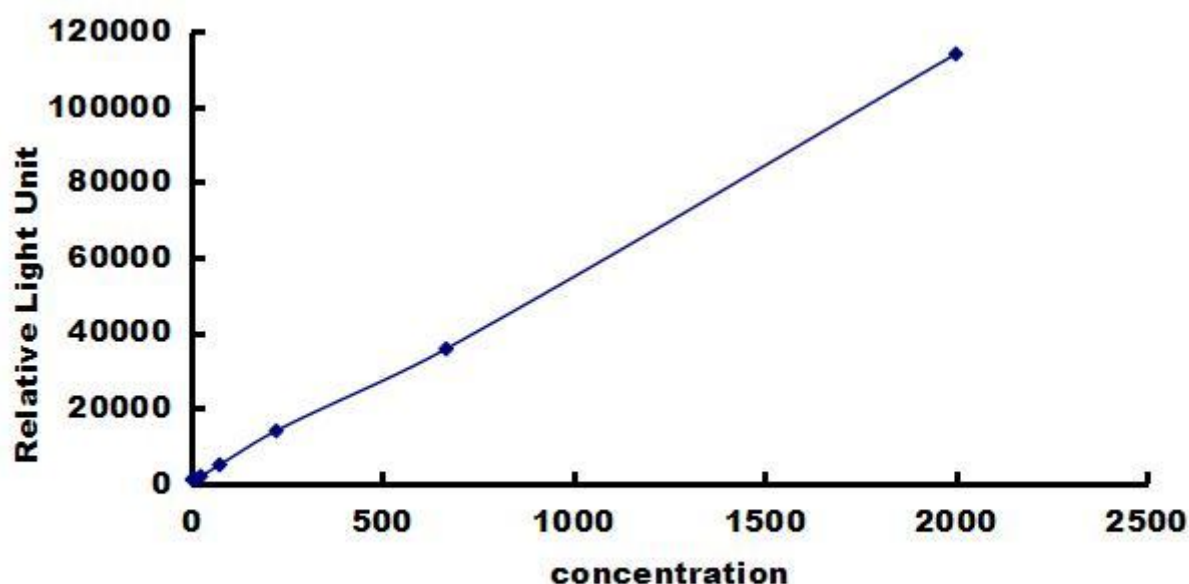
1. Prepare all reagents, samples and standards;
2. Add 100µL standard or sample to each well. Incubate 1 hour at 37°C;
3. Aspirate and add 100µL prepared Detection Reagent A. Incubate 1 hour at 37°C;
4. Aspirate and wash 3 times;
5. Add 100µL prepared Detection Reagent B. Incubate 30 minutes at 37°C;
6. Aspirate and wash 5 times;
7. Add 100µL Substrate Solution. Incubate 10 minutes at 37°C;
8. Read RLU value immediately.

## **[ CALCULATION OF RESULTS ]**

Average the duplicate readings for each standard, control, and samples and subtract the average zero standard relative light unit (RLU). Create a standard curve with PDGF-B concentration on the x-axis and the RLU value on the y-axis. Draw a best fit curve through the points and it can be determined by regression analysis. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

## **[ TYPICAL DATA ]**

Plot the RLU value of the standard on the Y-axis against the known concentration of the standard on the X-axis. Further, in this part, in order to help the customer perform the assay more visual, we provide the customer with the raw data (not the log of data). However, plotting log of the data to construct the curve will be recommended. The RLU values of the standard curve may vary according to the conditions of assay performance (e.g. operator, pipetting technique, washing technique or temperature effects). This curve is provided for demonstration only. The customers should establish their own standard curve for each test conducted.



**Typical Standard Curve for PDGF-B, Porcine CLIA.**

## **[ DETECTION RANGE ]**

2.74-2,000pg/mL. The standard curve concentrations used for the CLIA's were 2,000pg/mL, 666.67pg/mL, 222.22pg/mL, 74.07pg/mL, 24.69pg/mL, 8.23pg/mL, 2.74pg/mL.

## **[ SENSITIVITY ]**

The minimum detectable dose of PDGF-B is typically less than 0.95pg/mL.

The sensitivity of this assay, or Lower Limit of Detection (LLD) was defined as the lowest protein concentration that could be differentiated from zero. It was determined the mean RLU value of 20 replicates of the zero standard added by their two standard deviations.

## **[ SPECIFICITY ]**

This assay has high sensitivity and excellent specificity for detection of PDGF-B.

No significant cross-reactivity or interference between PDGF-B and analogues was observed.

**Note:**

Limited by current skills and knowledge, it is impossible for us to complete the cross- reactivity detection between PDGF-B and all the analogues, therefore, cross reaction may still exist.

### **[ RECOVERY ]**

Matrices listed below were spiked with certain level of recombinant PDGF-B and the recovery rates were calculated by comparing the measured value to the expected amount of PDGF-B in samples.

| Matrix                     | Recovery range (%) | Average(%) |
|----------------------------|--------------------|------------|
| sodium citrate plasma(n=5) | 99-105             | 101        |

### **[ LINEARITY ]**

The linearity of the kit was assayed by testing samples spiked with appropriate concentration of PDGF-B and their serial dilutions. The results were demonstrated by the percentage of calculated concentration to the expected.

| Sample                     | 1: 2    | 1: 4    | 1: 8   | 1: 16  |
|----------------------------|---------|---------|--------|--------|
| sodium citrate plasma(n=5) | 86-103% | 90-101% | 83-97% | 85-96% |

### **[ PRECISION ]**

Intra-assay Precision (Precision within an assay): 3 samples with low, middle and high level PDGF-B were tested 20 times on one plate, respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, middle and high level PDGF-B were tested on 3 different plates, 8 replicates in each plate.

$CV(\%) = SD/mean \times 100$

Intra-Assay:  $CV < 10\%$

Inter-Assay:  $CV < 12\%$

### **[ STABILITY ]**

The stability of CLIA kit is determined by the loss rate of activity. The loss rate of this kit is less than 5% within the expiration date under appropriate storage condition.

To minimize extra influence on the performance, operation procedures and lab conditions, especially room temperature, air humidity, incubator temperature should be strictly controlled. It is also strongly suggested that the whole assay is performed by the same operator from the beginning to the end.

### **[ IMPORTANT NOTE ]**

1. Limited by the current conditions and scientific technology, we can't completely conduct the comprehensive identification and analysis on the raw material provided by suppliers. So there might be some qualitative and technical risks to use the kit.
2. The final experimental results will be closely related to validity of the products, so the kit should be used prior to the expiration date. And please store the kits exactly according to the instruction.
3. Kits from different batches may be a little different in detection range, sensitivity and color developing time.

Please perform the experiment exactly according to the instruction attached in kit while electronic ones from our website is only for reference.

4. Do not mix or substitute reagents from one kit lot to another. Use only the reagents supplied by manufacturer.
5. Protect all reagents from strong light during storage and incubation. All the bottle caps of reagents should be covered tightly to prevent the evaporation and contamination of microorganism. TMB Substrate should remain colorless till it is reacted with the enzyme which binds to the microplate.
6. There may be some foggy substance in the wells when the plate is opened at the first time. It will not have any effect on the final assay results. Do not remove microplate from the storage bag until needed.
7. Wrong operations during the reagents preparation and loading, as well as incorrect parameter setting for the plate reader may lead to incorrect results. It is recommended to use a single-mode luminometer or a multi-mode detection instrument set for luminescence measurement to measure light emission from 96-well microplates.
8. Variation in sample preparation and each step of experimental operation may cause different results. In order to get better reproducible results, the operation of each step in the assay should be controlled.
9. Each kit has been strictly passed Q.C test. However, results from end users might be inconsistent with our in-house data due to some unexpected transportation conditions or different lab equipments. Intra-assay variance among kits from different batches might arise from above factors, too.
10. Kits from different manufacturers with the same item might produce different results, since we haven't compared our products with other manufacturers.
11. The standard of the kit and immunogen used for antibody preparation are commonly recombinant proteins, as different fragments, expression systems, purification methods might be used in recombinant protein preparation, we can not guarantee the kit could detect recombinant protein from other companies. So, it is not recommended to use the kit for the detection of recombinant protein.
12. Please predict the concentration of target molecules in samples, or arrange a preliminary experiment, it is a good way to solve specific problem, e.g. the concentration of samples are beyond the detection range of the kit.
13. The kit might not be suitable for detection of samples from some special experiment, for instance, knock-out experiments, due to their uncertainty of effectiveness.
14. The kit is designed for research use only, we will not be responsible for any issue if the kit was used in clinical diagnostic or any other procedures.

## [ **TROUBLE SHOOTING** ]

| Problem                    | Possible Source                             | Correction Action                                       |
|----------------------------|---------------------------------------------|---------------------------------------------------------|
| <b>Poor Standard Curve</b> | Improper standard curve preparation         | Ensure accurate operation of the dilution               |
|                            | Incomplete washing and aspiration           | Adequate washing and adequate aspiration                |
|                            | Inaccurate Pipetting                        | Check and Calibrate pipettes                            |
| <b>Poor Precision</b>      | Incomplete washing of wells                 | Ensure sufficient washing                               |
|                            | Inadequate mixing and aspiration reagents   | Adequate aspiration and mixing reagents                 |
|                            | Reused pipette tips, containers and sealers | Change and use new pipette tips, containers and sealers |

|                               |                                            |                                                             |
|-------------------------------|--------------------------------------------|-------------------------------------------------------------|
|                               | Inaccurate Pipetting                       | Check and Calibrate pipettes                                |
| <b>Low<br/>RLU<br/>Values</b> | Inadequate reagent volumes added to wells  | Calibrate pipettes and Add adequate reagents                |
|                               | Incorrect incubation times                 | Ensure sufficient incubation times                          |
|                               | Incorrect incubation temperature           | Reagents balanced to room temperature                       |
|                               | Conjugate or substrate reagent failure     | Mix conjugate & substrate, color should develop immediately |
| <b>Sample<br/>Values</b>      | Improper Sample Storage                    | Store the sample properly and use the fresh sample          |
|                               | Improper sample collection and preparation | Take proper sample collection and preparation method        |
|                               | Low quantity of analyte in samples         | Use new sample and repeat assay                             |