

Product Datasheet

beta-Catenin Antibody (12F7) - BSA Free NBP1-54467

Unit Size: 0.1 ml

Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.

www.novusbio.com



technical@novusbio.com

Reviews: 4 **Publications: 22**

Protocols, Publications, Related Products, Reviews, Research Tools and Images at:
www.novusbio.com/NBP1-54467

Updated 9/9/2025 v.20.1

Earn rewards for product
reviews and publications.

Submit a publication at www.novusbio.com/publications

Submit a review at www.novusbio.com/reviews/destination/NBP1-54467



NBP1-54467

beta-Catenin Antibody (12F7) - BSA Free

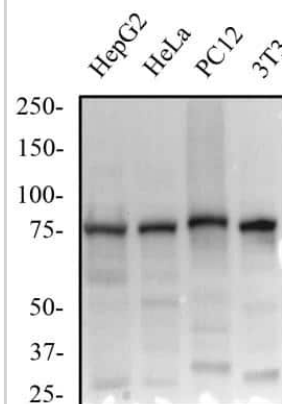
Product Information	
Unit Size	0.1 ml
Concentration	1.0 mg/ml
Storage	Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.
Clonality	Monoclonal
Clone	12F7
Preservative	0.02% Sodium Azide
Isotype	IgG1
Purity	Protein G purified
Buffer	PBS
Target Molecular Weight	92 kDa

Product Description	
Description	Novus Biologicals Mouse beta-Catenin Antibody (12F7) - BSA Free (NBP1-54467) is a monoclonal antibody validated for use in IHC, WB, Flow, ICC/IF, Simple Western and IP. Anti-beta-Catenin Antibody: Cited in 19 publications. All Novus Biologicals antibodies are covered by our 100% guarantee.
Host	Mouse
Gene ID	1499
Gene Symbol	CTNNB1
Species	Human, Mouse, Rat, Duck, Chicken, Primate
Reactivity Notes	Use in Duck reported in scientific literature (PMID:32692763).
Marker	Epithelial Cell Marker, Adherens Junction Marker
Immunogen	Recombinant chicken beta Catenin fused to maltose binding protein. [UniProt# O42486]

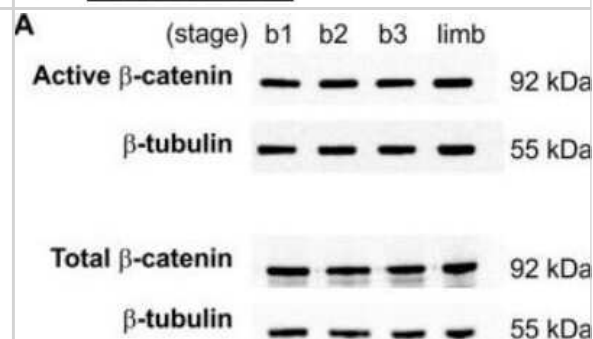
Product Application Details	
Applications	Western Blot, Simple Western, Immunohistochemistry-Paraffin, Flow Cytometry, Flow (Intracellular), Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunoprecipitation
Recommended Dilutions	Western Blot 1:1000, Simple Western 1:100, Flow Cytometry, Immunohistochemistry 1:100-1:200, Immunocytochemistry/ Immunofluorescence 1:50-1:100, Immunoprecipitation 1:10-1:500, Immunohistochemistry-Paraffin 1:100-1:200, Flow (Intracellular)
Application Notes	<p>This beta Catenin (12F7) antibody is useful for IHC-P sections, ICC/IF, IP and WB, where a band can be seen at approx. 92 kDa.</p> <p>In Simple Western only 10 - 15 uL of the recommended dilution is used per data point.</p> <p>See Simple Western Antibody Database for Simple Western validation: Tested in HepG2 lysate 0.5 mg/mL, separated by Size, antibody dilution of 1:100, apparent MW was 90 kDa. Separated by Size-Wes, Sally Sue/Peggy Sue.</p>

Images

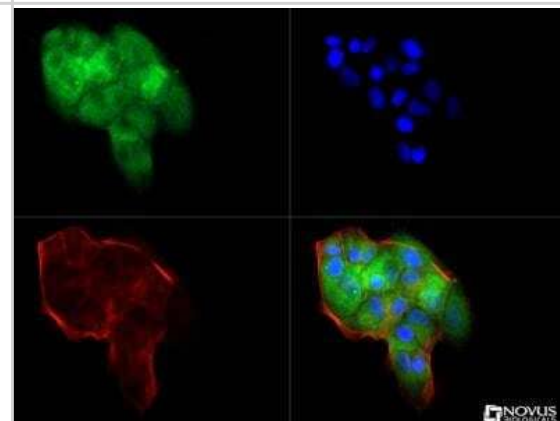
Western Blot: beta-Catenin Antibody (12F7) [NBP1-54467] - Total protein from human HepG2 and HeLa cells, rat PC12 cells and mouse 3T3 cells was separated on a 7.5% gel by SDS-PAGE, transferred to PVDF membrane and blocked in 5% non-fat milk in TBST. The membrane was probed with 1.0 ug/ml anti-Beta catenin in 1% non-fat milk in TBST and detected with an anti-mouse HRP secondary antibody using chemiluminescence.



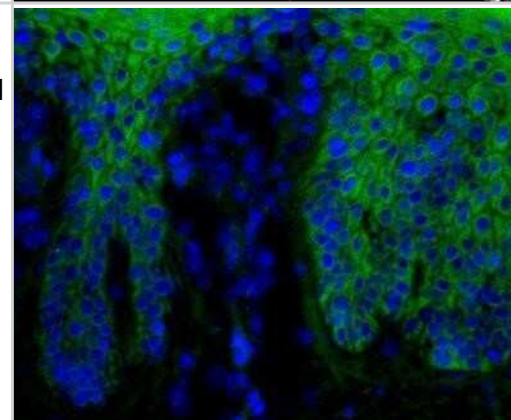
Western Blot: beta-Catenin Antibody (12F7) [NBP1-54467] - Activity of Wnt/beta-catenin pathway in the embryonic chick lung. Western blot analysis of active and total beta-catenin in stage b1, b2 and b3 lungs, and stage 24 limb (as positive control). Control loading was performed using beta-tubulin (55 kDa). Total and active beta-catenin correspond to 92 kDa. Image collected and cropped by CiteAb from the following publication (<https://dx.plos.org/10.1371/journal.pone.0112388>), licensed under a CC-BY license.



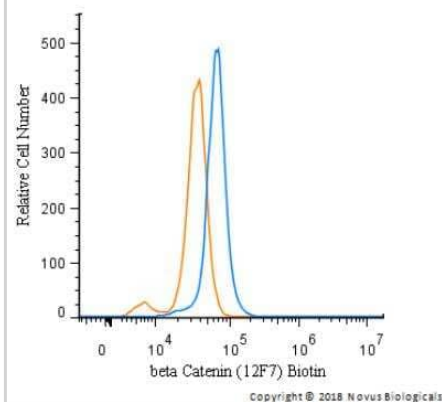
Immunocytochemistry/Immunofluorescence: beta-Catenin Antibody (12F7) [NBP1-54467] - The beta- Catenin antibody was tested in MCF-7 cells against Dylight 488 (Green). Actin and nuclei were counterstained against Phalloidin 550 (Red) and DAPI (Blue).



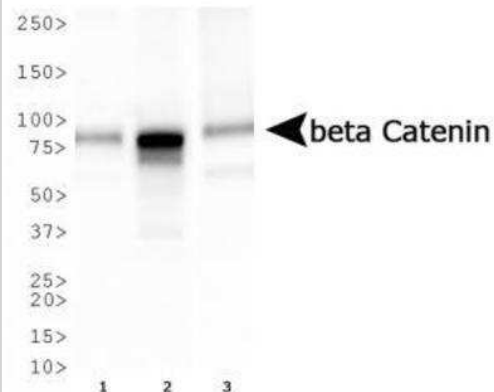
Immunohistochemistry: beta-Catenin Antibody (12F7) [NBP1-54467] - Beta-catenin (green) was detected in human skin (psoriasis) using beta-catenin-FITC antibody (1:40) in PBS for 1 hour. Nuclei were stained with Dapi (blue). Image from verified customer review. Image using the FITC format of this antibody.



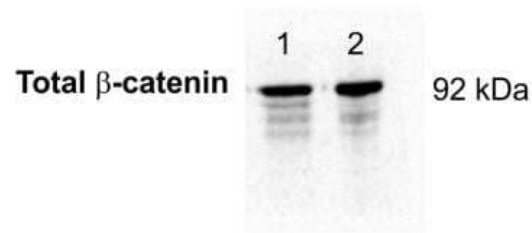
Flow Cytometry: beta-Catenin Antibody (12F7) [NBP1-54467] - An intracellular stain was performed on SK-MEL-28 cells with beta-Catenin Antibody [12F7] NBP1-54467B (blue) and a matched isotype control (orange). Both antibodies were conjugated to Biotin. Cells were fixed with 4% PFA and then permeabilized with 0.1% saponin. Cells were incubated in an antibody dilution of 2.5 ug/mL for 30 minutes at room temperature, followed by Streptavidin - R-Phycoerythrin Protein (2012-1000, Novus Biologicals).



Western Blot: beta-Catenin Antibody (12F7) [NBP1-54467] - Analysis of beta Catenin expression in 1) HepG2, 2) MCF7, and 3) Cos7 whole cell lysates using NBP1-54467.



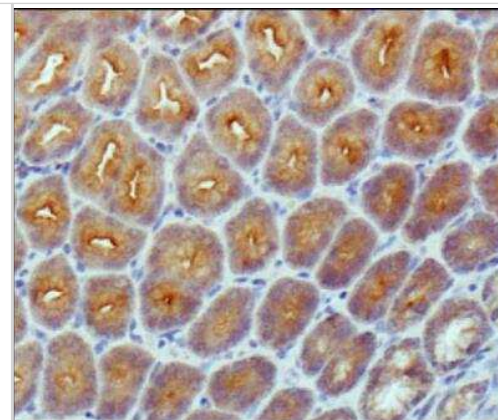
Western Blot: beta-Catenin Antibody (12F7) [NBP1-54467] - Analysis of beta- Catenin in embryonic lung (lane 1) and embryonic limb (lane 2) lysates using anti-beta- Catenin antibody. Each lane was loaded with 5ug of protein sample. Image from verified customer review.



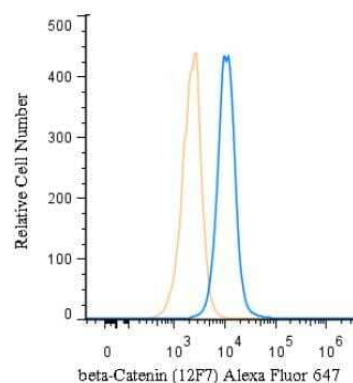
Immunohistochemistry-Paraffin: beta-Catenin Antibody (12F7) [NBP1-54467] - Analysis of beta Catenin in mouse intestine using DAB with hematoxylin counterstain.



Immunohistochemistry-Paraffin: beta-Catenin Antibody (12F7) [NBP1-54467] - Analysis of beta- Catenin in mouse intestine using DAB with hematoxylin counterstain.



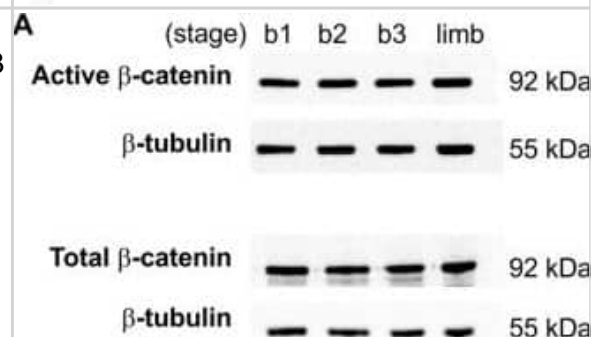
Flow (Intracellular): beta-Catenin Antibody (12F7) [NBP1-54467] - An intracellular stain was performed on HeLa cells with beta-Catenin Antibody (12F7) NBP1-54467AF647 (blue) and a matched isotype control (orange). Cells were fixed with 4% PFA and then permeabilized with 0.1% saponin. Cells were incubated in an antibody dilution of 5 ug/mL for 30 minutes at room temperature. Both antibodies were conjugated to Alexa Fluor 647.



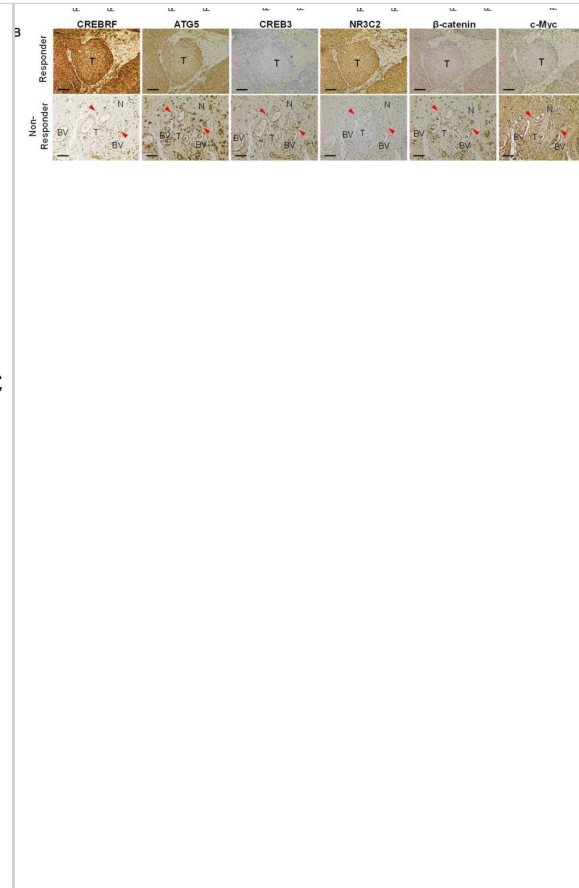
Simple Western: beta-Catenin Antibody (12F7) [NBP1-54467] - Simple Western lane view shows a specific band for Beta- Catenin in 0.5 mg/ml of HepG2 lysate. This experiment was performed under reducing conditions using the 12-230 kDa separation system.



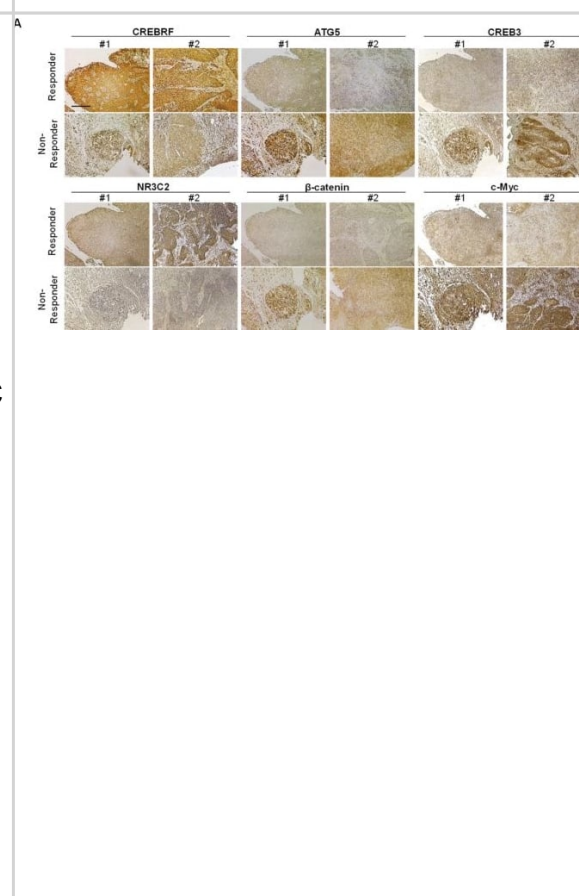
Activity of Wnt/ β -catenin pathway in the embryonic chick lung. (A) Western blot analysis of active and total β -catenin in stage b1, b2 and b3 lungs, and stage 24 limb (as positive control). Control loading was performed using β -tubulin (55 kDa). Total and active β -catenin correspond to 92 kDa. (B) Semi-quantitative analysis for active and total β -catenin. Results are presented as arbitrary units normalized for β -tubulin. $p < 0.05$: * vs limb.



Downregulation of CREBRF and NR3C2 increase poor prognosis in HNSCC. (A) Histological analysis of miR124-3p and miR766-3p target gene expression (CREBRF-ATG5/CREB3, NR3C2- β -catenin/c-Myc) in Responder vs. Non-Responder HNSCC clinical samples. Magnification, x100. Scale bar, 210 μ m. The quantitative data from all specimens are shown in the bar chart. Each dot in the graph represents an individual clinical sample. Two-sided unpaired Student t test was used to analyze comparisons, and data are presented as means \pm SEM. * $p < 0.05$ and *** $p < 0.001$. (B) Histological analysis of tumor morphology in relation to miR124-3p and miR766-3p target gene expression. Representative images of CREBRF, ATG5, CREB3, NR3C2, β -catenin, and c-Myc expression in the serial section of responder and non-responder HNSCC specimens. BV: Blood Vessel. T: Tumor. N: Normal tissue. The invasive cancer cells are indicated by red arrowhead. Magnification, $\times 200$. Scale bar, 100 μ m. (C) Summary of resistance mechanisms regulated by miR124-3p and miR766-3p. Our data indicated that upon acquired resistance in HNSCC cells or in non-responder HNSCC tumors, the levels of miR124-3p and miR766-3p go up, which in turn down-regulate its direct target genes: CREBRF and NR3C2, and consequently the expression of downstream targets of CREBRF (ATG5/CREB3) and NR3C2 (β -catenin/c-Myc) increased in resistant tumors, which are positively correlated with poor prognosis. Thus, by enhancing the CREBRF-ATG5/CREB3 and NR3C2- β -catenin/c-Myc axis, miR124-3p and miR766-3p support aggressive HNSCC progression. Image collected and cropped by CiteAb from the following open publication (<https://pubmed.ncbi.nlm.nih.gov/36358691>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Downregulation of CREBRF and NR3C2 increase poor prognosis in HNSCC. (A) Histological analysis of miR124-3p and miR766-3p target gene expression (CREBRF-ATG5/CREB3, NR3C2- β -catenin/c-Myc) in Responder vs. Non-Responder HNSCC clinical samples. Magnification, x100. Scale bar, 210 μ m. The quantitative data from all specimens are shown in the bar chart. Each dot in the graph represents an individual clinical sample. Two-sided unpaired Student t test was used to analyze comparisons, and data are presented as means \pm SEM. * $p < 0.05$ and *** $p < 0.001$. (B) Histological analysis of tumor morphology in relation to miR124-3p and miR766-3p target gene expression. Representative images of CREBRF, ATG5, CREB3, NR3C2, β -catenin, and c-Myc expression in the serial section of responder and non-responder HNSCC specimens. BV: Blood Vessel. T: Tumor. N: Normal tissue. The invasive cancer cells are indicated by red arrowhead. Magnification, $\times 200$. Scale bar, 100 μ m. (C) Summary of resistance mechanisms regulated by miR124-3p and miR766-3p. Our data indicated that upon acquired resistance in HNSCC cells or in non-responder HNSCC tumors, the levels of miR124-3p and miR766-3p go up, which in turn down-regulate its direct target genes: CREBRF and NR3C2, and consequently the expression of downstream targets of CREBRF (ATG5/CREB3) and NR3C2 (β -catenin/c-Myc) increased in resistant tumors, which are positively correlated with poor prognosis. Thus, by enhancing the CREBRF-ATG5/CREB3 and NR3C2- β -catenin/c-Myc axis, miR124-3p and miR766-3p support aggressive HNSCC progression. Image collected and cropped by CiteAb from the following open publication (<https://pubmed.ncbi.nlm.nih.gov/36358691>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Publications

Shin-Ichiro Hino, Kiyoka Inenaga, Takuto Miyazaki, Chika Tanaka-Mizota Suppression of HCT116 Human Colon Cancer Cell Motility by Polymethoxyflavones is Associated with Inhibition of Wnt/ β -Catenin Signaling. *Nutrition and cancer* 2022-09-08 [PMID: 35658755]

Aamir K, Sethi G, Afrin MR et al. Arjunolic acid modulate pancreatic dysfunction by ameliorating pattern recognition receptor and canonical Wnt pathway activation in type 2 diabetic rats *Life sciences* 2023-08-15 [PMID: 37307966] (Simple Western, IHC, Rat)

Urasaki Y, Le TT Functional Complementation of Anti-Adipogenic Phytonutrients for Obesity Prevention and Management *Nutrients* 2022-10-16 [PMID: 36297009] (WB, Human)

Bhatia R, Thompson CM, Clement EJ et al. Malondialdehyde-Acetaldehyde Extracellular Matrix Protein Adducts Attenuate Unfolded Protein Response During Alcohol and Smoking-Induced Pancreatitis *Gastroenterology* 2022-07-03 [PMID: 35788346] (ICC/IF)

Details:

Fig, 6E.

Cameron S Anti-Cancer and Stress Response Pathway Effects of Nanosilver and Sodium Ascorbate Carleton University 2022-07-11 (WB, Human)

Clinch M The Role of Hypoxia on PORCN and WLS Expression in Human Embryonic Kidney (HEK293T) and Human Colon Cancer (HCT-116T) Cells Carleton University 2022-04-05 (WB, Human)

Sheng J Cellular Effects Nanosilver on Cancer and Non-cancer Cells: Potential Environmental and Human Health Impacts Thesis

Sidor J, Gillette M, Dezi L et al. Role of Presenilin-1 in Aggressive Human Melanoma *International Journal of Molecular Sciences* 2022-04-28 [PMID: 35563300] (WB, Human)

Urasaki, Y, Beaumont, C Et al. Potency Assessment of CBD Oils by Their Effects on Cell Signaling Pathways. *Nutrients* 2020-01-30 [PMID: 32019055] (WB, Human)

Gul HF, Ilhan N, Ilhan N et al. The Combined Effect of Pomegranate Extract and Tangeretin on the DMBA-induced Breast Cancer Model *The Journal of nutritional biochemistry* 2020-12-13 [PMID: 33326843] (WB, Rat)

GonCalves AN, Correia-Pinto J, Nogueira-Silva C ROBO2 signaling in lung development regulates SOX2/SOX9 balance, branching morphogenesis and is dysregulated in nitrofen-induced congenital diaphragmatic hernia *Respir Res* 2020-11-18 [PMID: 33208157] (WB)

Liu Z, Selby CP, Yang Y et al. Circadian regulation of c-MYC in mice *Proc. Natl. Acad. Sci. U.S.A.* 2020-08-19 [PMID: 32817420] (WB, Mouse)

More publications at <http://www.novusbio.com/NBP1-54467>

Procedures

Western Blot protocol for beta-Catenin Antibody (NBP1-54467)

Western Blot Protocol

1. Perform SDS-PAGE on samples to be analyzed, loading 40 ug of total protein per lane.
2. Transfer proteins to membrane according to the instructions provided by the manufacturer of the membrane and transfer apparatus.
3. Stain according to standard Ponceau S procedure (or similar product) to assess transfer success, and mark molecular weight standards where appropriate.
4. Rinse the blot.
5. Block the membrane using standard blocking buffer for at least 1 hour.
6. Wash the membrane in wash buffer three times for 10 minutes each.
7. Dilute primary antibody in blocking buffer and incubate 1 hour at room temperature.
8. Wash the membrane in wash buffer three times for 10 minutes each.
9. Apply the diluted HRP conjugated secondary antibody in blocking buffer (as per manufacturers instructions) and incubate 1 hour at room temperature.
10. Wash the blot in wash buffer three times for 10 minutes each (this step can be repeated as required to reduce background).
11. Apply the detection reagent of choice in accordance with the manufacturers instructions.

Note: Tween-20 can be added to the blocking or antibody dilution buffer at a final concentration of 0.05-0.2%.

*The above information is only intended as a guide. The researcher should determine what protocol best meets their needs. Please follow safe laboratory procedures.

Immunohistochemistry-Paraffin protocol for beta-Catenin Antibody (NBP1-54467)

Immunohistochemistry-Paraffin Embedded Sections

Antigen Unmasking:

Bring slides to a boil in 10 mM sodium citrate buffer (pH 6.0) then maintain at a sub-boiling temperature for 10 minutes. Cool slides on bench-top for 30 minutes.

Staining:

1. Wash sections in deionized water three times for 5 minutes each.
2. Wash sections in wash buffer for 5 minutes.
3. Block each section with 100-400 ul blocking solution for 1 hour at room temperature.
4. Remove blocking solution and add 100-400 ul diluted primary antibody. Incubate overnight at 4 C.
5. Remove antibody solution and wash sections in wash buffer three times for 5 minutes each.
6. Add 100-400 ul biotinylated diluted secondary antibody. Incubate 30 minutes at room temperature.
7. Remove secondary antibody solution and wash sections three times with wash buffer for 5 minutes each.
8. Add 100-400 ul Streptavidin-HRP reagent to each section and incubate for 30 minutes at room temperature.
9. Wash sections three times in wash buffer for 5 minutes each.
10. Add 100-400 ul DAB substrate to each section and monitor staining closely.
11. As soon as the sections develop, immerse slides in deionized water.
12. Counterstain sections in hematoxylin.
13. Wash sections in deionized water two times for 5 minutes each.
14. Dehydrate sections.
15. Mount coverslips.

*The above information is only intended as a guide. The researcher should determine what protocol best meets their needs. Please follow safe laboratory procedures.

Immunocytochemistry/ Immunofluorescence Protocol for beta-Catenin Antibody (NBP1-54467)

Immunocytochemistry Protocol

Culture cells to appropriate density in 35 mm culture dishes or 6-well plates.

1. Remove culture medium and add 10% formalin to the dish. Fix at room temperature for 30 minutes.
2. Remove the formalin and add ice cold methanol. Incubate for 5-10 minutes.
3. Remove methanol and add washing solution (i.e. PBS). Be sure to not let the specimen dry out. Wash three times for 10 minutes.
4. To block nonspecific antibody binding incubate in 10% normal goat serum from 1 hour to overnight at room temperature.
5. Add primary antibody at appropriate dilution and incubate at room temperature from 2 hours to overnight at room temperature.
6. Remove primary antibody and replace with washing solution. Wash three times for 10 minutes.
7. Add secondary antibody at appropriate dilution. Incubate for 1 hour at room temperature.
8. Remove antibody and replace with wash solution, then wash for 10 minutes. Add Hoechst 33258 to wash solution at 1:25,000 and incubate for 10 minutes. Wash a third time for 10 minutes.
9. Cells can be viewed directly after washing. The plates can also be stored in PBS containing Azide covered in Parafilm (TM). Cells can also be cover-slipped using Fluoromount, with appropriate sealing.

*The above information is only intended as a guide. The researcher should determine what protocol best meets their needs. Please follow safe laboratory procedures.



Flow (Intracellular) Protocol for beta-Catenin Antibody (NBP1-54467)

Protocol for Flow Cytometry Intracellular Staining

Sample Preparation.

1. Grow cells to 60-85% confluency. Flow cytometry requires between 2×10^5 and 1×10^6 cells for optimal performance.
2. If cells are adherent, harvest gently by washing once with staining buffer and then scraping. Avoid using trypsin as this can disrupt certain epitopes of interest. If enzymatic harvest is required, use Accutase, Collagenase, or TrypLE Express for a less damaging option.
3. Reserve 100 μ L for counting, then transfer cell volume into a 50 mL conical tube and centrifuge for 8 minutes at 400 RCF.
 - a. Count cells using a hemocytometer and a 1:1 trypan blue exclusion stain to determine cell viability before starting the flow protocol. If cells appear blue, do not proceed.
4. Re-suspend cells to a concentration of 1×10^6 cells/mL in staining buffer (NBP2-26247).
5. Aliquot out 1 mL samples in accordance with your experimental samples.

Tip: When cell surface and intracellular staining are required in the same sample, it is advisable that the cell surface staining be performed first since the fixation and permeabilization steps might reduce the availability of surface antigens.

Intracellular Staining.

Tip: When performing intracellular staining, it is important to use appropriate fixation and permeabilization reagents based upon the target and its subcellular location. Generally, our Intracellular Flow Assay Kit (NBP2-29450) is a good place to start as it contains an optimized combination of reagents for intracellular staining as well as an inhibitor of intracellular protein transport (necessary if staining secreted proteins). Certain targets may require more gentle or transient permeabilization protocols such as the commonly employed methanol or saponin-based methods.

Protocol for Cytoplasmic Targets:

Optional: Perform cell surface staining as described in the previous section.

1. Fix the cells by adding 100 μ L fixation solution (such as 4% PFA) to each sample for 10-15 minutes.
2. Permeabilize cells by adding 100 μ L of a permeabilization buffer to every 1×10^6 cells present in the sample. Mix well and incubate at room temperature for 15 minutes.
 - a. For cytoplasmic targets, use a gentle permeabilization solution such as 1X PBS + 0.5% Saponin or 1X PBS + 0.5% Tween-20.
 - b. To maintain the permeabilized state throughout your experiment, use staining buffer + 0.1% of the permeabilization reagent (i.e. 0.1% Tween-20 or 0.1% Saponin).
3. Following the 15 minute incubation, add 2 mL of the staining buffer + 0.1% permeabilizer to each sample.
4. Centrifuge for 5 minutes at 400 RCF.
5. Discard supernatant and re-suspend in 1 mL of staining buffer + 0.1% permeabilizer.
6. Stain each sample at 1 μ L/ 1×10^6 cells of primary antibody or 1-3 μ L/ 1×10^6 cells for directly conjugated antibodies. Mix well and incubate at room temperature for 30 minutes- 1 hour. Gently mix samples every 10-15 minutes.
7. Following the primary/conjugate incubation, add 2 mL/sample of staining buffer +0.1% permeabilizer and centrifuge for 5 minutes at 400 RCF.
8. Remove supernatant and re-suspend each sample in 2 mL staining buffer + 0.1% permeabilizer, repeat wash for 5 minutes at 400 RCF.
9. If using a directly conjugated antibody, after the second wash, re-suspend cell pellet to a final volume of 500 μ L per sample and proceed with flow analysis.





Novus Biologicals USA

10730 E. Briarwood Avenue
Centennial, CO 80112
USA
Phone: 303.730.1950
Toll Free: 1.888.506.6887
Fax: 303.730.1966
nb-customerservice@bio-techne.com

Bio-Techne Canada

21 Canmotor Ave
Toronto, ON M8Z 4E6
Canada
Phone: 905.827.6400
Toll Free: 855.668.8722
Fax: 905.827.6402
canada.inquires@bio-techne.com

Bio-Techne Ltd

19 Barton Lane
Abingdon Science Park
Abingdon, OX14 3NB, United Kingdom
Phone: (44) (0) 1235 529449
Free Phone: 0800 37 34 15
Fax: (44) (0) 1235 533420
info.EMEA@bio-techne.com

General Contact Information

www.novusbio.com
Technical Support: nb-technical@bio-techne.com
Orders: nb-customerservice@bio-techne.com
General: novus@novusbio.com

Products Related to NBP1-54467

NBP2-33376H	Blue Marker Antibody (6F4-F6) [HRP]
HAF007	Goat anti-Mouse IgG Secondary Antibody [HRP]
NB7539	Goat anti-Mouse IgG (H+L) Secondary Antibody [HRP]
NBP1-97005-0.5mg	Mouse IgG1 Isotype Control (MG1)

Limitations

This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

For more information on our 100% guarantee, please visit www.novusbio.com/guarantee

Earn gift cards/discounts by submitting a review: www.novusbio.com/reviews/submit/NBP1-54467

Earn gift cards/discounts by submitting a publication using this product:
www.novusbio.com/publications

