Product Datasheet

Nrf2 Antibody - BSA Free

NBP1-32822

Unit Size: 0.1 ml

Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.

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Reviews: 4 Publications: 77

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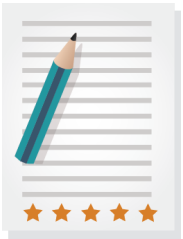
Updated 9/25/2025 v.20.1



NBP1-32822

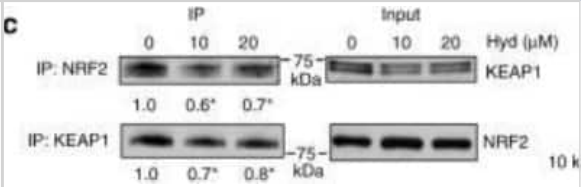
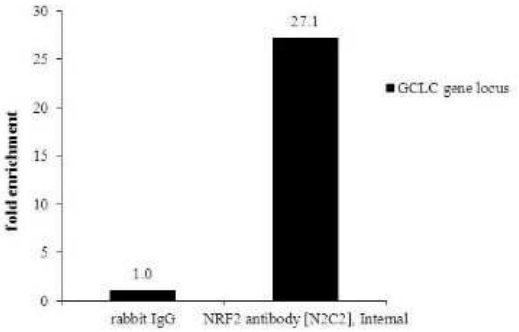
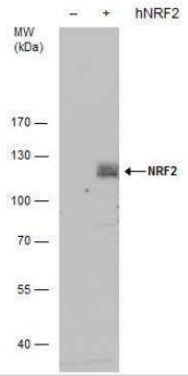
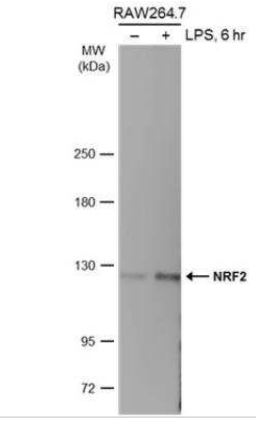
Nrf2 Antibody - BSA Free

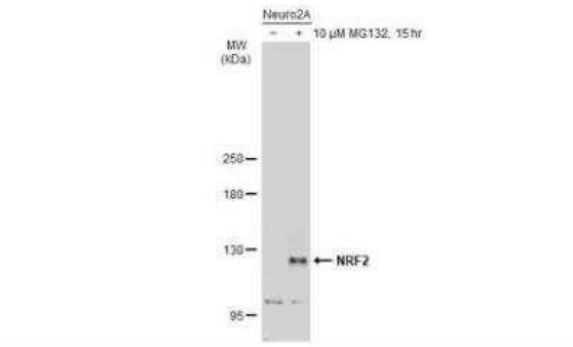
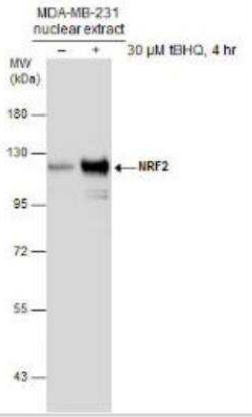
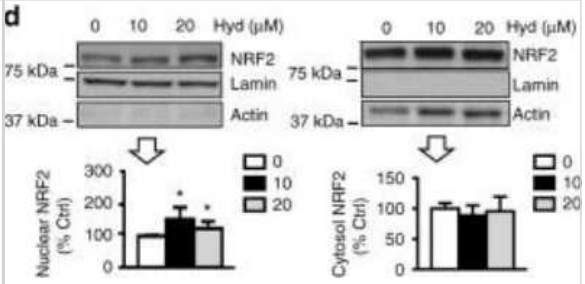
Product Information	
Unit Size	0.1 ml
Concentration	Concentrations vary lot to lot. See vial label for concentration. If unlisted please contact technical services.
Storage	Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.
Clonality	Polyclonal
Preservative	0.025% Proclin 300
Isotype	IgG
Purity	Antigen Affinity-purified
Buffer	PBS, 1% BSA, 20% Glycerol
Target Molecular Weight	68 kDa

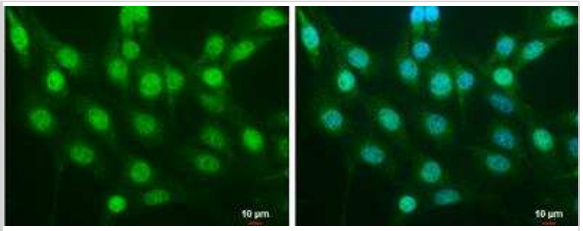
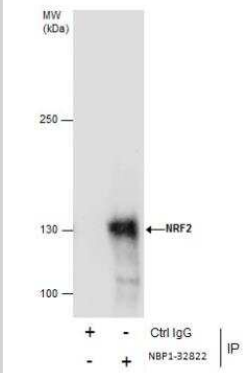
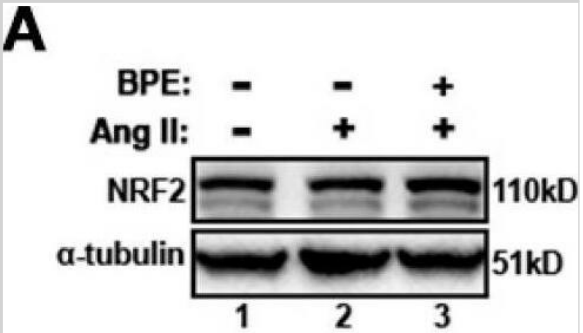
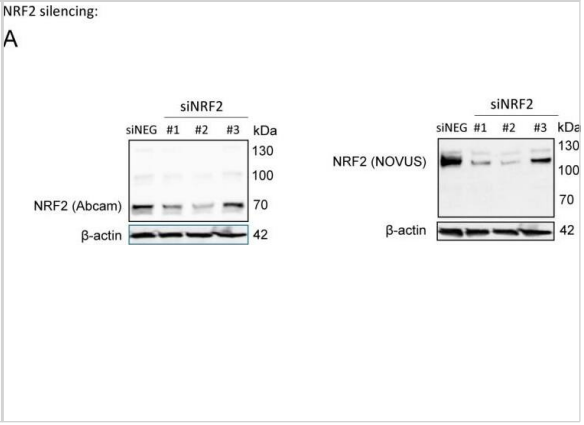
Product Description	
Description	Novus Biologicals Rabbit Nrf2 Antibody - BSA Free (NBP1-32822) is a polyclonal antibody validated for use in IHC, WB, Flow, ICC/IF, Simple Western, IP and ChIP. Anti-Nrf2 Antibody: Cited in 72 publications. All Novus Biologicals antibodies are covered by our 100% guarantee.
Host	Rabbit
Gene ID	4780
Gene Symbol	NFE2L2
Species	Human, Mouse, Rat, Alligator, Avian, Plant, Zebrafish
Immunogen	Recombinant protein encompassing a sequence within the center region of human NRF2. The exact sequence is proprietary.

Product Application Details	
Applications	Western Blot, Simple Western, Immunohistochemistry-Paraffin, Flow Cytometry, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunoprecipitation, Chromatin Immunoprecipitation (ChIP), Knockdown Validated
Recommended Dilutions	Western Blot 1:500-1:3000, Simple Western, Flow Cytometry Assay dependent, Immunohistochemistry 1:100-1:1000, Immunocytochemistry/ Immunofluorescence 1:100-1:1000, Immunoprecipitation 1:100-1:500, Immunohistochemistry-Paraffin 1:100-1:1000, Chromatin Immunoprecipitation (ChIP) Assay dependent, Knockdown Validated
Application Notes	See Simple Western Antibody Database for Simple Western validation: Tested in Hep-G2, separated by Size, apparent MW was 108 kDa

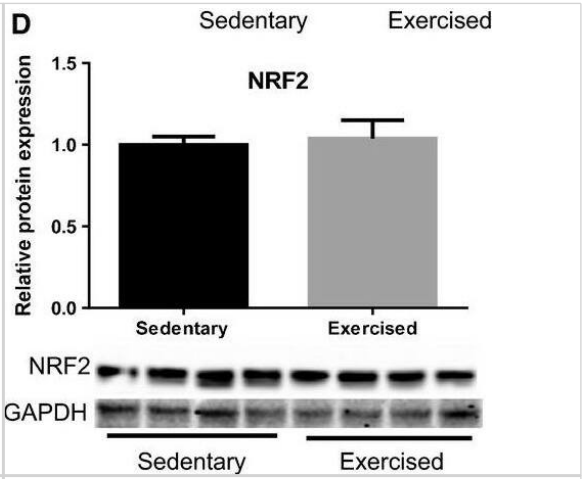
Images

Hydralazine enhances NRF2 signaling in SH-SY5Y cells. c Hydralazine reduced the interaction between NRF2 and KEAP1. Interactions were measured by reciprocal Co-IPs followed by western blot analysis. *p < 0.05, two-tailed Student's t test, n = 3, mean +/- SD. Image collected and cropped by CiteAb from the following publication (https://www.nature.com/articles/s41467-017-02394-3), licensed under a CC-BY license.	
ChIP was performed with HepG2 chromatin extract and 5 ug of either normal rabbit IgG or anti-NRF2 antibody. The precipitated DNA was detected by PCR with primer set targeting to GCLC gene locus.	
Non-transfected (-) and NRF2-transfected (+, including 3xFlag-tag) 293T whole cell extracts (30ug) were separated by 5% SDS-PAGE, and the membrane was blotted with NRF2 antibody diluted by 1:1000.	
Untreated (-) and treated (+) RAW264.7 whole cell extracts (30 ug) were separated by 5% SDS-PAGE, and the membrane was blotted with NRF2 antibody [N2C2], Internal diluted at 1:500. The HRP-conjugated anti-rabbit IgG antibody (NBP2-19301) was used to detect the primary antibody.	

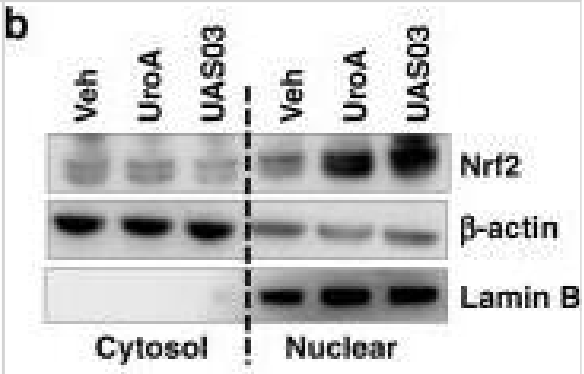
Untreated (-) and treated (+) Neuro2A whole cell extracts (30 ug) were separated by 5% SDS-PAGE, and the membrane was blotted with NRF2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:500. The HRP-conjugated anti-rabbit IgG antibody (NBP2-19301) was used to detect the primary antibody.	
Untreated (-) and treated (+) HepG2 whole cell extracts (30 ug) were separated by 5% SDS-PAGE, and the membrane was blotted with NRF2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:500. The HRP-conjugated anti-rabbit IgG antibody (NBP2-19301) was used to detect the primary antibody.	
Untreated (-) and treated (+) MDA-MB-231 nuclear extracts (30 ug) were separated by 7.5% SDS-PAGE, and the membrane was blotted with NRF2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:1000.	
Nrf2-Antibody-Western-Blot-NBP1-32822-img0047.jpg	

NIH/3T3 cells were fixed in 4% paraformaldehyde at RT for 15 min. Green: NRF2 protein stained by NRF2 antibody [N2C2], Internal diluted at 1:500. Blue: Hoechst 33342 staining. Scale bar = 10 um.	
Immunoprecipitation of NRF2 protein from HepG2 whole cell extracts using 5 ug of NRF2 antibody [N2C2], Internal Western blot analysis was performed using NRF2 antibody [N2C2], Internal.. EasyBlot anti-Rabbit IgG was used as a secondary reagent.	
Western Blot: Nrf2 Antibody [NBP1-32822] - Blueberry polyphenol extract (BPE) increases the expression of NRF2 & HO-1 while reducing NF-κB p65 phosphorylation in angiotensin (Ang) II-treated human aortic endothelial cells (HAECs). HAECs were treated with 200 µg/mL of BPE for 1 h then treated with 200 nM of Ang II for 12 h. Protein expression of NRF2 (A,B), HO-1 (C,D), & NF-κB p65 (E,F) were determined by Western blot. Quantification was performed using Image Lab (Bio-Rad Laboratories, Inc.). Data are expressed as mean ± SD from nine (HO-1), & three (NRF2 & NF-κB) independent experiments. Values that do not share the same letter are significantly different from each other ($p \leq 0.05$). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/35453301), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	
Western Blot: Nrf2 Antibody [NBP1-32822] - NRF2 antibody validation. (A) NRF2 expression was silenced in MCF7 cells by transiently transfecting NRF2-specific siRNAs or a negative control siRNA for 48 h, then NRF2 protein expression was determined using two different antibodies (Abcam: ab31163; NOVUS: NBP1-32822). (B–D) 4T1 cells were treated with NRF2 activators, RA839 or tBHQ, or MG-132, a proteasome inhibitor, in the concentrations indicated for 48 h, then NRF2 protein expression was determined by western blotting using two different antibodies (Abcam: ab31163; NOVUS: NBP1-32822). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/31461945), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	

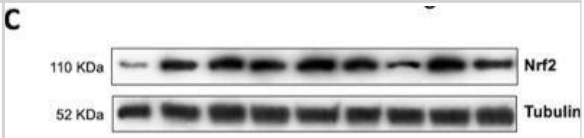
Western Blot: Nrf2 Antibody [NBP1-32822] - Maternal exercise during pregnancy on mitochondrial biogenesis in the fetal hearts. (A) Levels of relative mRNA expression measured by qRT□PCR. n = 9–12/group. Maternal exercise during pregnancy did not alter levels of mRNA in Ppargc1a & Tfam, while it significantly upregulated the levels of mRNA in Nrf1 & Nrf2. (B–D) Densitometric analyses of protein expression levels relative to the sedentary group with representative images of western blots were shown. No significant differences in PGC□1α, NRF1, & NRF2 (P > 0.05). n = 5–6/group. * P < 0.05, significantly different from the sedentary group. Black bar: fetal hearts from sedentary dams; gray bar: fetal hearts from exercised dams. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/28292876), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



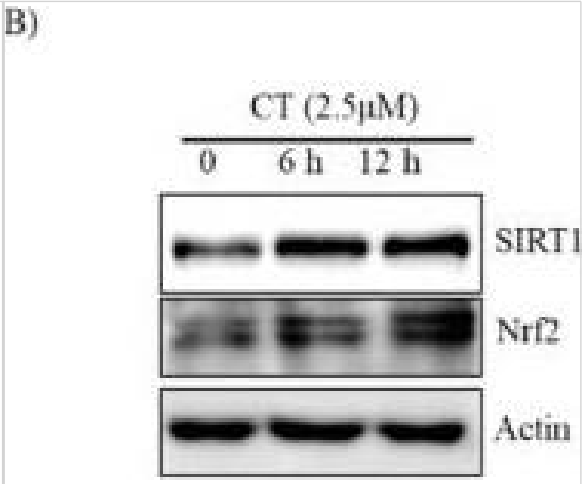
Western Blot: Nrf2 Antibody [NBP1-32822] - Nrf2 is required for UroA/UAS03 mediated upregulation of tight junction proteins. a Nrf2 levels were determined by immunoblots in HT29 cells treated with vehicle/UroA/UAS03 (50 μM) for 24 h. b Nrf2 expression in cytosolic & nuclear fractions of HT29 cells treated with Veh/UroA/UAS03 (50 μM) for 6 h. c Immunofluorescence confocal images of HT29 cells treated with vehicle/UroA/UAS03 (50 μM) for 6 h. The cells were stained with anti-Nrf2 antibody & DAPI. Relative green fluorescence (n = ~20 cells) intensity was measured. Scale bars indicate 25 μm. d Expression of Cldn4 & NQO1 in colon explants from WT, Nrf2−/−, & AhR−/− mice treated with vehicle/UroA/UAS03 (50 μM) for 24 h. Immunoblots were quantified using Image J software. e mRNA levels of Cldn4, Nrf2, & HO1 from colon explant cultures was measured by real-time PCR using SyBr green method. f C57BL/6, Nrf2−/−, & AhR−/− mice (n = 3) treated orally daily with veh or UroA/UAS03 (20 mg/kg) for 1 week. Cldn4 & NQO1 protein levels in colons were measured by immunoblots & quantified by Image J software. All in vitro studies were performed in triplicates. The immunoblots of colon explants & colon tissues were quantified from at least 6 independent runs. The levels of proteins were normalized to β-actin & Wild type vehicle treatment was set to 1 & calculated the fold changes. Statistics performed using unpaired t-test using Graphpad Prism software. Error bars, ±SEM; *p < 0.05; **p < 0.01; ***p < 0.001. Source Data are provided as a Source Data File Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/30626868), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



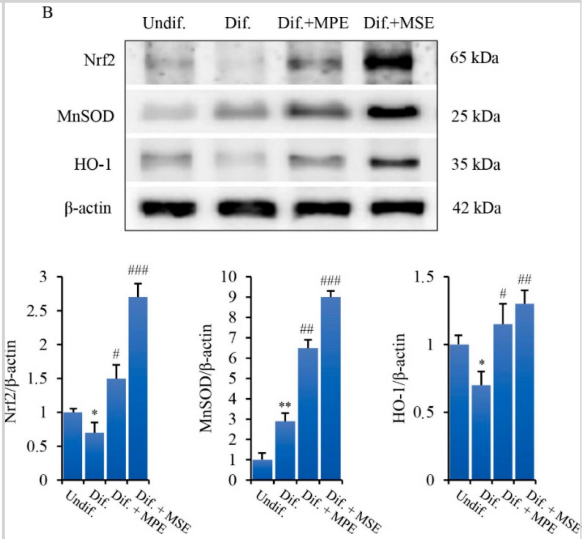
Western Blot: Nrf2 Antibody [NBP1-32822] - Modulation of Nrf2 & Keap1 mRNA & protein levels by compounds 1–6, curcumin (CURC), & dimethyl fumarate (DMF). (A–B) RNA from total cellular extracts of SH-SY5Y cells treated for 24 hours with 5 μM compounds or 20 μM DMF were analyzed for Nrf2 (A) & Keap1 (B) mRNA expression by RT-qPCR. GAPDH was used as housekeeping gene. Results are shown as mean ± SEM; no statistically significant data with Dunnett's multiple comparison test (A, n = 3, F ratio = 1.249; B, n = 3, F ratio = 1.671). (C–D) Cellular extracts of SH-SY5Y cells treated for 24 hours with compounds at 5 μM or 20 μM DMF were analyzed for Nrf2 (C) & Keap1 (D) protein levels by Western blot. Anti-tubulin was used as protein loading control. Results are shown as ratio (% of CTR) ± SEM; **p < 0.01, versus CTR; Dunnett's multiple comparison test (C, n ≥ 5, F ratio = 3.981; D, n = 3, F ratio = 0.4049). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32047434), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



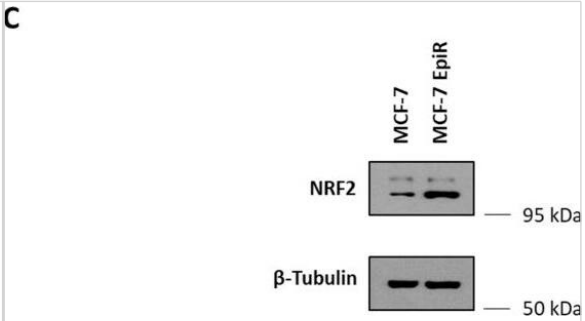
Western Blot: Nrf2 Antibody [NBP1-32822] - CT activated AMPK/SIRT1 signaling. (A,B) HepG2 cells were treated with 2.5 μM CT for indicated times. Western blot analysis of phosphorylated AMPK, ACC, SIRT1, & Nrf2. (C) C57BL/6 mice were pair-fed either control or ethanol-containing diet with or without CT (20 or 40 mg/kg) for four weeks. Western blot analysis of phosphorylated AMPK, SIRT1. CYP2E1, & Nrf2. (D) HepG2 cells were incubated with 50 mM ethanol & treated with CT (2.5 or 5 μM) for 24 h. Western blot analysis of phosphorylated AMPK, SIRT1, CYP2E1, & Nrf2. (E) AML-12 cells were incubated with 50 mM ethanol & treated with CT (2.5 or 5 μM) for 24 h. Western blot analysis of phosphorylated AMPK & SIRT1. The images are representative (F) HepG2 cells were pretreated with CT (2.5 μM) for 3 h or with compound C (comp C) (10 μM) for 6 h, followed by ethanol (100 μM) treatment. Measurement of intracellular TG levels. Data are shown as mean ± SD of three independent experiments. #p < 0.05 vs. untreated control, ** p < 0.01 vs. ethanol-treated group. §§p < 0.01 vs. ethanol & CT-treated group. Densitometric analysis of western blots are given in Supplementary Figures S2 & S3A–G. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/31906014), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



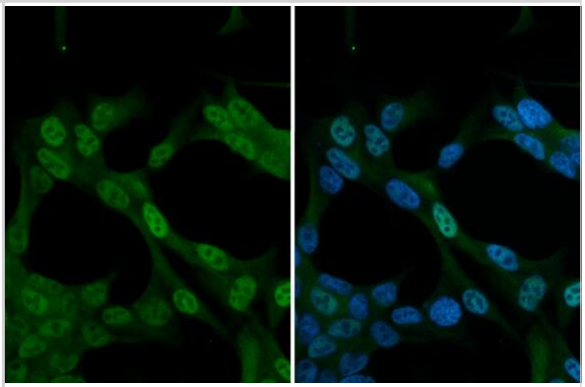
Western Blot: Nrf2 Antibody [NBP1-32822] - MPE & MSE exert anti-oxidant effects in 3T3-L1 adipocytes. 3T3-L1 cells were treated with pro-differentiative agents for 8 days in the presence or absence of 100 μg/mL MPE or MSE, as reported in Methods. (A) Intracellular ROS were detected using the redox-sensitive fluorochrome H2-DCFDA. After differentiation, the medium was replaced with 10 μM H2DCFDA solution & the incubation was protracted for 30 min at 37 °C. The oxidation of the fluorochrome generates green fluorescence, which was visualized by a Leica microscope equipped with a DC300F camera using a FITC filter. Representative micrographs of fluorescence microscopy were taken at 200× magnification. (B). Western blotting analysis of Nrf2, MnSOD & HO-1 in 3T3-L1 cells differentiated without or with 100 μg/mL MPE or MSE. Equal loading of proteins was verified by immunoblotting for β-actin & showed values were assigned in relation to undifferentiated cells (Undif.). The bar graphs represent the mean of three independent experiments ± SD. * p < 0.05, ** p < 0.01 with respect to the undifferentiated 3T3-L1 cells, # p < 0.05, ## p < 0.01 & ### p < 0.001 with respect to the differentiated untreated 3T3-L1 cells (Dif.). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/35204243), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



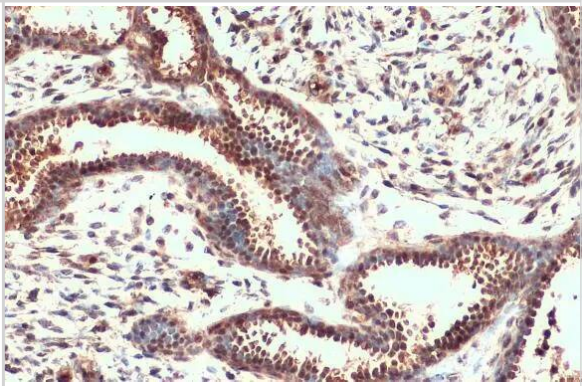
Western Blot: Nrf2 Antibody [NBP1-32822] - NRF2 modulates epirubicin resistance in breast cancer cells. (A) MCF-7 & MCF-7 EpiR cells were treated for 24 h with epirubicin at 1 μM. MCF-7 & MCF-7 EpiR cells were stained with CellROX Deep Red reagent & analyzed for ROS levels by flow cytometry. Data were analyzed by FlowJo software. The mean fluorescence values were presented as relative ROS level compared to untreated cells (0 μM). Data presented as mean ± SD. Student's t-test was used to compare the means: ** $p < 0.01$; *** $p < 0.001$; n.s. nonsignificant. (N = 4) (B) Knockdown of NRF2 was achieved by transfecting 4 specific siRNA against NRF2 (siNRF2, 150pmol) to MCF-7 EpiR cells in a 6-well plate. Non-targeting siRNAs were used as control (NSC). At 24 h post-transfection, these cells were seeded in 6-well plates & treated with increasing doses of epirubicin for 14 days. Their sensitivity to epirubicin was assessed by clonogenic assay. Their clonogenicity in response to epirubicin was analyzed by two-way ANOVA & found to be significantly different (** $p < 0.01$) from one another. (N = 3) (C) Expression of NRF2 in MCF-7 cells & MCF-7 EpiR cells was detected by Western blot. β-Tubulin served as the loading control. (N = 3). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32110852), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

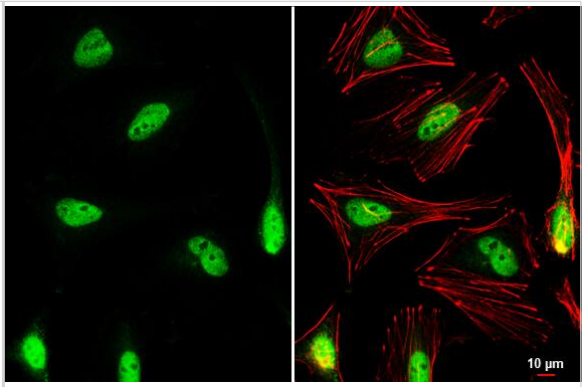
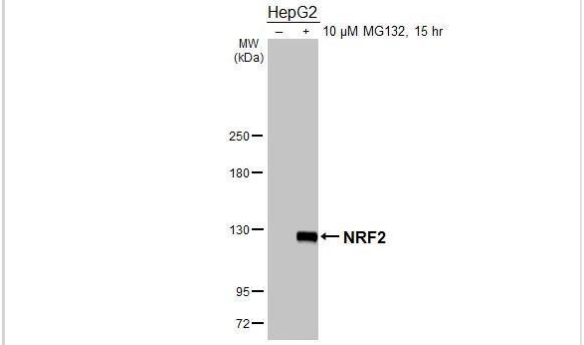
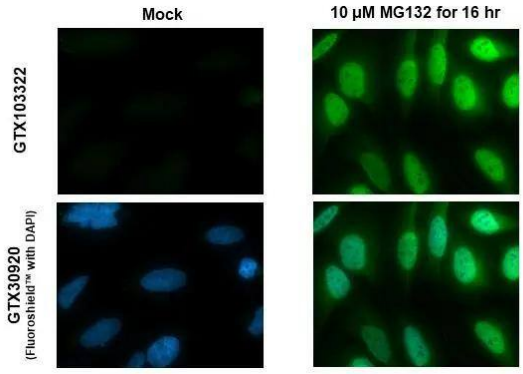
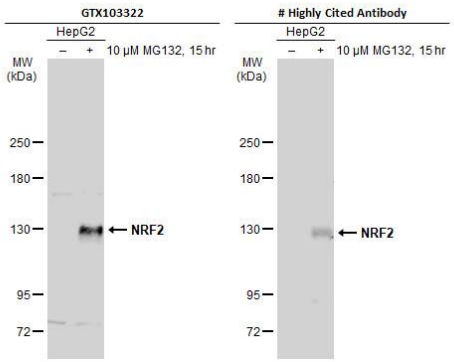


Immunocytochemistry/ Immunofluorescence: Nrf2 Antibody [NBP1-32822] - Nrf2 antibody [N2C2], Internal detects Nrf2 protein at nucleus by immunofluorescent analysis.Sample: Neuro2A cells were fixed in 4% paraformaldehyde at RT for 15 min.Green: Nrf2 stained by Nrf2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:1000.



Immunohistochemistry-Paraffin: Nrf2 Antibody [NBP1-32822] - Nrf2 antibody [N2C2], Internal detects Nrf2 protein at cytoplasm and nucleus by immunohistochemical analysis.Sample: Paraffin-embedded human breast carcinoma.Nrf2 stained by Nrf2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:500.Antigen Retrieval: Citrate buffer, pH 6.0, 15 min



Immunocytochemistry/ Immunofluorescence: Nrf2 Antibody [NBP1-32822] - Nrf2 antibody [N2C2], Internal detects Nrf2 protein at nucleus by immunofluorescent analysis.Sample: HeLa cells were fixed in 4% paraformaldehyde at RT for 15 min.Green: Nrf2 stained by Nrf2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:1000.Red: phalloidin, a cytoskeleton marker, diluted at 1:200.Scale bar= 10 um.	
Untreated (-) and treated (+) HepG2 whole cell extracts (30 ug) were separated by 5% SDS-PAGE, and the membrane was blotted with Nrf2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:2000. The HRP-conjugated anti-rabbit IgG antibody was used to detect the primary antibody.	
NRF2 antibody [N2C2], Internal detects NRF2 protein at nucleus by immunofluorescent analysis.Sample: Mock and treated HeLa cells were fixed in 4% paraformaldehyde at RT for 15 min.Green: NRF2 stained by NRF2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:500.Blue: Fluoroshield with DAPI .	
Untreated (-) and treated (+) HepG2 whole cell extracts (30 ug) were separated by 5% SDS-PAGE, and the membranes were blotted with NRF2 antibody [N2C2], Internal (NBP1-32822) diluted at 1:500 and competitor's antibody diluted at 1:500. The HRP-conjugated anti-rabbit IgG antibody was used to detect the primary antibody. *The competitor is not affiliated with Novus and does not endorse this product.	

Publications

Tamburini B, Liberto D, Pratelli G et al. Extra Virgin Olive Oil Polyphenol-Enriched Extracts Exert Antioxidant and Anti-Inflammatory Effects on Peripheral Blood Mononuclear Cells from Rheumatoid Arthritis Patients. Antioxidants (Basel, Switzerland) 2025-01-31 [PMID: 40002358]

Sipos A, Kerekes □, Szeőcs D et al. Ursodeoxycholic acid prompts glycolytic dominance, reductive stress and epithelial-to-mesenchymal transition in ovarian cancer cells through NRF2 activation Cell Death Discovery 2025-04-03 [PMID: 40175359]

Di Lollo V, Canciello A, Peserico A et al. Unveiling the immunomodulatory shift: Epithelial-mesenchymal transition Alters immune mechanisms of amniotic epithelial cells iScience 2023-09-15 [PMID: 37680464]

Seok JH, Kim DH, Kim HJ et al. Epigallocatechin-3-gallate suppresses hemin-aggravated colon carcinogenesis through Nrf2-inhibited mitochondrial reactive oxygen species accumulation Journal of Veterinary Science 2022-08-18 [PMID: 36174978]

Hussein MM, Sayed RKA, Mokhtar DM. Structural and immunohistochemical analysis of the cellular compositions of the liver of molly fish (Poecilia sphenops), focusing on its immune role Zoological Letters 2023-01-05 [PMID: 36604695]

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