



PRODUCT INFORMATION & MANUAL

Mouse HGF Valukine™ ELISA

VAL617

For the quantitative determination of natural and recombinant mouse Hepatocyte Growth Factor HGF concentrations

For research use only.
Not for diagnostic or therapeutic procedures.

Bio-Techne China Co. Ltd

P: +86 (21) 52380373 P: 8009881270 F: +86 (21) 52381001
info.cn@bio-techne.com

Please refer to the kit label for expiry date.
Novus kits are guaranteed for 3 months from date of receipt

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I. BACKGROUND

HGF, also known as Scatter Factor and Hepatopoietin A, is a pleiotropic protein in the Plasminogen subfamily of S1 peptidases. It is a multidomain molecule that includes an N-terminal PAN/APPLE-like domain, four Kringle domains, and a serine proteinase-like domain that has no detectable protease activity (1 - 5). Mouse HGF is secreted as an inactive 728 amino acid (aa) single chain propeptide. It is cleaved after the fourth Kringle domain by a serine protease to form bioactive disulfide-linked HGF with a 60 kDa alpha and 30 kDa beta chain. Alternate splicing generates an isoform that lacks the peptidase and the second, third, and fourth Kringle domains. Mouse HGF shares 91% - 95% aa sequence identity with bovine, canine, feline, human, and rat HGF. HGF binds heparan-sulfate proteoglycans and the widely expressed receptor tyrosine kinase, HGF R/c-MET (6, 7). HGF - dependent c-MET activation is implicated in the development of many human cancers (8). HGF regulates epithelial morphogenesis by inducing cell scattering and branching tubulogenesis (9, 10). HGF induces the up - regulation of integrin alpha 2 beta 1 in epithelial cells by a selective increase in alpha 2 gene transcription (11). This integrin serves as a collagen I receptor, and its blockade disrupts epithelial cell branching tubulogenesis (11, 12). HGF can also alter epithelium morphology by the induction of nectin - 1 alpha ectodomain shedding, an adhesion protein component of adherens junctions (13). In the thyroid, HGF induces the proliferation, motility, and loss of differentiation markers of thyrocytes and inhibits TSH-stimulated iodine uptake (14). HGF promotes the motility of cardiac stem cells in damaged myocardium (15).

II. OVERVIEW

A. PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for mouse HGF has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any HGF present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated detection antibody specific for mouse HGF are pipetted into the wells. After washing away any unbound substances, streptavidin-HRP are pipetted into the wells. Following a wash to remove any unbound reagent, a substrate solution is added to the wells and color develops in proportion to the amount of HGF bound in the initial step. The color development is stopped, and the intensity of the color is measured.

B. LIMITATIONS OF THE PROCEDURE

- **FOR RESEARCH USE ONLY NOT FOR USE IN DIAGNOSTIC PROCEDURES.**
- This kit is suitable for cell culture supernate and mouse serum.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples generate values higher than the highest standard, dilute the samples with Calibrator Diluent and repeat the assay.
- Any variation in operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

III. ADVANTAGES

A. PRECISION

Intra-assay Precision (Precision within an assay)

Three samples were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

Three samples were tested in twenty separate assays to assess inter-assay precision.

Sample	Intra-assay Precision			Inter-assay Precision		
	1	2	3	1	2	3
Mean (pg/mL)	264.2	1071.4	3722.6	269.1	1064.8	3758.2
Standard Deviation	12.7	21.2	60.7	16.1	25.5	117.6
CV%	4.8	2.0	1.6	6.0	2.4	3.1

B. RECOVERY

The recovery of mouse HGF spiked to different levels throughout the range of the assay in cell culture media was evaluated. The recovery ranged from 102.1-108% with an average of 104.9%.

The recovery of mouse HGF spiked to different levels throughout the range of the assay in mouse serum was evaluated. The recovery ranged from 88-107.2% with an average of 93.2%.

C. SENSITIVITY

The minimum detectable dose (MDD) of mouse HGF is typically less than 5.18 pg/mL.

The MDD was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

D. CALIBRATION

This immunoassay is calibrated against highly purified *NS0*-expressed recombinant mouse HGF produced at R&D Systems.

E. LINEARITY

To assess the linearity of the assay, different samples were containing or spiked with high concentrations of mouse HGF and diluted with Calibrator Diluent (1×) to produce samples with values within the dynamic range of the assay.

Dilution	Average % of Expected	Range (%)
1:2	100.0	89 - 107
1:4	99.5	94.9 - 114.1
1:8	103.8	95.3 - 119.5
1:16	100.9	85.2 - 110

F. SAMPLE VALUES

Serum - Five mouse serum samples were evaluated for the presence of mouse HGF in this assay. All samples measured ranged from 2158 to 2966 pg/mL with an average of 2586 pg/mL.

G. SPECIFICITY

The following factors prepared at 100 ng/mL were assayed and exhibited no cross-reactivity or interference.

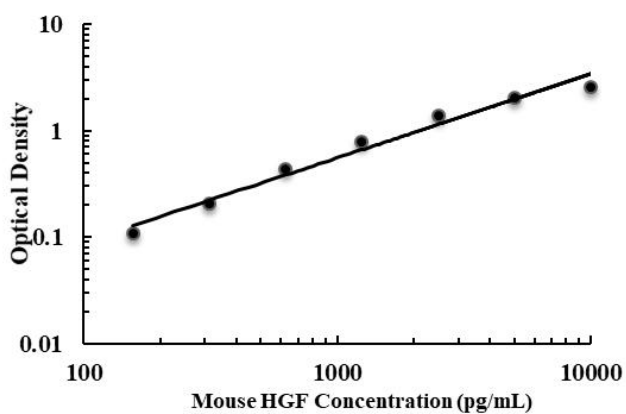
Recombinant mouse	Recombinant human
HGF Activator	HGF
	HGF Activator
	HGF R/Fc Chimera
	MSP
	MSP β chain

Recombinant mouse HGF R/Fc Chimera does not cross-react in this assay but does interfere at concentrations > 3.12 ng/mL.

IV. EXPERIMENT

EXAMPLE STANDARD

The standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.



pg/mL	OD	Average	Corrected
0	0.057 0.057	0.057	-
156.25	0.159 0.172	0.166	0.109
312.5	0.263 0.258	0.261	0.204
625	0.486 0.488	0.487	0.430
1250	0.845 0.839	0.842	0.785
2500	1.444 1.449	1.447	1.390
5000	2.073 2.159	2.116	2.059
10000	2.638 2.673	2.656	2.599

V. KIT COMPONENTS AND STORAGE

A. MATERIALS PROVIDED

Parts	Description	Size
Mouse HGF Microplate	96 well polystyrene microplate (12 strips of 8 wells) coated with a goat antibody against mouse HGF.	1 plate
Mouse HGF Standard	Recombinant mouse HGF in a buffered protein base; lyophilized. Refer to the vial label for reconstitution volume.	2 vials
Mouse HGF Detection Antibody	Biotinylated HGF antibody, lyophilized. Refer to the vial label for reconstitution volume.	1 vial
Calibrator Diluent (4×)	Concentrated buffered diluent used to dilute standard and samples.	1 vial
Streptavidin-HRP A (200×)	200× Streptavidin conjugated to horseradish peroxidase.	1 vial
Reagent Diluent (10×)	A 10× concentrated buffered protein base used to dilute Detection Antibody and HRP.	1 vial
Wash Buffer Concentrate (25×)	A 25× concentrated solution of buffered surfactant with preservatives.	1 vial
Color Reagent A	Stabilized hydrogen peroxide.	1 vial
Color Reagent B	Stabilized chromogen (tetramethylbenzidine).	1 vial
Stop Solution	2 N sulfuric acid.	1 vial
Plate Sealers	Adhesive strip.	3 strips

B. STORAGE

Unopened Kit	Store at 2-8 °C. Do not use past kit expiration date.	
Opened/ Reconstituted Reagents	Streptavidin-HRP A	May be stored for up to 1 month at 2-8 °C.*
	Diluted Wash Solution	
	Unmixed Color Reagent A	
	Unmixed Color Reagent B	
	Stop Solution	
	Standard	Prepare fresh for each assay.
	Detection Antibody	Aliquot and store for up to 1 month at -20 °C in a manual defrost freezer. *
	Reagent Diluent (10×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Reagent Diluent (1×). Prepare fresh for each assay.
	Calibrator Diluent (4×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Calibrator Diluent (1×). Prepare fresh for each assay.
Microplate Wells	Return unused wells to the foil pouch containing the desiccant pack, reseal along entire edge of zip-seal. May be stored for up to 1 month at 2-8 °C.*	

* Provided this is within the expiration date of the kit.

C. OTHER SUPPLIES REQUIRED

- ◆ Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- ◆ Pipettes and pipette tips.
- ◆ Deionized or distilled water.
- ◆ Squirt bottle, manifold dispenser, or automated microplate washer.
- ◆ 500 mL graduated cylinder.

D. PRECAUTION

- ◆ Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.
- ◆ Color Reagent B may cause skin, eye, and respiratory irritation. Avoid breathing fumes.
- ◆ The Stop Solution provided with this kit is an acid solution. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.

VI. PREPARATION

A. SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. Samples may require dilution with Calibrator Diluent (1 \times).

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

B. SAMPLE PREPARATION

Serum samples require a 2-fold dilution. A suggested 2-fold dilution is 100 μL of sample + 100 μL of Calibrator Diluent (1 \times).

C. REAGENT PREPARATION

Note: *Bring all reagents to room temperature before use.*

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate (25 \times) into deionized or distilled water to prepare 500 mL of Wash Buffer.

Calibrator Diluent (1 \times) - Use deionized or distilled water to prepare Calibrator Diluent (1 \times).

Reagent Diluent (1 \times) - Use deionized or distilled water to prepare Reagent Diluent (1 \times).

Detection Antibody- Reconstitution Volume refer to vial label with Reagent Diluent (1 \times). Aliquot and store if needed. Dilute stock solution in Calibrator Diluent (1 \times) to the working concentration of 200 ng/mL. Prepare at least 15 minutes prior to use.

Streptavidin-HRP A (1 \times) - Dilute to the working concentration specified on the vial label using Reagent Diluent (1 \times).

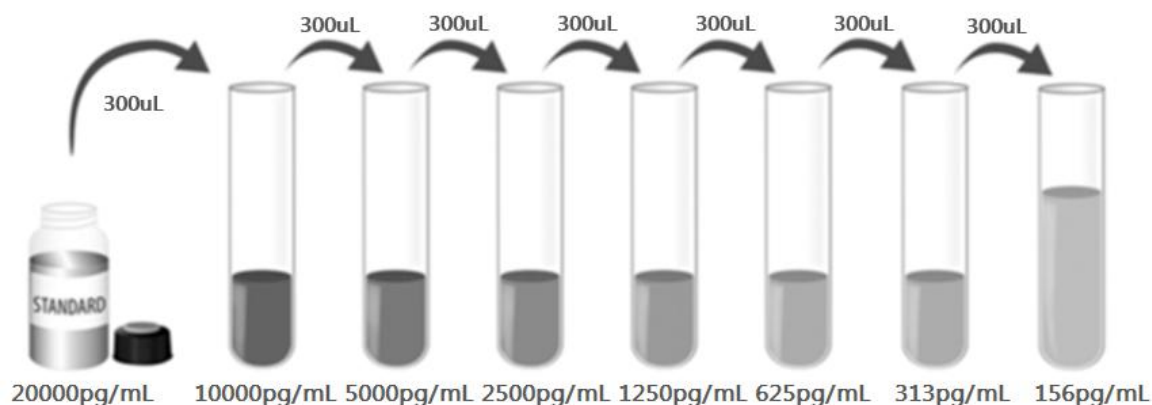
Substrate Solution - Color Reagent A and B should be mixed together in equal

volumes within 15 minutes of use. Protect from light. 100 μ L of the resultant mixture is required per well.

Mouse HGF Standard - Refer to the vial label for the reconstitution volume*. This reconstitution produces a stock solution of 20000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

*if you have any question, please seek help from our Technical Support.

Pipette 300 μ L of the appropriate Calibrator Diluent (1 \times) into each tube. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 10000 pg/mL standard serves as the high standard. The Calibrator Diluent (1 \times) serves as the zero standard (0 pg/mL).



D. TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- It is recommended that the samples be pipetted within 15 minutes.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Substrate Solution should remain colorless until added to the plate. Keep Substrate Solution protected from light. Substrate Solution should change from colorless to gradations of blue.
- Stop Solution should be added to the plate in the same order as the Substrate Solution. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.

VII. ASSAY PROCEDURE

Note: *Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.*

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 100 μ L of standard, or prepared sample per well. Cover with the adhesive strip provided. Incubate for 2 hours at room temperature. A plate layout is provided for a record of standards and samples assayed. (Samples may require dilution. See Sample Preparation section.)
4. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Solution (400 μ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 μ L of the Detection Antibody diluted in Calibrator Diluent, to each well. Cover with a new adhesive strip and incubate 2 hours at room temperature.
6. Repeat the aspiration/wash as in step 4.
7. Add 100 μ L of the working dilution of Streptavidin-HRP A to each well. Cover the plate and incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
8. Repeat the aspiration/wash as in step 4.
9. Add 100 μ L of Substrate Solution to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
10. Add 50 μ L of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
11. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections

in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

12. **CALCULATION OF RESULTS**

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density (O.D.). Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the HGF concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

VIII. REFERENCES

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PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

1																	
2																	
3																	
4																	
5																	
6																	
7																	
8																	
9																	
10																	
11																	
12																	
	A	B	C	D	E	F	G	H									



产品信息及操作手册

小鼠 HGF Valukine™ ELISA 试剂盒

目录号: **VAL617**

适用于定量检测天然和重组小鼠 HGF 的浓度

科研专用, 不可用于临床诊断

Bio-Techne China Co. Ltd

P: +86 (21) 52380373 P: 8009881270 F: +86 (21) 52381001

info.cn@bio-techne.com

有效期详见试剂盒包装标签

Novus 试剂盒确保在你收货日期 3 个月内有效

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I. 背景

HGF又称分散因子和肝细胞生成素A,是S1肽酶的纤溶酶原亚家族中的一种多效性蛋白。它是一个多结构域分子,包括一个N端的PAN/APPLE样结构域、四个Kringle结构域和一个未检测到蛋白酶活性的丝氨酸蛋白酶样结构域 (1-5)。小鼠HGF以一种无活性的728氨基酸(aa)单链前体形式分泌。它在第四个Kringle结构域后被丝氨酸蛋白酶切割,形成具有生物活性的二硫键连接的HGF,其 α 链为60 kDa, β 链为30 kDa。交替剪接产生一个缺乏肽酶异构体,和第二、第三和第四Kringle结构域的异构体。小鼠HGF与牛、犬、猫、人和大鼠HGF具有91%-95%的aa序列同源性。HGF结合硫酸乙酰肝素蛋白多糖和广泛表达的受体酪氨酸激酶,HGF R/c-MET (6, 7)。HGF依赖的c-MET激活与许多人类癌症的发展有关(8)。HGF通过诱导细胞散射和分支小管形成来调节上皮形态发生(9, 10)。HGF通过选择性增加 $\alpha 2$ 基因转录,诱导上皮细胞整合素 $\alpha 2\beta 1$ 上调(11)。该整合素作为I型胶原受体,其阻断会破坏上皮细胞分支小管形成(11, 12)。HGF还可以通过诱导黏附连接的粘附蛋白成分nectin-1 α 外结构域脱落来改变上皮形态(13)。在甲状腺中,HGF可诱导甲状腺细胞的增殖、运动和分化标志物的丢失,并抑制TSH刺激的碘摄入(14)。HGF促进受损心肌的心脏干细胞的运动(15)。

II. 概述

A. 检测原理

本实验采用双抗体夹心ELISA法。抗小鼠HGF抗体包被于微孔板上，样品和标准品中的小鼠HGF会与固定在板上的抗体结合，游离的成分被洗去；接着加入生物素化的抗小鼠HGF检测抗体进行孵育，洗涤去除未结合的物质后，加入链霉亲和素标记的辣根过氧化物酶（streptavidin-HRP）孵育。洗涤去除未结合的试剂后，加入底物溶液（显色剂）。溶液颜色与结合的目标蛋白成正比；加入终止液；用酶标仪测定吸光度。

B. 检测局限

- ◆ 仅供科研使用，不可用于体外诊断；
- ◆ 该试剂盒适用于细胞培养上清样本和小鼠血清样本；
- ◆ 请在试剂盒有效期内使用；
- ◆ 不同试剂盒及不同批号试剂盒的组分不能混用；
- ◆ 样本值若大于标准曲线的最高值，应将样本用标准品稀释剂（1×）稀释后重新检测；
- ◆ 检测结果的不同可由多种因素引起，包括实验人员的操作、移液器的使用方式、洗板技术、反应时间或温度、试剂盒的储存等。

III. 优势

A. 精确度

板内精确度（同一板内不同孔间的精确度）

已知浓度的三个样本，在同一板内分别检测20次，以确定板内精确度。

板间精确度（不同板之间的精确度）

已知浓度的三个样本，在不同板中分别检测20次，以确定板间精确度。

样本	板内精确度			板间精确度		
	1	2	3	1	2	3
平均值 (pg/mL)	264.2	1071.4	3722.6	269.1	1064.8	3758.2
标准差	12.7	21.2	60.7	16.1	25.5	117.6
CV%	4.8	2.0	1.6	6.0	2.4	3.1

B. 回收率

在细胞培养基样本中掺入检测范围内不同水平的小鼠HGF，测定其回收率。回收率范围在102.1-108%，平均回收率在104.9%。

在小鼠血清样本中掺入检测范围内不同水平的小鼠HGF，测定其回收率。回收率范围在88-107.2%，平均回收率在93.2%。

C. 灵敏度

小鼠HGF的最低可测剂量（MDD）一般小于5.18pg/mL。

MDD是根据20个重复的零标准品孔的吸光度值的平均值加两倍标准差计算得到的相对浓度。

D. 校正

此ELISA试剂盒经由R&D Systems生产的高纯度重组小鼠HGF蛋白所校正。

E. 线性

不同的样本中含有或掺入高浓度的小鼠HGF，然后用标准品稀释剂（1×）将样本稀释到检测范围内，测定其线性。

稀释倍数	平均值/期待值 (%)	范围 (%)
1:2	100.0	89 - 107
1:4	99.5	94.9 - 114.1
1:8	103.8	95.3 - 119.5
1:16	100.9	85.2 - 110

F. 样本预值

血清样本 - 使用本试剂盒检测了5份小鼠血清样本中HGF的水平。5份样本的检测值范围为2158 – 2966 pg/mL，平均值为2586 ng/mL。

G. 特异性

将以下因子配置成100ng/mL的浓度来检测，没有观察到明显的交叉反应或干扰。

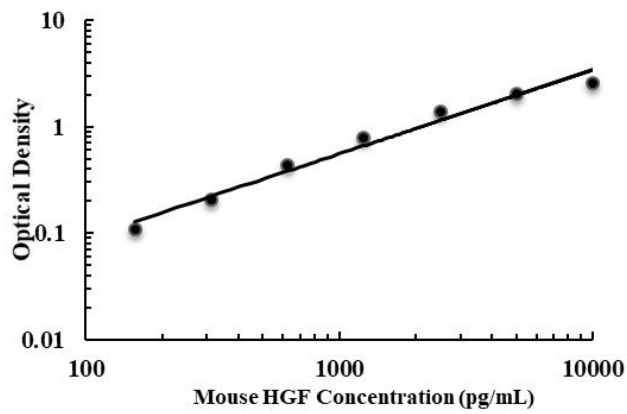
Recombinant mouse	Recombinant human
HGF Activator	HGF
	HGF Activator
	HGF R/Fc Chimera
	MSP
	MSP β chain

重组小鼠HGF R/Fc嵌合体在本实验中没有交叉反应，但在浓度>3.12ng/mL时有干扰作用。

IV. 实验

标准曲线实例

该标准曲线数据仅供参考，每次实验应绘制其对应的标准曲线。



pg/mL	OD	Average	Corrected
0	0.057 0.057	0.057	-
156.25	0.159 0.172	0.166	0.109
312.5	0.263 0.258	0.261	0.204
625	0.486 0.488	0.487	0.430
1250	0.845 0.839	0.842	0.785
2500	1.444 1.449	1.447	1.390
5000	2.073 2.159	2.116	2.059
10000	2.638 2.673	2.656	2.599

V. 试剂盒组成及储存

A. 试剂盒组成

组成	描述	规格
Mouse HGF Microplate	包被山羊抗小鼠 HGF 抗体的 96 孔聚苯乙烯板，8 孔×12 条	1 块板
Mouse HGF Standard	标准品（冻干粉），参考瓶身标签进行重溶	2 瓶
Mouse HGF Detection antibody	生物素化的 HGF 检测抗体，冻干粉，参考瓶身标签进行重溶	1 瓶
Streptavidin-HRP A (200×)	200×浓缩的链霉亲和素标记的 HRP	1 瓶
Reagent Diluent (10×)	浓缩的试剂稀释液（10×）	1 瓶
Calibrator Diluent (4×)	浓缩的标准品稀释剂（4×）	1 瓶
Wash Buffer Concentrate (25×)	浓缩洗涤缓冲液（25×）	1 瓶
Color Reagent A	显色液 A	1 瓶
Color Reagent B	显色液 B	1 瓶
Stop Solution	终止液	1 瓶
Plate Sealers	封板膜	3 张

B. 试剂盒储存

未开封试剂盒	2-8℃储存；请在试剂盒有效期内使用	
已打开，稀释 或重溶的试剂	链霉亲和素-HRP A	2-8℃储存，最多 30 天*
	洗涤缓冲液（1×）	
	显色剂 A	
	显色剂 B	
	终止液	
	标准品	使用时新鲜配制*
	检测抗体	分装， -20℃储存，最多 30 天*
	标准品稀释剂（4×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×标准品稀释剂
	试剂稀释液（10×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×试剂稀释液
包被的微孔板条	将未用的板条放回带有干燥剂的铝箔袋内，密封：2-8℃储存，最多 30 天*	

*必须在试剂盒有效期内

C. 实验所需自备试验器材

- ◆ 酶标仪（可测量450nm检测波长的吸收值及540nm或570nm校正波长的吸收值）
- ◆ 高精度加液器及一次性吸头
- ◆ 蒸馏水或去离子水
- ◆ 洗瓶（喷瓶）、多通道洗板器或自动洗板机
- ◆ 500mL量筒

D. 注意事项

- ◆ 试剂盒中的一些组分含有防腐剂，可能引起皮肤过敏反应，避免吸入。
- ◆ Color Reagent B可能引起皮肤、眼睛和呼吸道刺激，避免吸入。
- ◆ 试剂盒中的终止液是酸性溶液，使用时请做好眼镜、手、面部及衣服的保护。使用后请彻底洗手。

VI. 实验前准备

A. 样品收集及储存

细胞培养上清液：颗粒物应离心去除；立刻检测样本。样本收集后若不及时检测，需按一次使用量分装，冻存于-20℃冰箱内，避免反复冻融。样本可能需要用标准品稀释剂（1×）稀释。

血清样本：用血清分离管(SST)分离血清。使血样室温凝集30分钟，然后1000 x g离心15分钟。吸取血清样本之后即刻用于检测，或者分装，-20℃贮存备用。避免反复冻融。

B. 样本准备工作

血清样本需要用标准品稀释剂（1×）2倍稀释后进行检测，例如：100μL血清样本+100μL标准品稀释剂（1×）。

C. 检测前准备工作

使用前请将所有试剂放置于室温

洗涤液：从冰箱中取出的浓缩洗涤液可能有结晶，属于正常现象；放置室温，轻摇混匀，待结晶完全溶解后再配制洗涤液。可将20mL浓缩洗涤液用蒸馏水或去离子水稀释配置成500mL工作浓度的洗涤液。未用完的放回4℃。

标准品稀释剂（1×）：使用蒸馏水或去离子水稀释配置成标准品稀释剂（1×）。

试剂稀释液（1×）：使用蒸馏水或去离子水稀释配置成试剂稀释液（1×）。

检测抗体：参考检测抗体瓶标签指示，用试剂稀释液（1×）将冻干粉进行重溶。再用标准品稀释剂（1×）稀释至工作浓度200ng/mL，至少在使用前15分钟准备。

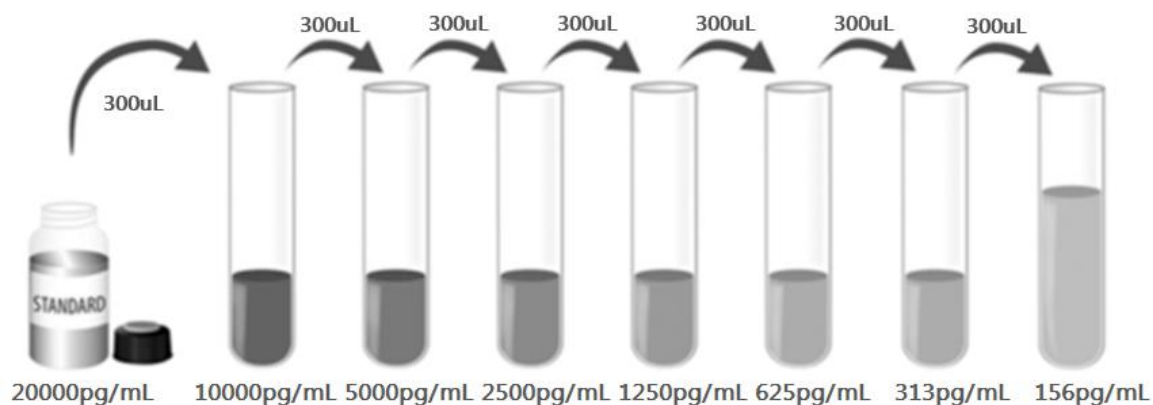
链霉亲和素- HRP A：用试剂稀释液（1×）将链霉亲和素- HRP A（200×）稀释至工作浓度链霉亲和素- HRP A（1×）。

显色剂：按试验所需用量（100μL/孔）将显色液A和显色液B等体积混合，避光保存，现用现配，须在15分钟内使用。

HGF 标准品：冻干标准品的重溶体积请参考瓶身标签，得到浓度为20000pg/mL标准品母液。轻轻震荡至少15分钟，使其充分溶解。

*如有疑问，请咨询我们的技术支持。

向各稀释管中加入300μL标准品稀释剂（1×）。将标准品母液参照下图做系列稀释，每管须充分混匀后再移液到下一管。10000pg/mL管作标准曲线最高点，标准品稀释剂（1×）可用作标准品零点（0pg/mL）。



D. 技术小提示

- ◆ 当混合或重溶蛋白液时，尽量避免起沫；
- ◆ 为了避免交叉污染，配置不同浓度标准品、上样、加不同试剂都需要更换枪头。另外不同试剂请分别使用不同的移液槽；
- ◆ 建议15分钟内完成一块板的上样；
- ◆ 每次孵育时，正确使用封板胶纸可保证结果的准确性；
- ◆ 混合后的显色底物在上板前应为无色，请避光保存；加入微孔板后，将由无色变成不同深度的蓝色；
- ◆ 终止液上板顺序应同显色底物上板顺序一致；加入终止液后，孔内颜色由蓝变黄；若孔内有绿色，则表明孔内液体未混匀，请充分混合。

VII. 操作步骤

使用前请将所有试剂和样本放置于室温，建议所有的实验样本和标准品做复孔检测

1. 按照上一节的说明，准备好所有需要的试剂和标准品；
2. 从已平衡至室温的密封袋中取出微孔板，未用的板条请放回铝箔袋内，重新封口；
3. 分别将不同浓度标准品，实验样本或者质控品加入相应孔中，每孔100 μ L。用封板膜封住反应孔，室温孵育2小时。说明书提供了一张96孔模板图，可用于记录标准品和试验样本的板内位置；（样本需要稀释，详情参见样本制备部分。）
4. 将板内液体吸去，使用洗瓶、多通道洗板器或自动洗板机洗板。每孔加洗涤液400 μ L，然后将板内洗涤液吸去。重复操作3次，共洗4次。每次洗板尽量吸去残留液体会有助于得到好的实验结果。最后一次洗板结束，请将板内所有液体吸干或将板倒置，在吸水纸拍干所有残留液体；
5. 在每个微孔内加入100 μ L配置好的检测抗体（用标准品稀释剂稀释）。用封板胶纸封住反应孔，室温孵育2小时；
6. 重复第4步洗板操作；
7. 在每个微孔内加入100 μ L稀释好的链霉亲和素- HRP A工作液。用封板膜封住反应孔，室温孵育20分钟，**注意避光**；
8. 重复第4步洗板操作；
9. 在每个微孔内加入100 μ L显色剂，室温孵育20分钟，**注意避光**；
10. 在每个微孔内加入50 μ L终止液，请轻拍微孔板，使溶液混合均匀。孔内溶液颜色会从蓝色变为黄色；
11. 加入终止液后30分钟内，使用酶标仪测量450nm的吸光度值，设定540nm或570nm作为校正波长。如果没有使用双波长校正，结果准确度可能会受影响；
12. **计算结果：**将每个标准品和样品的校正吸光度值（OD₄₅₀-OD₅₄₀/OD₅₇₀），复孔读数取平均值，然后减去平均零标准品OD值。使用计算机软件作四参数逻辑（4-PL）曲线拟合创建标准曲线。另一种方法是，可以通过绘制标准品浓度做对数与相应OD值对数生成曲线，并通过回归分析确定最佳拟合线。通过样本的OD值，可从标准曲线上得到样本中小鼠HGF的浓度。如果样品被稀释，从标准曲线读取的浓度必须乘以稀释倍数。

VIII. 参考文献

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96 孔模板图

请使用 96 孔模板图来记录标准品及样本在板内的位置

