



PRODUCT INFORMATION & MANUAL

Human PD-1 Valukine™ ELISA

Catalog Number: VAL118

For the quantitative determination of natural and recombinant human Programmed Death-1(PD-1) concentrations

For research use only.
Not for diagnostic or therapeutic procedures.

Bio-Techne China Co. Ltd

P: +86 (21) 52380373 P: 8009881270 F: +86 (21) 52381001
info.cn@bio-techne.com

Please refer to the kit label for expiry date.
Novus kits are guaranteed for 3 months from date of receipt

Version 202407.7

TABLE OF CONTENTS

I. BACKGROUND	2
II. OVERVIEW	3
III. ADVANTAGES	4
IV. EXPERIMENT	6
V. KIT COMPONENTS AND STORAGE	7
VI. PREPARATION	9
VII. ASSAY PROCEDURE	11
VIII. REFERENCES	13

I. BACKGROUND

Programmed Death-1 receptor (PD-1), also known as CD279, is type I transmembrane protein belonging to the CD28 family of immune regulatory receptors (1). Other members of this family include CD28, CTLA-4, ICOS, and BTLA (2-5). The cytoplasmic tail contains two tyrosine residues that form the immunoreceptor tyrosine-based inhibitory motif (ITIM) and immunoreceptor tyrosine-based switch motif (ITSM) that are important for mediating PD-1 signaling. PD-1 acts as a monomeric receptor and interacts in a 1:1 stoichiometric ratio with its ligands PD-L1 (B7-H1) and PD-L2 (B7-DC) (6, 7). PD-1 is expressed on activated T cells, B cells, monocytes, and dendritic cells while PD-L1 expression is constitutive on the same cells and also on non hematopoietic cells such as lung endothelial cells and hepatocytes (8, 9). Ligation of PD-L1 with PD-1 induces co-inhibitory signals on T cells promoting their apoptosis, anergy, and functional exhaustion (10). Thus, the PD-1: PD-L1 interaction is a key regulator of the threshold of immune response and peripheral immune tolerance (11). Finally, blockade of the PD-1: PD-L1 interaction by either antibodies or genetic manipulation accelerates tumor eradication and shows potential for improving cancer immunotherapy (12, 13).

II. OVERVIEW

A. PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for human PD-1 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any PD-1 present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated detection antibody specific for human PD-1 is pipetted into the wells. After washing away any unbound substances, Streptavidin-HRP is pipetted into the wells. Following a wash to remove any unbound reagent, TMB substrate solution (Chromogenic agent) is added to the wells and color develops in proportion to the amount of human PD-1 bound in the initial step. The color development is stopped, and the intensity of the color is measured.

B. LIMITATIONS OF THE PROCEDURE

- ◆ **FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.**
- ◆ This kit is suitable for cell culture supernate, human serum and plasma.
- ◆ The kit should not be used beyond the expiration date on the kit label.
- ◆ Do not mix or substitute reagents with those from other lots or sources.
- ◆ If samples generate values higher than the highest standard, dilute the samples with Reagent Diluent (1×) and repeat the assay.
- ◆ Any variation in operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

III. ADVANTAGES

A. PRECISION

Intra-assay Precision (Precision within an assay)

Three samples were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

Three samples were tested in twenty separate assays to assess inter-assay precision.

Sample	Intra-assay Precision			Inter-assay Precision		
	1	2	3	1	2	3
Mean (pg/mL)	510.0	1350.0	6895.8	493.6	1389.0	6945.4
Standard Deviation	35.8	70.3	556.7	37.6	94.5	365.8
CV%	7.0	5.2	8.1	7.6	6.8	5.3

B. RECOVERY

The recovery of human PD-1 spiked to levels throughout the range of the assay in various matrices was evaluated. The recovery ranged from 76.6 to 114.3% with an average of 92.9%.

C. SENSITIVITY

The minimum detectable dose (MDD) of human PD-1 is typically less than 14.7 pg/mL.

The MDD was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

D. CALIBRATION

This immunoassay is calibrated against a highly purified NS0-expressed recombinant human PD-1 produced at R&D Systems®.

E. LINEARITY

To assess the linearity of the assay, samples were spiked with high concentrations of human PD-1 in various matrices and diluted with Reagent Diluent (1×) to produce samples with values within the dynamic range of the assay.

Dilution	Average % of Expected	Range (%)
1:2	104.1	86.2-113.2
1:4	107.7	95.9-118.9
1:8	105.8	90.1-122.3
1:16	108.6	90.1-126.8

F. SPECIFICITY

This assay recognizes both natural and recombinant human PD-1. The following factors prepared at 100 ng/mL were assayed and exhibited no cross-reactivity or interference.

Recombinant human
B7-H1/Fc Chimera
CD28/Fc Chimera
CTLA-4/Fc Chimera
ICOS/ Fc Chimera

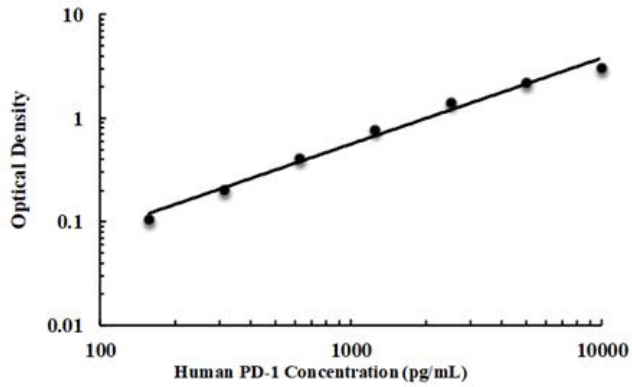
A sample containing 50 ng/mL of recombinant mouse PD-1/Fc Chimera reads as 724 pg/mL (1.4% cross-reactivity).

Recombinant human PD-L2/Fc Chimera does not cross-react in this assay but does interfere at concentrations > 25 ng/mL.

IV. EXPERIMENT

EXAMPLE STANDARD

The standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.



pg/mL	OD	Average	Corrected
0	0.027 0.027	0.027	—
156.3	0.130 0.135	0.133	0.106
312.5	0.222 0.236	0.229	0.202
625	0.419 0.448	0.434	0.407
1250	0.781 0.813	0.797	0.770
2500	1.376 1.506	1.441	1.414
5000	2.215 2.264	2.239	2.212
10000	3.065 3.078	3.072	3.045

V. KIT COMPONENTS AND STORAGE

A. MATERIALS PROVIDED

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

Parts	Description	Size
Human PD-1 Microplate	96 well polystyrene microplate (12 strips of 8 wells) coated with an antibody against human PD-1.	1 plate
Human PD-1 Standard	Recombinant human PD-1 in a buffered protein base; lyophilized. Refer to the vial label for reconstitution volume.	2 vials
Human PD-1 Detection Antibody	Biotinylated human PD-1 antibody, lyophilized. Refer to the vial label for reconstitution volume.	1 vial
Streptavidin-HRP A (200×)	200× Streptavidin conjugated to horseradish peroxidase.	1 vial
Reagent Diluent Concentrate (10×)	A 10× concentrated buffered protein base used to dilute standard, sample, detection antibody and HRP.	1 vial
Wash Buffer Concentrate (25×)	A 25× concentrated solution of buffered surfactant with preservatives.	1 vial
TMB Substrate	TMB ELISA Substrate Solution/TMB Substrate Solution	1 vial
Stop Solution	2 N sulfuric acid.	1 vial
Plate Sealers	Adhesive strip.	3 strips

B. STORAGE

Unopened Kit	Store at 2-8 °C. Do not use past kit expiration date.	
Opened/ Reconstituted Reagents	Streptavidin-HRP A	May be stored for up to 1 month at 2-8 °C.*
	Wash Buffer (1×)	
	TMB Substrate	
	Stop Solution	
	Standard	Prepare fresh for each assay. Standards may be stored for up to 1 month at -20 °C.*
	Detection Antibody	Aliquot and store for up to 1 month at -20 °C in a manual defrost freezer. *
	Reagent Diluent Concentrate (10×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Reagent Diluent (1×). Prepare fresh for each assay.
Microplate Wells	Return unused wells to the foil pouch containing the desiccant pack, reseal along entire edge of zip-seal. May be stored for up to 1 month at 2-8 °C.*	

* Provided this is within the expiration date of the kit.

C. OTHER SUPPLIES REQUIRED

- ◆ Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- ◆ Pipettes and pipette tips.
- ◆ Deionized or distilled water.
- ◆ Squirt bottle, manifold dispenser, or automated microplate washer.
- ◆ Test tubes for dilution of standards.
- ◆ 100mL and 500 mL graduated cylinder.

D. PRECAUTION

- ◆ Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.
- ◆ The Stop Solution provided with this kit is an acid solution. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.

VI. PREPARATION

A. SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. Samples may require dilution with Reagent Diluent (1 \times).

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. Samples may require dilution with Reagent Diluent (1 \times).

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. Samples may require dilution with Reagent Diluent (1 \times).

B. SAMPLE PREPARATION

Human serum samples recommend a 10-fold dilution. A suggested 10-fold dilution is 20 μL of sample + 180 μL of Reagent Diluent (1 \times). Optimal dilutions should be determined by the end user.

Human plasma samples recommend a 2-fold dilution. A suggested 2-fold dilution is 100 μL of sample + 100 μL of Reagent Diluent (1 \times). Optimal dilutions should be determined by the end user.

C. REAGENT PREPARATION

Note: *Bring all reagents to room temperature before use.*

Wash Buffer (1 \times) - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate (25 \times) into deionized or distilled water to prepare 500 mL of Wash Buffer (1 \times).

Reagent Diluent (1 \times) - Use deionized or distilled water to prepare Reagent Diluent (1 \times).

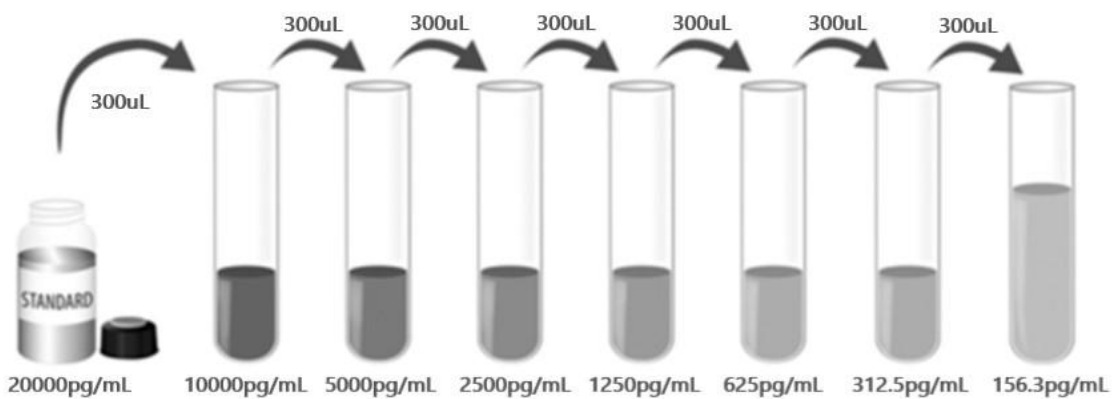
Detection Antibody (1 \times) - **Centrifuge briefly before opening. Reconstitution volume refer to vial label to prepare Detection Antibody (100 \times).** Allow the Detection Antibody to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Aliquot and store if needed. Dilute to Detection Antibody (1 \times) with Reagent Diluent (1 \times). Prepare at least 15 minutes prior to use.

Streptavidin-HRP A (1 \times) - **Centrifuge briefly before opening.** Dilute to the working concentration specified on the vial label using Reagent Diluent (1 \times).

Human PD-1 Standard - Centrifuge briefly before opening. Refer to the vial label for the reconstitution volume*. This reconstitution produces a stock solution of 20000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

*If you have any question, please seek help from our Technical Support.

Pipette 300 μ L of Reagent Diluent (1 \times) into each tube. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 10000 pg/mL standard serves as the high standard. The Reagent Diluent (1 \times) serves as the zero standard (0 pg/mL).



D. TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- It is recommended that the samples be pipetted within 15 minutes.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- TMB Substrate should remain colorless until added to the plate. Keep TMB Substrate protected from light. TMB Substrate should change from colorless to gradations of blue.
- Stop Solution should be added to the plate in the same order as the TMB Substrate. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the TMB Substrate.

VII. ASSAY PROCEDURE

Note: *Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.*

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 100 μ L of standard and prepared sample per well. Cover with the adhesive strip provided. **Incubate for 2 hours at room temperature.** A plate layout is provided for a record of standards and samples assayed. (Samples may require dilution. See Sample Preparation section.)
4. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (400 μ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 μ L of the Detection Antibody (1 \times) diluted in Reagent Diluent (1 \times), to each well. Cover with a new adhesive strip and **incubate 2 hours at room temperature.**
6. Repeat the aspiration/wash as in step 4.
7. Add 100 μ L of the working dilution of Streptavidin-HRP A to each well. Cover the plate and **incubate for 20 minutes at room temperature. Protect from light.**
8. Repeat the aspiration/wash as in step 4.
9. Add 100 μ L of TMB Substrate to each well. **Incubate for 20 minutes at room temperature. Protect from light.**
10. Add 50 μ L of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
11. Determine the optical density of each well within 10 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in

the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

12. **CALCULATION OF RESULTS**

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density (O.D.). Create a standard curve by reducing the data using computer software capable of generating a four-parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the human PD-1 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

VIII. REFERENCES

1. Ishida, Y. et al. (1992) EMBO J. 11:3887.
2. Sharpe, A.H. and G. J. Freeman (2002) Nat. Rev. Immunol. 2:116.
3. Coyle, A. and J. Gutierrez-Ramos (2001) Nat. Immunol. 2:203.
4. Nishimura, H. and T. Honjo (2001) Trends Immunol. 22:265.
5. Watanabe, N et al. (2003) Nat. Immunol. 4:670.
6. Zhang, X. et al. (2004) Immunity 20:337.
7. Lázár-Molnár, E. et al. (2008) Proc. Natl. Acad. Sci. USA 105:10483.
8. Nishimura, H et al. (1996) Int. Immunol. 8:773.
9. Keir, M.E. et al. (2008) Annu. Rev. Immunol. 26:677.
10. Butte, M.J. et al. (2007) Immunity 27:111.
11. Okazaki, T. et al. (2013) Nat. Immunol. 14:1212.
12. Iwai, Y. et al. (2002) Proc. Natl. Acad. Sci. USA 99: 12293.
13. Nogrady, B. (2014) Nature 513:S10.

PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

1											
2											
3											
4											
5											
6											
7											
8											
9											
10											
11											
12											
	A	B	C	D	E	F	G	H			



产品信息及操作手册

人 PD-1 Valukine™ ELISA 试剂盒

目录号: VAL118

适用于定量检测天然和重组人 PD-1 的浓度

科研专用, 不可用于临床诊断

Bio-Techne China Co. Ltd

P: +86 (21) 52380373 P: 8009881270 F: +86 (21) 52381001

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有效期详见试剂盒包装标签

Novus 试剂盒确保在你收货日期 3 个月内有效

版本号 202407.7

目录

I. 背景	17
II. 概述	18
III. 优势	19
IV. 实验	21
V. 试剂盒组成及储存	22
VI. 实验前准备	24
VII. 操作步骤	26
VIII. 参考文献	27

I. 背景

程序性死亡-1受体(PD-1)又称CD279,是免疫调节受体CD28家族的I型跨膜蛋白(1),该家族的其他成员包括CD28、CTLA-4、ICOS和BTLA(2-5)。其细胞质尾部含有两个酪氨酸残基,组成了基于酪氨酸的免疫受体抑制结构域(ITIM)和开关结构域(ITSM),这两个结构域对介导PD-1信号转导非常重要。PD-1作为一个单体受体,能与其配体PD-L1(B7-H1)和PD-L2(B7-DC)以1:1的比例相互作用(6,7)。PD-1在活化的T细胞、B细胞、单核细胞和树突状细胞中有表达,而PD-L1除了在以上细胞中,还在非造血细胞(如肺内皮细胞和肝细胞等)中都有表达(8,9)。PD-L1和PD-1的结合能够诱导产生共抑制信号,促使T细胞失活、凋亡、功能衰竭(10)。因此,PD-1和PD-L1相互作用是免疫反应阈值和外周免疫耐受的关键调控因子(11)。通过抗体或基因调控阻断PD-1和PD-L1的相互作用可以加速肿瘤根除,提示了PD-1作为癌症免疫治疗靶标的潜力(12,13)。

II. 概述

A. 检测原理

本实验采用双抗体夹心 ELISA 法。抗人 PD-1 捕获抗体包被于微孔板上，经过孵育，样品和标准品中的 PD-1 会与固定在板上的抗体结合，游离的成分被洗去；接着加入生物素化的抗人 PD-1 检测抗体进行孵育，洗涤去除未结合的物质后，加入链霉亲和素标记的辣根过氧化物酶(Streptavidin-HRP)孵育。洗涤去除未结合的试剂后，加入 TMB 底物溶液（显色剂）。溶液颜色与结合的目标蛋白成正比；加入终止液；用酶标仪测定吸光度。

B. 检测局限

- ◆ 仅供科研使用，不可用于体外诊断；
- ◆ 该试剂盒适用于细胞培养上清、人血清和血浆样本；
- ◆ 请在试剂盒有效期内使用；
- ◆ 不同试剂盒及不同批号试剂盒的组分不能混用；
- ◆ 样本值若大于标准曲线的最高值，应将样本用试剂稀释液（1×）稀释后重新检测；
- ◆ 检测结果的不同可由多种因素引起，包括实验人员的操作、移液器的使用方式、洗板技术、反应时间或温度、试剂盒的效期等。

III. 优势

A. 精确度

板内精确度（同一板内不同孔间的精确度）

已知浓度的三个样本，在同一板内分别检测 20 次，以确定板内精确度。

板间精确度（不同板之间的精确度）

已知浓度的三个样本，在不同板中分别检测 20 次，以确定板间精确度。

	板内精确度			板间精确度		
样本	1	2	3	1	2	3
平均值 (pg/mL)	510.0	1350.0	6895.8	493.6	1389.0	6945.4
标准差	35.8	70.3	556.7	37.6	94.5	365.8
CV%	7.0	5.2	8.1	7.6	6.8	5.3

B. 回收率

在不同类型样本中掺入检测范围内不同水平的人PD-1，测定其回收率。回收率范围为 76.6-114.3%，平均回收率为92.9%。

C. 灵敏度

人 PD-1 的最低可测剂量（MDD）一般小于 14.7 pg/mL。

MDD 是根据 20 个重复的零标准品孔的吸光度值的平均值加两倍标准差计算得到的相对应浓度。

D. 校正

此 ELISA 试剂盒经由 R&D Systems®生产的 NS0 表达的高纯度重组人 PD-1 蛋白所校正。

E. 线性

在不同类型样本中掺入高浓度的人PD-1，然后用试剂稀释液（1×）将样本稀释到检测范围内，测定其线性。

稀释倍数	平均值 (%)	范围 (%)
1:2	104.1	86.2-113.2
1:4	107.7	95.9-118.9
1:8	105.8	90.1-122.3
1:16	108.6	90.1-126.8

F. 特异性

此 ELISA 法可检测天然及重组人 PD-1 蛋白。对制备的 100 ng/mL 的下列因素进行了测定，无交叉反应或干扰。

Recombinant human
B7-H1/Fc Chimera
CD28/Fc Chimera
CTLA-4/Fc Chimera
ICOS/ Fc Chimera

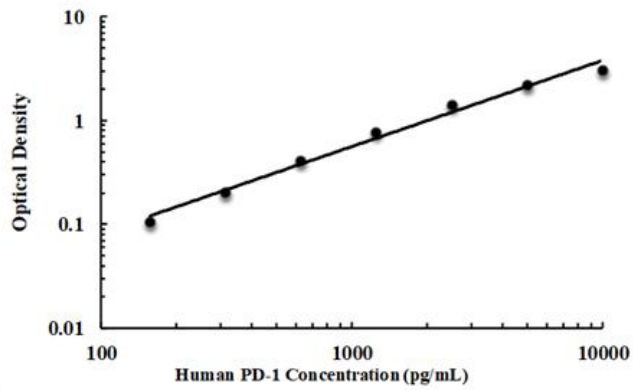
含 50 ng/mL 重组小鼠 PD-1/Fc Chimera 的样本检测值为 724 pg/mL (1.4%的交叉反应)

重组人 PD-L2/Fc Chimera 没有交叉反应，但当浓度 > 25 ng/mL 时有干扰。

IV. 实验

标准曲线实例

该标准曲线数据仅供参考，每次实验应绘制其对应的标准曲线。



pg/mL	OD	Average	Corrected
0	0.027 0.027	0.027	—
156.3	0.130 0.135	0.133	0.106
312.5	0.222 0.236	0.229	0.202
625	0.419 0.448	0.434	0.407
1250	0.781 0.813	0.797	0.770
2500	1.376 1.506	1.441	1.414
5000	2.215 2.264	2.239	2.212
10000	3.065 3.078	3.072	3.045

V. 试剂盒组成及储存

A. 试剂盒组成

组成	描述	规格
Human PD-1 Microplate	包被抗人 PD-1 抗体的 96 孔聚苯乙烯板, 8 孔 ×12 条	1 块板
Human PD-1 Standard	标准品 (冻干粉), 参考瓶标签进行重溶	2 瓶
Human PD-1 Detection Antibody	生物素化的人 PD-1 检测抗体, 冻干粉, 参考瓶标签进行重溶	1 瓶
Streptavidin-HRP A (200×)	200×浓缩的链霉亲和素标记的 HRP	1 瓶
Reagent Diluent Concentrate (10×)	浓缩的试剂稀释液 (10×) 用于稀释标准品, 样本, 检测抗体和 HRP	1 瓶
Wash Buffer Concentrate (25×)	浓缩洗涤缓冲液 (25×)	1 瓶
TMB Substrate	TMB ELISA 底物溶液/TMB 底物溶液	1 瓶
Stop Solution	终止液	1 瓶
Plate Sealers	封板膜	3 张

B. 试剂盒储存

未开封试剂盒	2-8℃储存；请在试剂盒有效期内使用	
已打开，稀释 或重溶的试剂	链霉亲和素-HRP A	2-8℃储存，最多 30 天*
	洗涤液（1×）	
	TMB 底物溶液	
	终止液	
	标准品	使用时新鲜配制* 标准品-20℃储存，最多 30 天*
	检测抗体	分装，-20℃储存，最多 30 天*
	试剂稀释液（10×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×试剂稀释液，多余的丢弃
	包被的微孔板条	将未用的板条放回带有干燥剂的铝箔袋内，密封： 2-8℃储存，最多 30 天*

*必须在试剂盒有效期内

C. 实验所需自备试验器材

- ◆ 酶标仪（可测量 450nm 检测波长的吸收值及 540nm 或 570nm 校正波长的吸收值）
- ◆ 高精度加液器及一次性吸头
- ◆ 蒸馏水或去离子水
- ◆ 洗瓶（喷瓶）、多通道洗板器或自动洗板机
- ◆ 用来稀释标准品的试管
- ◆ 100mL 及 500mL 量筒

D. 注意事项

- ◆ 试剂盒中的一些组分含有防腐剂，可能引起皮肤过敏反应，避免吸入。
- ◆ 试剂盒中的终止液是酸性溶液，使用时请做好眼睛、手、面部及衣服的保护。使用后请彻底洗手。

VI. 实验前准备

A. 样品收集及储存

细胞培养上清液：颗粒物应离心去除；立刻检测样本。样本收集后若不及时检测，需按一次使用量分装，冻存于 $\leq -20^{\circ}\text{C}$ 冰箱内，避免反复冻融。样本需要用试剂稀释液（1×）稀释。

血清样本：用血清分离管（SST）分离血清。使血样室温凝集 30 分钟，然后 $1000 \times g$ 离心 15 分钟。吸取血清样本之后即刻用于检测，或者分装， $\leq -20^{\circ}\text{C}$ 贮存备用。避免反复冻融。样本需要用试剂稀释液（1×）稀释。

血浆样本：使用 EDTA、肝素钠或枸橼酸钠作为抗凝剂收集血浆。然后 $1000 \times g$ 离心 15 分钟，需在 30 分钟内收集血浆样本之后即刻用于检测，或者分装， $\leq -20^{\circ}\text{C}$ 贮存备用。避免反复冻融。样本需要用试剂稀释液（1×）稀释。

B. 样本准备工作

人血清样本建议用试剂稀释液（1×）10 倍稀释后进行检测，即 $20 \mu\text{L}$ 样本+ $180 \mu\text{L}$ 试剂稀释液（1×）。最佳稀释度应由最终用户确定。

人血浆样本建议用试剂稀释液（1×）2 倍稀释后进行检测，即 $100 \mu\text{L}$ 样本+ $100 \mu\text{L}$ 试剂稀释液（1×）。最佳稀释度应由最终用户确定。

C. 检测前准备工作

使用前请将所有试剂放置于室温

洗涤液（1×）：从冰箱中取出的浓缩洗涤液可能有结晶，属于正常现象；放置室温，轻摇混匀，待结晶完全溶解后再配制洗涤液。可将 20 mL 浓缩洗涤液（25×）用蒸馏水或去离子水稀释配制成 500 mL 工作浓度的洗涤液（1×）。

试剂稀释液（1×）：使用去离子水或蒸馏水稀释配制成试剂稀释液（1×）。

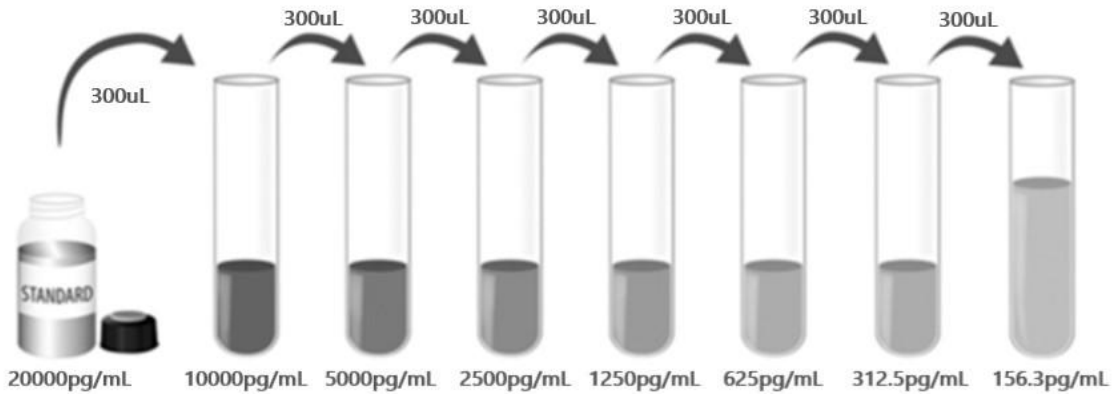
检测抗体（1×）：开盖前请瞬时离心。参考检测抗体瓶标签重溶冻干粉，制备检测抗体（100×）。轻轻震摇至少 15 分钟，使其充分溶解。如有需要分装保存。再用试剂稀释液（1×）稀释至检测抗体（1×），至少在使用前 15 分钟准备。

链霉亲和素-HRP A（1×）：开盖前请瞬时离心。用试剂稀释液（1×）将链霉亲和素-HRP A 稀释至工作浓度。

人 PD-1 标准品：开盖前请瞬时离心。冻干标准品的重溶体积请参考瓶身标签*，得到浓度为 20000 pg/mL 标准品母液。轻微震摇至少 15 分钟，使其充分溶解。

*如有疑问，请咨询我们的技术支持。

每个稀释管中加入 300 μL 试剂稀释液 (1 \times)。将标准品母液参照下图做系列稀释，每管须充分混匀后再移液到下一管。10000 pg/mL 管作标准曲线最高点，试剂稀释液 (1 \times) 可用作标准品零点 (0 pg/mL)。



D. 技术小提示

- ◆ 当混合或重溶蛋白液时，尽量避免起沫；
- ◆ 为了避免交叉污染，配制不同浓度标准品、上样、加不同试剂都需要更换枪头。另外不同试剂请分别使用不同的移液槽；
- ◆ 建议 15 分钟内完成一块板的上样；
- ◆ 每次孵育时，正确使用封板膜可保证结果的准确性；
- ◆ TMB 底物溶液在上板前应为无色，请避光保存；加入微孔板后，将由无色变成不同深度的蓝色；
- ◆ 终止液上板顺序应同 TMB 底物溶液上板顺序一致；加入终止液后，孔内颜色由蓝变黄；若孔内有绿色，则表明孔内液体未混匀，请充分混合。

VII. 操作步骤

使用前请将所有试剂和样本放置于室温，建议所有的实验样本和标准品做复孔检测。

1. 按照上一节的说明，准备好所有需要的试剂和标准品；
2. 从已平衡至室温的密封袋中取出微孔板，未用的板条请放回铝箔袋内，重新封口；
3. 分别将不同浓度标准品和实验样本加入相应孔中，每孔 100 μL 。用封板膜封住反应孔，**室温孵育 2 小时**。说明书提供了一张 96 孔模板图，可用于记录标准品和实验样本的板内位置；（样本需要稀释，详情参见样本制备部分。）
4. 将板内液体吸去，使用洗瓶、多通道洗板器或自动洗板机洗板。每孔加洗涤液 400 μL ，然后将板内洗涤液吸去。重复操作 3 次，共 4 次。每次洗板尽量吸去残留液体会有助于得到好的实验结果。最后一次洗板结束，请将板内所有液体吸干或将板倒置，在吸水纸拍干所有残留液体；
5. 在每个微孔内加入 100 μL 配制好的检测抗体（1 \times ）。用封板膜封住反应孔，**室温孵育 2 小时**；
6. 重复第 4 步洗板操作；
7. 在每个微孔内加入 100 μL 稀释好的链霉亲和素- HRP A 工作液。用封板膜封住反应孔，**室温孵育 20 分钟，注意避光**；
8. 重复第 4 步洗板操作；
9. 在每个微孔内加入 100 μL TMB 底物溶液，**室温孵育 20 分钟，注意避光**；
10. 在每个微孔内加入 50 μL 终止液，请轻拍微孔板，使溶液混合均匀。
11. 加入终止液后 10 分钟内，使用酶标仪测量 450nm 的吸光度值，设定 540nm 或 570nm 作为校正波长。如果波长校正不可用，以 450 nm 的读数减去 540 nm 或 570 nm 的读数。这种减法将校正酶标板上的光学缺陷。没有校正而直接在 450 nm 处进行的读数可能会更高且更不准确；
12. **计算结果：**将每个标准品和样品的复孔吸光值取平均值，然后减去零标准品平均 OD 值（O.D.），使用计算机软件作四参数逻辑（4-PL）曲线拟合创建标准曲线。另一替代方法是，通过绘制 y 轴上每个标准品的平均吸光值与 x 轴上的浓度来构建标准曲线，并通过图上的点绘制最佳拟合曲线。数据可以通过绘制人 PD-1 浓度的对数与 O.D. 的对数来线性化，并且最佳拟合线可以通过回归分析来确定。该程序将产生足够但不太精确的数据拟合。
如果样品被稀释，从标准曲线读取的浓度必须乘以稀释倍数。

VIII. 参考文献

1. Ishida, Y. et al. (1992) EMBO J. 11:3887.
2. Sharpe, A.H. and G. J. Freeman (2002) Nat. Rev. Immunol. 2:116.
3. Coyle, A. and J. Gutierrez-Ramos (2001) Nat. Immunol. 2:203.
4. Nishimura, H. and T. Honjo (2001) Trends Immunol. 22:265.
5. Watanabe, N et al. (2003) Nat. Immunol. 4:670.
6. Zhang, X. et al. (2004) Immunity 20:337.
7. Lázár-Molnár, E. et al. (2008) Proc. Natl. Acad. Sci. USA 105:10483.
8. Nishimura, H et al. (1996) Int. Immunol. 8:773.
9. Keir, M.E. et al. (2008) Annu. Rev. Immunol. 26:677.
10. Butte, M.J. et al. (2007) Immunity 27:111.
11. Okazaki, T. et al. (2013) Nat. Immunol. 14:1212.
12. Iwai, Y. et al. (2002) Proc. Natl. Acad. Sci. USA 99: 12293.
13. Nogrady, B. (2014) Nature 513:S10.

96 孔模板图

请使用 96 孔模板图来记录标准品及样本在板内的位置

