Magnetic Luminex® Assay

Rat Premixed Multi-Analyte Kit

Catalog Number LXSARM

For the simultaneous detection of multiple rat biomarkers in cell culture supernates, serum, and plasma.

This package insert must be read in its entirety before using this product. For research use only. Not for use in diagnostic procedures.
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INTRODUCTION
This kit contains the components required to screen multiple rat biomarkers in cell culture supernate, serum, and plasma samples in multiplexed sandwich ELISAs.

Magnetic Luminex® Assays can be used to assess the levels of biomarkers of your choosing in a single sample. For ease of use, the microparticles are premixed in one vial as are the biotinylated detection antibodies.

PRINCIPLE OF THE ASSAY
Magnetic Luminex® Assay multiplex kits are designed for use with the Luminex® MAGPIX® CCD Imager. Alternatively, kits can be used with the Luminex® 200 or Bio-Rad® Bio-Plex®, dual laser, flow-based sorting and detection platforms.

Analyte-specific antibodies are pre-coated onto color-coded magnetic microparticles. Microparticles, standards and samples are pipetted into wells and the immobilized antibodies bind the analytes of interest. After washing away any unbound substances, a biotinylated antibody cocktail specific to the analytes of interest is added to each well. Following a wash to remove any unbound biotinylated antibody, streptavidin-phycoerythrin conjugate (Streptavidin-PE), which binds to the biotinylated antibody, is added to each well. A final wash removes unbound Streptavidin-PE, the microparticles are resuspended in buffer and read using the Luminex® MAGPIX Analyzer. A magnet in the analyzer captures and holds the super paramagnetic microparticles in a monolayer. Two spectrally distinct Light Emitting Diodes (LEDs) illuminate the microparticles. One LED identifies the analyte that is being detected and the second LED determines the magnitude of the PE-derived signal, which is in direct proportion to the amount of analyte bound. Each well is imaged with a CCD camera. Kits can also be used with Luminex® 200 or a Bio-Rad Bio-Plex dual laser, flow-based systems.
LIMITATIONS OF THE PROCEDURE

• FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

• The kit should not be used beyond the expiration date on the kit label.

• Do not mix or substitute reagents with those from other lots or sources.

• If samples generate values higher than the highest standard, further dilute the samples with calibrator diluent and repeat the assay.

• Any variation in diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

• Variations in sample collection, processing, and storage may cause sample value differences.

• Discrepancies may exist in values obtained for the same analyte utilizing different technologies.

• Magnetic Luminex® Assays afford the user the benefit of multi-analyte analysis of biomarkers in a single sample. A multipurpose diluent is used to dilute samples, if necessary, and provide accurate estimates of natural analytes in cell culture supernates, serum, and plasma.

• Only the analytes listed on the enclosed Certificate of Analysis can be measured with this kit.

TECHNICAL HINTS

• When mixing or reconstituting protein solutions, always avoid foaming.

• To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.

• To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.

• Protect microparticles and Streptavidin-PE from light at all times to prevent photo bleaching.

PRECAUTIONS

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the MSDS on our website prior to use.
MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date. This kit contains sufficient materials to run multiplex assays on one 96 well plate.

<table>
<thead>
<tr>
<th>PART</th>
<th>PART #</th>
<th>DESCRIPTION</th>
<th>STORAGE OF OPENED, DILUTED, OR RECONSTITUTED MATERIAL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat Standard Cocktail A</td>
<td>893115</td>
<td>2 vials of recombinant rat biomarkers in a buffered protein base with preservatives; lyophilized.</td>
<td>Once reconstituted, any remaining standard must be discarded. Use fresh a standard for each assay.</td>
</tr>
<tr>
<td>Rat Magnetic Premixed Microparticle Cocktail</td>
<td>894782</td>
<td>0.6 mL of a concentrated microparticle cocktail with preservatives.</td>
<td>May be stored for up to 1 month at 2-8 °C.*</td>
</tr>
<tr>
<td>Rat Premixed Biotin-Ab Cocktail</td>
<td>894783</td>
<td>0.6 mL of a concentrated biotin antibody cocktail with preservatives.</td>
<td>Once diluted, these solutions must be discarded. Use fresh dilutions for each assay.</td>
</tr>
<tr>
<td>Streptavidin-PE Concentrate</td>
<td>893535</td>
<td>0.25 mL of a concentrated streptavidin-phycoerythrin conjugate with preservatives.</td>
<td></td>
</tr>
<tr>
<td>Assay Diluent RD1W</td>
<td>895117</td>
<td>11 mL of a buffered protein base with preservatives.</td>
<td>May be stored for up to 1 month at 2-8 °C.*</td>
</tr>
<tr>
<td>Calibrator Diluent RD6-52</td>
<td>895438</td>
<td>2 vials (21 mL/vial) of a buffered protein base with preservatives.</td>
<td></td>
</tr>
<tr>
<td>Wash Buffer Concentrate</td>
<td>895003</td>
<td>21 mL of a 25-fold concentrated solution of buffered surfactant with preservative. May turn yellow over time.</td>
<td></td>
</tr>
<tr>
<td>Microplate</td>
<td>641385</td>
<td>1 flat-bottomed 96 well microplate used as a vessel for the assay.</td>
<td></td>
</tr>
<tr>
<td>Certificate of Analysis</td>
<td>752296</td>
<td>1 sheet listing the selected analytes with the microparticle regions, standard reconstitution volumes, and concentrations for the provided standard(s).</td>
<td></td>
</tr>
<tr>
<td>Mixing Bottles</td>
<td>895505</td>
<td>2 empty 8 mL bottles used for mixing microparticles with Assay Diluent RD1W.</td>
<td></td>
</tr>
<tr>
<td>Plate Sealers</td>
<td>640445</td>
<td>3 adhesive foil strips.</td>
<td></td>
</tr>
</tbody>
</table>

*Provided this is within the expiration date of the kit.

OTHER SUPPLIES REQUIRED

- Luminex® MAGPIX, Luminex® 200, or Bio-Rad Bio-Plex analyzer with X-Y platform.
- Hand-held microplate magnet or plate washer with a magnetic platform.
- Pipettes and pipette tips.
- Deionized or distilled water.
- Multi-channel pipette, manifold dispenser, or automated dispensing unit.
- 500 mL graduated cylinder.
- Polypropylene test tubes for dilution of standards and samples.
- Horizontal orbital microplate shaker (0.12” orbit) capable of maintaining a speed of 800 ± 50 rpm.
- Microcentrifuge.
SAMPLE COLLECTION & STORAGE
The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

**Cell Culture Supernates** - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

**Serum** - Allow blood samples to clot for 2 hours at room temperature before centrifuging for 20 minutes at 2000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 20 minutes at 2000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

*Note:* *Citrate plasma has not been validated for use in this assay.*

SAMPLE PREPARATION
To determine the appropriate dilution for each analyte, refer to the table located in the following link [http://www.rndsystems.com/Products/LXSARM](http://www.rndsystems.com/Products/LXSARM).

Serum and plasma samples require at least a 2-fold dilution. A suggested 2-fold dilution is 75 μL of sample + 75 μL of Calibrator Diluent RD6-52. Mix thoroughly.

High abundance biomarkers may require additional dilution such as 50- or 200-fold.

A suggested 50-fold dilution is 10 μL of sample + 490 μL of Calibrator Diluent RD6-52. Mix thoroughly.

A suggested 200-fold dilution can be achieved by adding 10 μL of sample to 90 μL of Calibrator Diluent RD6-52. Complete the 200-fold dilution by adding 10 μL of the diluted sample to 190 μL Calibrator Diluent RD6-52.
REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to deionized or distilled water to prepare 500 mL of Wash Buffer.

Standard - Reconstitute the Standard Cocktail with Calibrator Diluent RD6-52. Refer to the Certificate of Analysis for the reconstitution volume. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

Use polypropylene tubes. Pipette 500 μL of the reconstituted Standard Cocktail into a tube labeled Standard 1. Pipette 200 μL of Calibrator Diluent RD6-52 into the remaining tubes. Use Standard 1 to produce a 3-fold dilution series (below). Mix each tube thoroughly before the next transfer. Standard 1 serves as the high standard. Calibrator Diluent RD6-52 serves as the blank. Refer to the Certificate of Analysis for values of Standard 1.
**DILUTED MICROPARTICLE COCKTAIL PREPARATION**

1. Centrifuge the Microparticle Cocktail vial for 30 seconds at 1000 x g prior to removing the cap.

2. Gently vortex the vial to resuspend the microparticles, taking precautions not to invert the vial.

3. Dilute the Microparticle Cocktail using Assay Diluent RD1W in the mixing bottle provided.

<table>
<thead>
<tr>
<th>Number of Wells Used</th>
<th>Microparticle Cocktail</th>
<th>+</th>
<th>Assay Diluent RD1W</th>
</tr>
</thead>
<tbody>
<tr>
<td>96</td>
<td>500 μL</td>
<td>+</td>
<td>5.00 mL</td>
</tr>
<tr>
<td>48</td>
<td>250 μL</td>
<td>+</td>
<td>2.50 mL</td>
</tr>
<tr>
<td>24</td>
<td>125 μL</td>
<td>+</td>
<td>1.25 mL</td>
</tr>
</tbody>
</table>

*Note: Protect microparticles from light during handling. Prepare microparticles within 30 minutes of use. Diluted microparticles cannot be stored.*

**DILUTED BIOTIN ANTIBODY COCKTAIL PREPARATION**

1. Centrifuge the Biotin Antibody Cocktail vial for 30 seconds at 1000 x g prior to removing the cap.

2. Gently vortex the vial, taking precautions not to invert the vial.

3. Dilute the Biotin Antibody Cocktail in Assay Diluent RD1W. Mix gently.

<table>
<thead>
<tr>
<th>Number of Wells Used</th>
<th>Biotin Antibody Cocktail</th>
<th>+</th>
<th>Assay Diluent RD1W</th>
</tr>
</thead>
<tbody>
<tr>
<td>96</td>
<td>500 μL</td>
<td>+</td>
<td>5.00 mL</td>
</tr>
<tr>
<td>48</td>
<td>250 μL</td>
<td>+</td>
<td>2.50 mL</td>
</tr>
<tr>
<td>24</td>
<td>125 μL</td>
<td>+</td>
<td>1.25 mL</td>
</tr>
</tbody>
</table>

**STREPTAVIDIN-PE PREPARATION**

*Use a polypropylene amber bottle or a polypropylene test tube wrapped with aluminum foil. Protect the Streptavidin-PE from light during handling and storage.*

1. Centrifuge the Streptavidin-PE vial for 30 seconds at 1000 x g prior to removing the cap.

2. Gently vortex the vial, taking precautions not to invert the vial.

3. Dilute the Streptavidin-PE Concentrate to a 1X concentration by adding 220 μL of Streptavidin-PE to 5.35 mL of Wash Buffer. This provides enough Streptavidin-PE to assay one 96-well microplate. If assaying less than 96 wells, adjust these volumes accordingly.
INSTRUMENT SETTINGS

Luminex® MAGPIX analyzer:

a) Assign the microparticle region for each analyte being measured (see Certificate of Analysis for details)
b) 50 events/bead
c) Sample size:
   I. 1-25 analytes: 50 μL
   II. >25 analytes: 35 μL
d) Collect Median Fluorescence Intensity (MFI)

Luminex® 200 and Bio-Rad Bio-Plex analyzers:

Note: Calibrate the analyzer using the proper reagents for superparamagnetic microparticles (refer to instrument manual).

a) Assign the microparticle region for each analyte being measured (see Certificate of Analysis for details)
b) 50 events/bead
c) Minimum events: 0
d) Flow rate: 60 μL/minute (fast)
e) Sample size: 50 μL
f) Doublet Discriminator gates at approximately 8000 and 16,500
g) Reporter Gain setting should remain on Default for accurate data analysis
h) Collect MFI

Note: The CAL2 setting for the Bio-Rad Bio-Plex analyzer should be set at the low RP1 target value.
ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.

Note: *Protect microparticles and Streptavidin-PE from light at all times.*

1. Prepare all reagents, standards, and samples as directed in the previous sections.

2. Resuspend the diluted microparticle cocktail by inversion or vortexing. Add 50 μL of the microparticle cocktail to each well of the microplate.

3. Add 50 μL of standard or sample* per well. Securely cover with a foil plate sealer. Incubate for 2 hours at room temperature on a horizontal orbital microplate shaker (0.12” orbit) set at 800 ± 50 rpm. A plate layout is provided to record standards and samples assayed.

4. Using a magnetic device designed to accommodate a microplate, wash by applying the magnet to the bottom of the microplate, allow 1 minute before removing the liquid, filling each well with Wash Buffer (100 μL) and allow 1 minute before removing the liquid again. Removal of liquid is essential for good performance. Perform the wash procedure three times.

   Note: Refer to the magnetic device user manual for proper wash technique using a flat bottom microplate.

5. Add 50 μL of diluted Biotin Antibody Cocktail to each well. Securely cover with a foil plate sealer and incubate for 1 hour at room temperature on the shaker set at 800 ± 50 rpm.

6. Repeat the wash as in step 4.

7. Add 50 μL of diluted Streptavidin-PE to each well. Securely cover with a foil plate sealer and incubate for 30 minutes at room temperature on the shaker set at 800 ± 50 rpm.

8. Repeat the wash as in step 4.

9. Resuspend the microparticles by adding 100 μL of Wash Buffer to each well. Incubate for 2 minutes on the shaker set at 800 ± 50 rpm.

10. Read within 90 minutes using a Luminex® or Bio-Rad analyzer.

    Note: Resuspend microparticles immediately prior to reading, by incubating plate for 2 minutes on the shaker set at 800 ± 50 rpm.

* Samples may require dilution. See Sample Preparation section.
**CALCULATION OF RESULTS**

Use the standard concentrations on the Certificate of Analysis and calculate 3-fold dilutions for the remaining levels. Average the duplicate readings for each standard and sample and subtract the average blank Median Fluorescence Intensity (MFI).

Create a standard curve for each analyte by reducing the data using computer software capable of generating a five parameter logistic (5-PL) curve-fit.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

**CALIBRATION**

This assay is calibrated against highly purified recombinant rat biomarkers produced at R&D Systems®.
PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

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