

Luminex® Performance Assay

Human Obesity Base Kit

Catalog Number LOBM000

For the simultaneous quantitative determination of multiple human obesity biomarker concentrations in cell culture supernates, serum, and plasma.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

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INTRODUCTION

Obesity is an escalating public health problem that markedly increases the risk of many of society's most prevalent diseases including diabetes, cardiovascular disease, cancer, and stroke. Consequently, much effort is focused on understanding the complex network of cytokines and hormones that regulate the balance between energy metabolism and appetite regulation. Altered levels of many obesity-related factors have the potential to impact these physiological processes.

Any combination of the following bead sets are suitable for use with this base kit.

Analyte	Catalog Number	Microparticle Region
Adiponectin/Acrp30	LOBM1065	46
CCL2/MCP-1	LUHM279	33
C-Reactive Protein (CRP)	LOBM1707	48
Complement Factor D	LOBM1824	47
IL-6	LUHM206	27
IL-10	LUHM217	29
Leptin	LUBM398	45
Resistin*	LOBM1359	56
Serpin E1/PAI-1	LOBM1786	26
TNF- α	LUHM210	37

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PRINCIPLE OF THE ASSAY

Luminex® Performance Assay multiplex kits are designed for use with any Luminex analyzer including the MAGPIX®, Luminex® 100/200™, FLEXMAP 3D®, xMAP INTELLIFLEX®, or Bio-Rad® Bio-Plex®, dual laser, flow-based sorting and detection platforms.

Analyte-specific antibodies are pre-coated onto magnetic microparticles embedded with fluorophores at set ratios for each unique microparticle region. Microparticles, standards and samples are pipetted into wells and the immobilized antibodies bind the analytes of interest. After washing away any unbound substances, a biotinylated antibody cocktail specific to the analytes of interest is added to each well. Following a wash to remove any unbound biotinylated antibody, streptavidin-phycoerythrin conjugate (Streptavidin-PE), which binds to the biotinylated antibody, is added to each well. Final washes remove unbound Streptavidin-PE, the microparticles are resuspended in buffer and read using the MAGPIX®. A magnet in the analyzer captures and holds the superparamagnetic microparticles in a monolayer. Two spectrally distinct Light Emitting Diodes (LEDs) illuminate the microparticles. One LED excites the dyes inside each microparticle to identify the region and the second LED excites the PE to measure the amount of analyte bound to the microparticle. A sample from each well is imaged with a CCD camera with a set of filters to differentiate excitation levels.

Analysis with the Luminex® 100/200™, FLEXMAP 3D®, xMAP INTELLIFLEX®, or Bio-Rad® Bio-Plex® uses one laser to excite the dyes inside each microparticle to identify the microparticle region and the second laser to excite the PE to measure the amount of analyte bound to the microparticle. All excitation emitted as each microparticle passes through the flow cell is then analyzed to differentiate excitation levels using a Photomultiplier Tube (PMT) and an Avalanche Photodiode.

LIMITATIONS OF THE PROCEDURE

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples fall outside the dynamic range of the assay, further dilute the samples with the appropriate calibrator diluent and repeat the assay.
- Any variation in diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- Variations in sample collection, processing, and storage may cause sample value differences.
- This assay is designed to eliminate interference by other factors present in biological samples. Until these factors have been tested in the Luminex® Performance Assay, the possibility of interference cannot be excluded.
- Luminex® Performance Assays afford the user the benefit of multi-analyte analysis of biomarkers in a single complex sample. For each sample type, a single multipurpose diluent is used to optimize recovery, linearity, and reproducibility. Such a multipurpose diluent may not optimize any single analyte to the same degree that a unique diluent selected for analysis of that analyte can optimize conditions, therefore, some performance characteristics may be more variable than those for assays designed specifically for single analyte analysis.
- **Only the analytes listed on the Standard Value Card can be measured with this base kit.**

MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past the kit expiration date.

PART	PART #	DESCRIPTION	STORAGE OF OPENED, DILUTED, OR RECONSTITUTED MATERIAL
Human Obesity Panel Standard Cocktail	892934	2 vials of recombinant human obesity biomarkers in a buffered protein base with preservatives; lyophilized.	Use fresh standards for each assay. Discard after use.
Microparticle Diluent 4	895547	6 mL of a buffered protein base with preservative.	May be stored for up to 1 month at 2-8 °C.* <i>Prepare fresh 1X solutions at the time of assay. Discard after use.</i>
Biotin Antibody Diluent 2	895832	5.5 mL of a buffered protein base with preservative.	
Streptavidin-PE	892525	0.07 mL of a concentrated streptavidin-phycoerythrin conjugate with preservatives.	
Calibrator Diluent RD5K	895119	21 mL of a buffered protein base with preservatives. <i>For cell culture supernate samples.</i>	May be stored for up to 1 month at 2-8 °C.*
Calibrator Diluent RD6-46	895883	2 vials (21 mL/vial) of a buffered protein base with preservatives. <i>For serum/plasma samples.</i>	
Wash Buffer Concentrate	895003	21 mL of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>	
Microplate	641385	1 flat-bottomed 96-well microplate used as a vessel for the assay.	
Mixing Bottles	895505	2 empty 8 mL bottles used for mixing microparticles with Microparticle Diluent.	
Plate Sealers	640445	4 adhesive foil strips.	
Standard Value Card	751249	1 card listing the standard reconstitution volume and working standard concentrations for this lot of base kit.	

*Provided this is within the expiration date of the kit.

Additional wash buffer ([#WA126](#)) and plates ([#LYX010](#)) are available for purchase.

OTHER SUPPLIES REQUIRED

- **Luminex® Performance Assay analyte-specific kit(s)** (see page 1)
- MAGPIX®, Luminex® 100/200™, FLEXMAP 3D®, xMAP INTELLIFLEX®, or Bio-Rad® Bio-Plex® analyzer with X-Y platform
- Hand-held microplate magnet or platewasher with a magnetic platform
- Pipettes and pipette tips
- Deionized or distilled water
- Multi-channel pipette, manifold dispenser, or automated dispensing unit
- 500 mL graduated cylinder
- Horizontal orbital microplate shaker (0.12" orbit) capable of maintaining a speed of 800 ± 50 rpm
- Microcentrifuge
- **Polypropylene** test tubes for dilution of standards and samples
- Luminex® Performance Assay Controls (optional; [R&D Systems®, # QC13](#))

PRECAUTIONS

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Protect microparticles and Streptavidin-PE from light at all times to prevent photobleaching.

SAMPLE COLLECTION & STORAGE

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifuging for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Note: *Citrate plasma has not been validated for use in this assay.*

SAMPLE PREPARATION

Use polypropylene tubes.

Note: *On the day of the assay, ALL fresh and previously frozen serum and plasma samples require centrifugation at 16,000 x g for 4 minutes immediately prior to use or dilution.*

Serum and plasma samples require a 4-fold dilution. A suggested 4-fold dilution is 30 μL of sample + 90 μL of Calibrator Diluent RD6-46. Mix thoroughly.

For Adiponectin, CRP, Complement Factor D, and Serpin E1, serum and plasma samples may require up to a 100-fold dilution. A suggested 100-fold dilution is 10 μL of the 4-fold dilution + 240 μL of Calibrator Diluent RD6-46. Mix thoroughly.

If expected sample values are unknown, it is recommended to evaluate both a 4-fold and a 100-fold sample dilution.

REAGENT PREPARATION

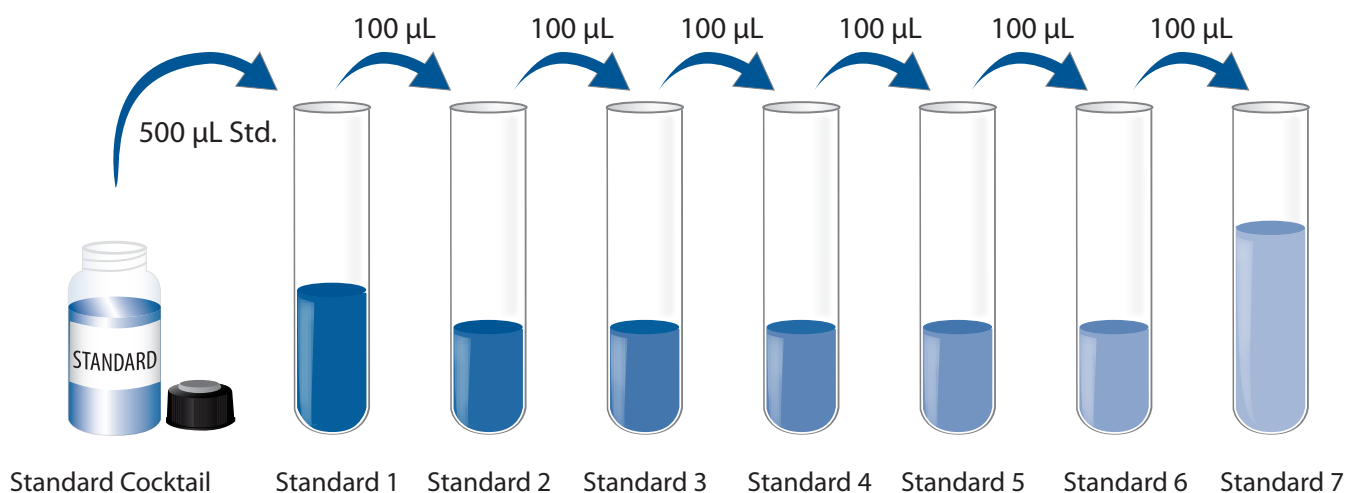
Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to 480 mL of deionized or distilled water to prepare 500 mL of Wash Buffer.

Standard - Refer to the Standard Value Card for the reconstitution volume and assigned values. Reconstitute the Standard Cocktail with Calibrator Diluent RD5K (*for cell culture supernate samples*) or Calibrator Diluent RD6-46 (*for serum/plasma samples*). Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

Note: Do NOT vortex standard cocktail.

Use polypropylene tubes. Pipette 500 μ L of the reconstituted Standard into the Standard 1 tube. Pipette 200 μ L of the appropriate calibrator diluent into the remaining tubes. Use Standard 1 to produce a 3-fold dilution series (below). Refer to the analyte specific datasheets for details. Mix each tube thoroughly before the next transfer. Standard 1 serves as the high standard. The appropriate calibrator diluent serves as the blank.



DILUTED MICROPARTICLE COCKTAIL PREPARATION

1. Centrifuge each Microparticle Concentrate vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vials to resuspend the microparticles, taking precautions not to invert the vials.
3. Dilute the Microparticle Concentrates in the mixing bottle provided. The volume of the Microparticle Concentrate listed in the table below is for each analyte (e.g. if measuring a full plate of CRP and IL-6, add 50 µL of CRP Microparticle Concentrate and 50 µL of IL-6 Microparticle Concentrate to 5 mL of Microparticle Diluent).

Number of Wells Used	Microparticle Concentrate	+	Microparticle Diluent
96	50.0 µL	+	5.00 mL
72	37.5 µL	+	3.75 mL
48	25.0 µL	+	2.50 mL
24	12.5 µL	+	1.25 mL

Note: Protect microparticles from light during handling. Diluted microparticles cannot be stored. Prepare microparticles within 30 minutes of use.

DILUTED BIOTIN-ANTIBODY COCKTAIL PREPARATION

1. Centrifuge each Biotin-Antibody vial for 30 seconds at 1000 x g prior to removing the caps.
2. Gently vortex the vials, taking precautions not to invert the vials.
3. Dilute the Biotin-Antibody Concentrates in Biotin Antibody Diluent 2. The volume of the Biotin-Antibody listed in the table below is for each analyte (e.g. if measuring a full plate, add 50 µL of each Biotin-Antibody to 5 mL of Biotin Antibody Diluent 2). Mix gently.

Number of Wells Used	Biotin-Antibody Concentrate	+	Biotin Antibody Diluent
96	50.0 µL	+	5.00 mL
72	37.5 µL	+	3.75 mL
48	25.0 µL	+	2.50 mL
24	12.5 µL	+	1.25 mL

STREPTAVIDIN-PE PREPARATION

Use a polypropylene amber bottle or a polypropylene tube wrapped with aluminum foil. Protect Streptavidin-PE from light during handling and storage.

1. Centrifuge the Streptavidin-PE vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vial, taking precautions not to invert the vial.
3. Dilute the Streptavidin-PE concentrate in Wash Buffer.

Number of Wells Used	Streptavidin-PE Concentrate	+	Wash Buffer
96	55.0 µL	+	5.50 mL
72	42.0 µL	+	4.10 mL
48	28.0 µL	+	2.75 mL
24	14.0 µL	+	1.35 mL

INSTRUMENT SETTINGS

Note: Adjust the probe height setting on the analyzer to avoid puncturing the plate. Calibrate the analyzer using the proper reagents for superparamagnetic microparticles (refer to instrument manual).

MAGPIX® analyzer:

- a) Sample volume: 50 µL
- b) Assign the microparticle region for each analyte being measured (see page 1)
- c) 50 count/region
- d) Collect Median Fluorescence Intensity (MFI)

Luminex® 100/200™, Luminex® FLEXMAP 3D®, Luminex® INTELLIFLEX, and Bio-Rad Bio-Plex analyzers:

Note: Ensure that the instrument flow rate is set to the default of 60 µL/minute (fast) for all flow based analyzers.

- a) Sample volume: 50 µL
- b) Bead Type:
 - i. Luminex® 100/200™, FLEXMAP 3D®, and xMAP INTELLIFLEX® select MagPlex
 - ii. Bio-Rad® Bio-Plex® Manager use Bio-Plex® MagPlex Beads (Magnetic)
- c) Doublet Discriminator gates:
 - i. Luminex® 100/200™ and FLEXMAP 3D®, set at 8000 and 16,500
 - ii. xMAP INTELLIFLEX® set at 7000 and 17,000
 - iii. Bio-Rad® Bio-Plex® Manager set at 8000 and 23,000
- d) Reporter Gain Setting:
 - i. Luminex® 100/200™ use Default setting
 - ii. FLEXMAP 3D® use Standard PMT setting
 - iii. xMAP INTELLIFLEX® use Luminex® 200™ Operating Mode on Low PMT setting
 - iv. Bio-Rad® Bio-Plex® Manager use the low RP1 target value for the CAL2 setting
- e) Assign the microparticle region for each analyte being measured (see page 1)
- f) 50 count/region
- g) Collect MFI

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that all standards, controls, and samples be assayed in duplicate.

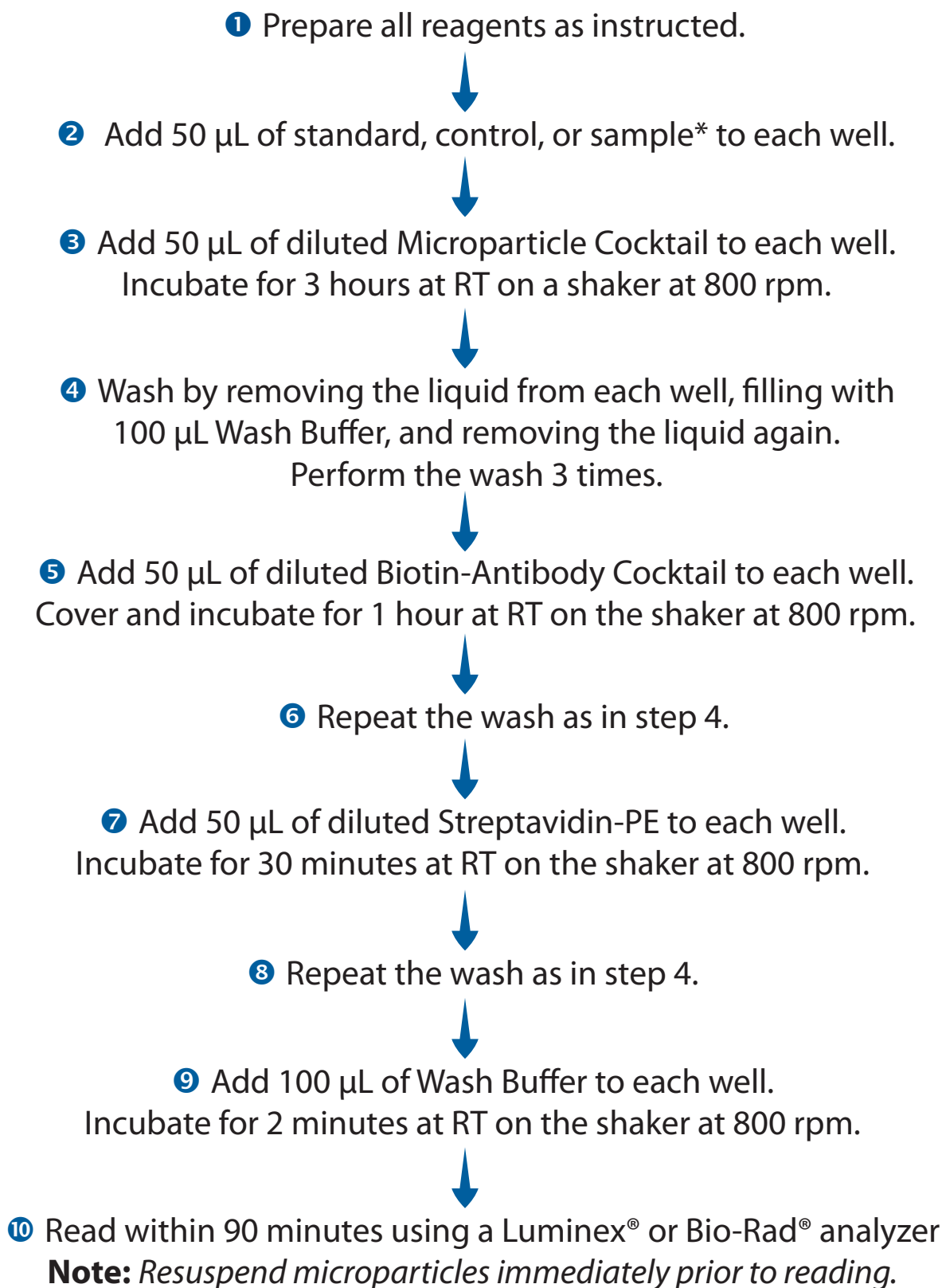
Note: *Protect microparticles and Streptavidin-PE from light at all times.*

1. Prepare all reagents, working standards, and samples as directed in the previous sections.
2. Add 50 μ L of standard, control, or sample* per well. A plate layout is provided to record standards and samples assayed.
3. Resuspend the diluted Microparticle Cocktail by inversion or vortexing. Add 50 μ L of the microparticle cocktail to each well of the microplate. Securely cover with a foil plate sealer. Incubate for 3 hours at room temperature on a horizontal orbital microplate shaker (0.12" orbit) set at 800 ± 50 rpm.
4. Using a magnetic device designed to accommodate a microplate, wash by applying the magnet to the bottom of the microplate, allow 1 minute before removing the liquid, filling each well with Wash Buffer (100 μ L) and allow 1 minute before removing the liquid again. Uniform removal of liquid is essential for good performance. **Note: Do NOT blot; this may cause a loss of microparticles.** Perform the wash procedure three times.
5. Add 50 μ L of diluted Biotin-Antibody Cocktail to each well. Securely cover with a foil plate sealer and incubate for 1 hour at room temperature on the shaker set at 800 ± 50 rpm.
6. Repeat the wash as in step 4.
7. Add 50 μ L of diluted Streptavidin-PE to each well. Securely cover with a foil plate sealer and incubate for 30 minutes at room temperature on the shaker set at 800 ± 50 rpm.
8. Repeat the wash as in step 4.
9. Resuspend the microparticles by adding 100 μ L of Wash Buffer to each well. Incubate for 2 minutes on the shaker set at 800 ± 50 rpm.
10. Read within 90 minutes using a Luminex® or Bio-Rad® analyzer.
Note: *Resuspend microparticles immediately prior to reading by shaking the plate for 2 minutes on the plate shaker at 800 ± 50 rpm.*

*Samples may require dilution. See the Sample Preparation section.

ASSAY PROCEDURE SUMMARY

Note: Protect microparticles and Streptavidin-PE from light at all times.



*Samples may require dilution. See Sample Preparation section.

CALCULATION OF RESULTS

Use the Standard concentrations on the Standard Value Card and calculate 3-fold dilutions for the remaining levels. Average the duplicate readings for each standard and sample and subtract the average blank Median Fluorescence Intensity (MFI).

Create a standard curve for each analyte by reducing the data using computer software, such as [Bio-Techne® Quantist™](#), capable of generating a five parameter logistic (5-PL) curve-fit.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

CALIBRATION

This assay is calibrated against highly purified recombinant human obesity biomarkers produced at R&D Systems®.

SPECIFICITY

The assay was tested for cross-reactivity and interference with the following factors. Less than 0.5% cross-reactivity and interference was observed unless otherwise noted on the analyte specific datasheet.

Recombinant human:

6Ckine	IL-11
CNTF	IL-12 p40
β-ECGF	IL-12 p70
FGF acidic	IL-13
FGF-4	IL-15
FGF-5	IL-16
FGF-6	IL-17
FGF-9	IL-18
FGF-10	LIF
FGF-18	LIF R
GCP-2	MIP-1δ
GROα	MIP-3α
GROβ	MIP-3β
GROγ	MCP-2
I-309	MCP-3
IGF-I	MCP-4
IGF-II	M-CSF
IL-1 RI	TNF RI
IL-1 RII	TNF-β
IL-2 Rα	VEGF ₁₂₁
IL-2 Rβ	VEGF ₁₆₅
IL-2 Ry	
IL-3 Rα	
IL-4 R	
IL-5 Rα	
IL-6 R	
IL-10 R	
IL-3	
IL-7	
IL-9	

Recombinant mouse:

G-CSF
GM-CSF
IFN-γ
IL-1α
IL-1ra
IL-1
IL-2
IL-4
IL-5
IL-6
IL-8
IL-10
IL-17
MIP-1α
MIP-1β
RANTES
Tpo
TNF-α
VEGF

Recombinant rat:

GM-CSF
IFN-γ
IL-1α
IL-1β
IL-2
IL-4
IL-6
IL-10
TNF-α

Recombinant porcine:

GM-CSF
IL-1α
IL-1β
IL-2
IL-4
IL-6
IL-8
IL-10

Recombinant human multiplex partners:

Complement Factor D
CRP
IL-6
IL-10
Leptin
MCP-1
Resistin
Serpin E1
TNF-α

PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

12								
11								
10								
9								
8								
7								
6								
5								
4								
3								
2								
1								
	A	B	C	D	E	F	G	H

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