

## PRODUCT DESCRIPTION

M1 Macrophages are associated with a high level of phagocytosis and secretion of pro-inflammatory cytokines. Their function is critical to host and anti-tumor defense against cancer. This panel contains 7 conjugated antibodies that can be used for the single step staining of M1 Macrophages.

## INTENDED USE

This multicolor flow cytometry panel was validated on human peripheral blood mononuclear cells (PBMCs).

## MATERIALS PROVIDED & STORAGE

Store the unopened kit at 2-8 °C. Do not use past expiration date.

MARKER	CLONE	FLUOROCHROME	SIZE	RECOMMENDED CONCENTRATION
CD3	UCHT1	Alexa Fluor® 405	100 tests	5 µL/10 <sup>6</sup> cells
CD11c	ICRF 3.9	PE	100 tests	10 µL/10 <sup>6</sup> cells
CD20	396444	Alexa Fluor® 405	25 tests	5 µL/10 <sup>6</sup> cells
CD38	240742	APC	100 tests	10 µL/10 <sup>6</sup> cells
CD86	37301	FITC	100 tests	10 µL/10 <sup>6</sup> cells
HLA-DR	L203	PerCP	100 tests	10 µL/10 <sup>6</sup> cells
NCAM1/CD56	2524C	Alexa Fluor® 700	100 µg	0.25 - 1 µg/10 <sup>6</sup> cells

*Note: Recommended concentrations are given as reference point for antibody titration. Optimal concentrations should be determined by each laboratory for their experimental conditions.*

## PRECAUTIONS

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

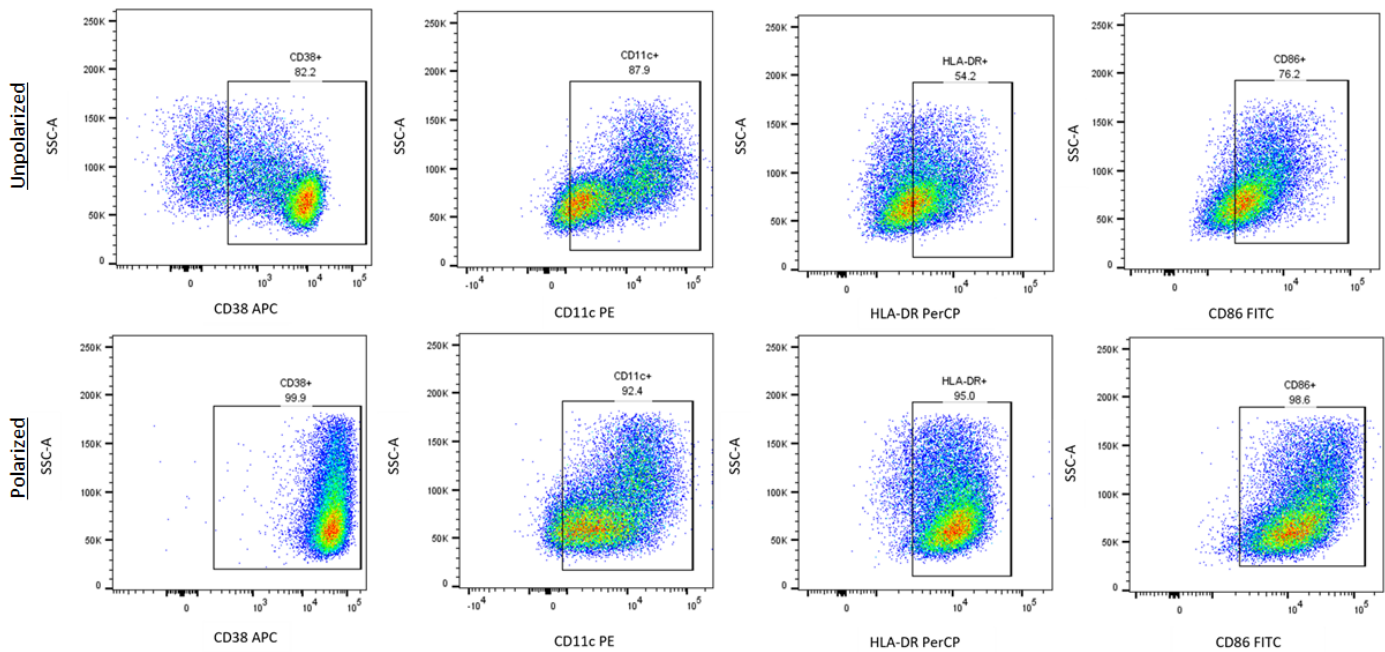
## OTHER SUPPLIES REQUIRED

- PBS
- Flow Cytometry Staining Buffer ([R&D Systems®, Catalog # FC001](#))
- Fc-block (blocking IgG)
- Isotype Control Antibodies (optional)
- 5 mL Flow cytometry tubes

## PROTOCOL

1. Wash human PBMCs (1 x 10<sup>6</sup> cells per sample) with 2 mL of Staining Buffer (1X) ([R&D Systems®, Catalog # FC001](#)) or other BSA-containing buffer, by spinning at 300 x g for 5 minutes, using 5 mL flow cytometry tubes. Decant/aspirate supernatant.
2. Fc-block cells with blocking IgG (1 µg IgG/10<sup>6</sup> cells) for 10 minutes at room temperature.
3. Add previously titrated amount of each surface marker. Vortex tubes.
4. (Optional) To a separate tube, add 5 µL of each of the isotype control antibodies. Vortex tubes.
5. Incubate the mixtures for 30-45 minutes at room temperature **in the dark**.
6. Wash with 2 mL of Staining Buffer (1X), by spinning at 300 x g for 5 minutes at end of incubation. Decant/aspirate supernatant.
7. Resuspend the cells in 0.2 - 0.5 mL Staining Buffer (1X) and acquire on a Flow Cytometer.

## DATA EXAMPLES



**Multicolor Flow Cytometry Panel to identify M1 Macrophages.** Monocytes were isolated from PBMCs via adherence depletion for 5 hours. Non-adherent cells were removed. Adherent cells were incubated for 6 days in C3<sup>+</sup> 10% huAB media with 50 ng/mL Recombinant Human GM-CSF ([Catalog # 215-GM](#)), media and cytokines are refreshed on day 3. For M1 polarization, 24 hours prior to staining, cells are incubated with 50 ng/mL human GM-CSF ([Catalog # 215-GM](#)), 50 ng/mL Recombinant Human IFN- $\gamma$  ([Catalog # 285-IF](#)), and 50 ng/mL LPS. Cells were stained with Anti-human CD3 Alexa Fluor<sup>®</sup> 405, CD20 Alexa Fluor 405, CD56 Alexa Fluor 700, CD86 FITC, HLA-DR PerCP, CD11c PE, and CD38 APC. Cells were previously gated on Live CD3<sup>+</sup>CD20<sup>-</sup>CD56<sup>-</sup>.