

# Intracellular NK Cell Activation

Multi-Color Flow Cytometry Panel
Catalog Number: FMC-P-005

## PRODUCT DESCRIPTION

Natural Killer (NK) cells are innate lymphocytes that are critical for host defense against viral infection and cancer. This panel contains 6 conjugated antibodies that can be used for investigating expression of intracellular effector molecules.

## **INTENDED USE**

This multicolor flow cytometry panel was validated on human peripheral blood mononuclear cells (PBMCs).

## **MATERIALS PROVIDED & STORAGE**

Store the unopened kit at 2-8 °C. Do not use past expiration date.

SURFACE/INTRACELLULAR	MARKER	CLONE	FLUOROCHROME	SIZE	RECOMMENDED CONCENTRATION
Surface	CD3	UCHT1	Alexa Fluor® 405	100 tests	5 μL/10 <sup>6</sup> cells
	NCAM1/CD56	2524C	Alexa Fluor® 647	100 μg	0.25-1 μg/10 <sup>6</sup> cells
	Fc Gamma R III (CD16)	245536	PerCP	100 tests	10 μL/10 <sup>6</sup> cells
Intracellular	Granzyme B	351927	Alexa Fluor® 488	100 tests	5 μL/10 <sup>6</sup> cells
	Granzyme K	2471A	Alexa Fluor® 594	100 μg	0.1-1 μg/10 <sup>6</sup> cells
	Perforin-1	1031751	Alexa Fluor® 750	100 μg	0.25 - 1 μg/10 <sup>6</sup> cells

Note: Recommended concentrations are given as reference point for antibody titration. Optimal concentrations should be determined by each laboratory for their experimental conditions.

## **PRECAUTIONS**

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

## **OTHER SUPPLIES REQUIRED**

- PBS
- Flow Cytometry Staining Buffer (R&D Systems®, Catalog # FC001)
- Flow Cytometry Fixation Buffer (R&D Systems, Catalog # FC004)
- Flow Cytometry Permeabilization Buffer/Wash Buffer I (R&D Systems, Catalog # FC005)
- Fc-block (blocking IgG)
- (Optional; Isotype Control Antibodies)
- 5 mL Flow cytometry tubes

#### **PROTOCOL**

### **Surface Staining**

- 1. Wash human PBMCs (1 x 10<sup>6</sup> cells per sample) with 2 mL of Staining Buffer (1X) (R&D Systems, Catalog # FC001) or other BSA-containing buffer, by spinning at 300 x g for 5 minutes, using 5 mL flow cytometry tubes. Decant/aspirate supernatant.
- 2. Fc-block cells with blocking IgG (1 µg IgG/106 cells) for 10 minutes at room temperature.
- 3. Add previously titrated amount of each surface marker. Vortex tubes.
- 4. (optional) To separate tube, add 5 µL of each of the isotype control antibodies. Vortex tubes.
- 5. Incubate the mixtures for 30-45 minutes at room temperature **in the dark**.
- 6. Wash with 2 mL of Staining Buffer (1X), by spinning at 300 x g for 5 minutes at end of incubation. Decant/aspirate supernatant.

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## **PROTOCOL** continued

## Intracellular Stain (with detergent permeabilization)

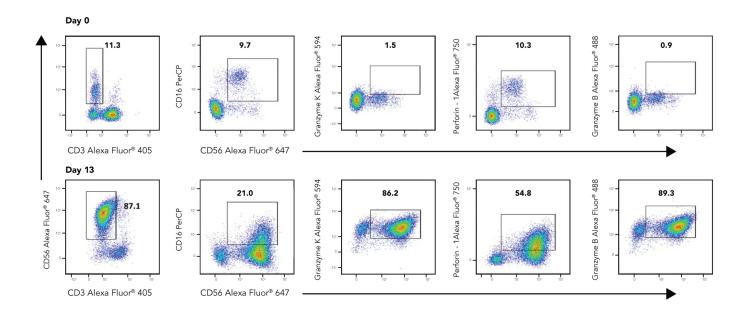
- 1. Add 0.5 mL of cold Flow Cytometry Fixation Buffer (R&D Systems® Catalog # FC004) and vortex. Incubate at room temperature for 10 minutes. Vortex cells intermittently to maintain a single cell suspension.
- 2. Centrifuge cells 350 500 x g for 5 minutes. Decant the Fixation Buffer.
- 3. Wash cells PBS (or HBSS) by adding 2 mL of PBS (or HBSS), centrifuge at 350 500 x g for 5 minutes, and decant buffer from pelleted cells. Repeat (2 total washes).
- 4. Resuspend the cell pellet in 0.1 0.2 mL of Flow Cytometry Permeabilization Buffer/Wash Buffer I (R&D Systems Catalog # FC005)

**Note:** Saponin-mediated cell permeabilization is a reversible process, it is important to keep the cells in the presence of Permeabilization Buffer I during intracellular staining.

**Note:** Depending on the specific antibody and cell sample being used, the fixation and permeabilization steps can be performed simultaneously using Flow Cytometry Fixation/Permeabilization Buffer I (R&D Systems, Catalog # FC007).

- 5. Add previously titrated amount of conjugated antibody to intracellular targets. Vortex tubes.
- 6. Incubate cells for 30 minutes at room temperature in the dark.
- 7. Wash cells 2 times with Flow Cytometry Permeabilization Buffer/Wash Buffer I (R&D Systems, Catalog # FC005) as described in Step 3.
- 8. Resuspend the cells in 0.2 0.5 mL PBS (or HBSS) buffer and acquire on a Flow Cytometer.

## **DATA EXAMPLES**



Intracellular NK Cell analysis over a 14-day time course. NK cells were expanded from PBMCs using plate-bound Anti-Human NKp46 Monoclonal Antibody (Catalog # MAB1850) in ExCellerate™ Human NK Cell Expansion Media, Xeno-Free (Catalog # CCM032) plus rhIL-2 (27 ng/mL, Catalog # 202-IL), rhIL-12 (10 ng/mL, Catalog # 219-IL), rhIL-18 (10 ng/mL, Catalog # 9124-IL), rhIL-21 (10 ng/mL, Catalog # 8879-IL) for 13 days. NK cells were assessed on Days 0 and 13 for expression of human CD16 PerCP, Granzyme K Alexa Fluor® 594, Granzyme B Alexa Fluor 488, Perforin Alexa Fluor 750, CD56 Alexa Fluor 647, and CD3 Alexa Fluor 405. Cells were gated on singlets (FSC-A by FSC-H) and live cells.

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