

## PRODUCT DESCRIPTION

Regulatory T (Treg) cells are a critical subset of CD4<sup>+</sup> T cells that regulate the immune response and prevent autoimmunity. They are capable of inducing apoptosis, increasing cellular adenosine, and inhibiting DC maturation. This panel contains 6 conjugated antibodies that can be used for staining Tregs.

## INTENDED USE

This multicolor flow cytometry panel was validated on human peripheral blood mononuclear cells (PBMCs).

## MATERIALS PROVIDED & STORAGE

Store the unopened kit at 2-8 °C. Do not use past expiration date.

SURFACE/INTRACELLULAR	MARKER	CLONE	FLUOROCHROME	SIZE	RECOMMENDED CONCENTRATION
Surface	CD3	UCHT1	Alexa Fluor® 405	100 tests	5 µL/10 <sup>6</sup> cells
	CD4	11830	PerCP	100 tests	10 µL/10 <sup>6</sup> cells
	CD25/IL-2R	24212	PE	100 tests	10 µL/10 <sup>6</sup> cells
	CD127/IL-7R	40131	Alexa Fluor® 700	100 tests	5 µL/10 <sup>6</sup> cells
	GITR/TNFRSF18	110416	FITC	100 tests	10 µL/10 <sup>6</sup> cells
Intracellular	FoxP3	376209	Alexa Fluor® 647	100 tests	10 µL/10 <sup>6</sup> cells

*Note: Recommended concentrations are given as reference point for antibody titration. Optimal concentrations should be determined by each laboratory for their experimental conditions.*

## PRECAUTIONS

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

## OTHER SUPPLIES REQUIRED

- PBS
- Flow Cytometry Staining Buffer ([R&D Systems®, Catalog # FC001](#))
- Flow Cytometry Fixation Buffer ([R&D Systems, Catalog # FC004](#))
- Flow Cytometry Permeabilization Buffer/Wash Buffer I ([R&D Systems, Catalog # FC005](#))
- Fc-block (blocking IgG)
- (Optional; Isotype Control Antibodies)
- 5 mL Flow cytometry tubes

## PROTOCOL

### Surface Staining

1. Wash human PBMCs (1 x 10<sup>6</sup> cells per sample) with 2 mL of Staining Buffer (1X) ([R&D Systems® Catalog # FC001](#)) or other BSA-containing buffer, by spinning at 300 x g for 5 minutes, using 5 mL flow cytometry tubes. Decant/aspirate supernatant.
2. Fc-block cells with blocking IgG (1 µg IgG/10<sup>6</sup> cells) for 10 minutes at room temperature.
3. Add previously titrated amount of each surface marker. Vortex tubes.
4. (optional) To separate tube, add 5 µL of each of the isotype control antibodies. Vortex tubes.
5. Incubate the mixtures for 30-45 minutes at room temperature **in the dark**.
6. Wash with 2 mL of Staining Buffer (1X), by spinning at 300 x g for 5 minutes at end of incubation. Decant/aspirate supernatant.

## PROTOCOL *continued*

### Intracellular Stain (with detergent permeabilization)

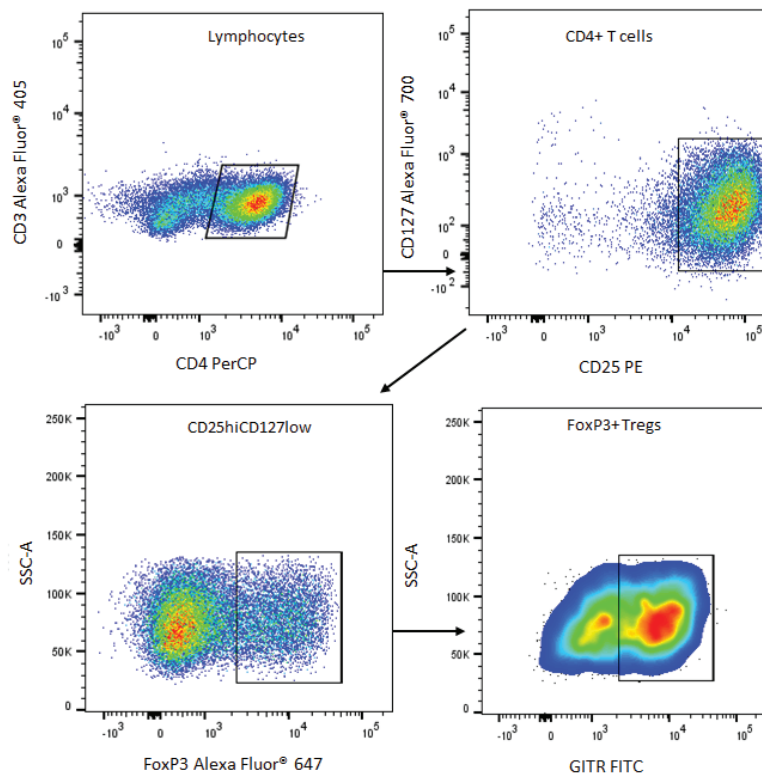
1. Add 0.5 mL of cold Flow Cytometry Fixation Buffer ([R&D Systems® Catalog # FC004](#)) and vortex. Incubate at room temperature for 10 minutes. Vortex cells intermittently to maintain a single cell suspension.
2. Centrifuge cells 350 - 500 x g for 5 minutes. Decant the Fixation Buffer.
3. Wash cells PBS (or HBSS) by adding 2 mL of PBS (or HBSS), centrifuge at 350-500 x g for 5 minutes, and decant buffer from pelleted cells. Repeat (2 total washes).
4. Resuspend the cell pellet in 0.1 - 0.2 mL of Flow Cytometry Permeabilization Buffer/Wash Buffer I ([R&D Systems Catalog # FC005](#))

**Note:** Saponin-mediated cell permeabilization is a reversible process, it is important to keep the cells in the presence of Permeabilization Buffer I during intracellular staining.

**Note:** Depending on the specific antibody and cell sample being used, the fixation and permeabilization steps can be performed simultaneously using Flow Cytometry Fixation/Permeabilization Buffer I ([R&D Systems, Catalog # FC007](#)).

5. Add previously titrated amount of conjugated antibody to intracellular targets. Vortex tubes.
6. Incubate cells for 30 minutes at room temperature **in the dark**.
7. Wash cells 2 times with Flow Cytometry Permeabilization Buffer/Wash Buffer I ([R&D Systems Catalog # FC005](#)) as described in Step 3.
8. Resuspend the cells in 0.2 - 0.5 mL PBS (or HBSS) buffer and acquire on a Flow Cytometer.

### DATA EXAMPLES



**Multicolor flow cytometry panel to identify human T Regulatory Cells (Tregs).** Human T regulatory cells were expanded from PBMCs using Anti-Human CD3 Monoclonal Antibody (10 µg/mL, [Catalog # MAB100](#)) and Anti-human CD28 Monoclonal Antibody (5 µg/mL, [Catalog # AF-342-PB](#)) plus rhIL-2 (20 ng/mL, [Catalog # 202-IL](#)), rhTGF-β (10 ng/mL, [Catalog # 7754-BH](#)) for 2 days. Cells were stained with CD3 Alexa Fluor<sup>®</sup> 405, CD4 PerCP, CD25 PE, CD127 Alexa Fluor 700, FoxP3 Alexa Fluor 647, and GITR FITC. Tregs are CD4<sup>+</sup>, CD3<sup>+</sup>, CD25<sup>hi</sup>, CD127<sup>low</sup>, Foxp3<sup>+</sup>. GITR is used as additional marker of human Tregs.