

biotechne®

R&D SYSTEMS

Luminex® Performance Assay

TGF- β Premixed Kit

Catalog Number FCSTM17

For the quantitative determination of transforming growth factor beta 1, 2, and 3 (TGF- β 1, TGF- β 2, and TGF- β 3) concentrations in cell culture supernates, serum, plasma, urine, and human milk.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

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INTRODUCTION

Transforming Growth Factor Beta 1, 2, and 3 (TGF- β 1, TGF- β 2, and TGF- β 3) are highly pleiotropic cytokines that are secreted by virtually all cell types. TGF- β molecules are proposed to act as cellular switches that regulate processes such as immune function, proliferation, and epithelial-mesenchymal transition (1-4). Targeted deletions of these genes in mice show that each TGF- β isoform has some non-redundant functions: TGF- β 1 is involved in hematopoiesis and endothelial differentiation; TGF- β 2 affects development of cardiac, lung, craniofacial, limb, eye, ear, and urogenital systems; and TGF- β 3 influences palatogenesis and pulmonary development (2). TGF- β ligands are initially synthesized as precursor proteins that undergo proteolytic cleavage. The mature segments form active ligand dimers via a disulfide-rich core consisting of the characteristic "cysteine knot" (1-4). TGF- β signaling begins with binding to a complex of the accessory receptor betaglycan (also known as TGF- β RIII) and a type II serine/threonine kinase receptor termed TGF- β RII. This receptor then phosphorylates and activates a type I serine/threonine kinase receptor, either ALK-1 or TGF- β RI (also called ALK-5). The activated type I receptor phosphorylates and activates Smad proteins that regulate transcription (3, 5, 6). Use of other signaling pathways that are Smad-independent allows for distinct actions observed in response to TGF- β in different contexts (5).

This kit can be used to assess the levels of three TGF- β molecules. Due to high cross-species homology, human, mouse, rat, and porcine samples can be tested in this assay. For ease of use, the TGF- β microparticles are pre-mixed in one vial and the biotinylated detection antibodies are pre-mixed as well. Due to a low level of cross-reactivity of the biotinylated anti-TGF- β 1 antibody with TGF- β 2, multiplexing TGF- β 1 and TGF- β 2 can result in falsely elevated values for TGF- β 2 in some sample types. We found in human serum samples, falsely elevated values for TGF- β 2 when multiplexing TGF- β 1 and TGF- β 2. Similar cross-reactivity was observed in competitor's multiplex assays. For studies using human serum samples, TGF- β 1 and TGF- β 3, or TGF- β 2 and TGF- β 3 can be multiplexed; however, careful consideration should be given when deciding to evaluate TGF- β 1 and TGF- β 2 as a multiplex. Testing with samples from healthy donors found the impact of the cross-reactivity observed became insignificant with human milk, platelet-poor plasma, cell culture supernates and urine samples. It is suggested that each lab evaluate their samples to determine if multiplexing will be problematic (as TGF- β levels will vary with disease state and serum source for media).

| Analyte | Microparticle Region |
|----------------|----------------------|
| TGF- β 1 | 34 |
| TGF- β 2 | 15 |
| TGF- β 3 | 22 |

PRINCIPLE OF THE ASSAY

Luminex® Performance Assay multiplex kits are designed for use with any Luminex analyzer including the MAGPIX®, Luminex® 100/200™, FLEXMAP 3D®, xMAP INTELLIFLEX®, or Bio-Rad® Bio-Plex®, dual laser, flow-based sorting and detection platforms.

Analyte-specific antibodies are pre-coated onto magnetic microparticles embedded with fluorophores at set ratios for each unique microparticle region. Microparticles, standards and samples are pipetted into wells and the immobilized antibodies bind the analytes of interest. After washing away any unbound substances, a biotinylated antibody cocktail specific to the analytes of interest is added to each well. Following a wash to remove any unbound biotinylated antibody, streptavidin-phycoerythrin conjugate (Streptavidin-PE), which binds to the biotinylated antibody, is added to each well. Final washes remove unbound Streptavidin-PE, the microparticles are resuspended in buffer and read using the MAGPIX®. A magnet in the analyzer captures and holds the superparamagnetic microparticles in a monolayer. Two spectrally distinct Light Emitting Diodes (LEDs) illuminate the microparticles. One LED excites the dyes inside each microparticle to identify the region and the second LED excites the PE to measure the amount of analyte bound to the microparticle. A sample from each well is imaged with a CCD camera with a set of filters to differentiate excitation levels.

Analysis with the Luminex® 100/200™, FLEXMAP 3D®, xMAP INTELLIFLEX®, or Bio-Rad® Bio-Plex® uses one laser to excite the dyes inside each microparticle to identify the microparticle region and the second laser to excite the PE to measure the amount of analyte bound to the microparticle. All excitation emitted as each microparticle passes through the flow cell is then analyzed to differentiate excitation levels using a Photomultiplier Tube (PMT) and an Avalanche Photodiode.

LIMITATIONS OF THE PROCEDURE

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples fall outside the dynamic range of the assay, further dilute the samples with calibrator diluent and repeat the assay.
- Any variation in diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- Variations in sample collection, processing, and storage may cause sample value differences.
- This assay is designed to eliminate interference by other factors present in biological samples. Until these factors have been tested in the Luminex® Performance Assay, the possibility of interference cannot be excluded.
- Luminex® Performance Assays afford the user the benefit of multi-analyte analysis of biomarkers in a single complex sample. For each sample type, a single multipurpose diluent is used to optimize recovery, linearity, and reproducibility. Such a multipurpose diluent may not optimize any single analyte to the same degree that a unique diluent selected for analysis of that analyte can optimize conditions. Therefore, some performance characteristics may be more variable than those for assays designed specifically for single analyte analysis.
- **Only the analytes listed on the Standard Value Card can be measured with this kit.**
- **Refer to the enclosed Certificate of Analysis for specific analytes included in this kit.**

MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past the kit expiration date.

| PART | PART # | DESCRIPTION | STORAGE OF OPENED, DILUTED, OR RECONSTITUTED MATERIAL |
|--|--------|--|--|
| TGF-β Standard Cocktail | 893244 | 2 vials of recombinant TGF-β1, 2, and 3 in a buffered protein base with preservatives; lyophilized. | Discard after use. Use a fresh standard for each assay. |
| TGF-β Premixed Magnetic Microparticles | 894892 | 1.2 mL of a concentrated microparticle cocktail with preservatives. | <i>May be stored for up to 1 month at 2-8 °C.* Prepare fresh 1X solutions at the time of assay. Discard after use.</i> |
| TGF-β Premixed Biotin-Antibody | 894893 | 1.2 mL of a concentrated biotinylated antibody cocktail with preservatives. | |
| Streptavidin-PE | 892525 | 0.07 mL of a concentrated streptavidin-phycoerythrin conjugate with preservatives. | |
| Microparticle Diluent 2 | 895815 | 6 mL of a buffered protein base with preservative. | May be stored for up to 1 month at 2-8 °C.* |
| Biotin Antibody Diluent 2 | 895832 | 5.5 mL of a buffered protein base with preservative. | |
| Calibrator Diluent RD5-49 | 895584 | 21 mL of a buffered protein base with preservatives. <i>For cell culture supernate/urine samples.</i> | |
| Calibrator Diluent RD6-50 | 895912 | 21 mL of a buffered animal serum with preservatives. <i>For serum/plasma/human milk samples.</i> | |
| Wash Buffer Concentrate | 895003 | 21 mL of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i> | |
| Microplate | 641385 | 1 flat-bottomed 96-well microplate used as a vessel for the assay. | |
| Mixing Bottles | 895505 | 2 empty 8 mL bottles used for mixing microparticles with Microparticle Diluent 2. | |
| Plate Sealers | 640445 | 4 adhesive foil strips. | |
| Standard Value Card | 749814 | 1 card listing the Standard reconstitution volume and working standard concentrations for this lot of base kit. | |

*Provided this is within the expiration date of the kit.

Note: Additional wash buffer is available for purchase ([R&D Systems®](#), # WA126).

OTHER SUPPLIES REQUIRED

- MAGPIX®, Luminex® 100/200™, FLEXMAP 3D®, xMAP INTELLIFLEX®, or Bio-Rad® Bio-Plex® analyzer with X-Y platform
- Hand-held microplate magnet or platewasher with a magnetic platform
- Pipettes and pipette tips
- Deionized or distilled water
- Multi-channel pipette, manifold dispenser, or automated dispensing unit
- 100 mL and 500 mL graduated cylinders
- Horizontal orbital microplate shaker (0.12" orbit) capable of maintaining a speed of 800 ± 50 rpm
- Microcentrifuge
- **Polypropylene** test tubes for dilution of standards and samples
- Luminex® Performance Assay Control (optional; [R&D Systems®, # QC17](#))

ADDITIONAL REAGENTS REQUIRED

For sample activation:

Note: *The reagents listed below are available as Sample Activation Kit 1 ([R&D Systems, # DY010](#)).*

- Hydrochloric acid (A.C.S. Grade, 12 N)
- Sodium hydroxide (A.C.S. Grade, 10 N)
- HEPES, free acid (Reagent Grade, M.W. 238.3)

PRECAUTIONS

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Protect microparticles and Streptavidin-PE from light at all times to prevent photobleaching.

SAMPLE COLLECTION AND STORAGE

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Samples must be assayed immediately after activation. Do not freeze activated samples.

Cell Culture Supernates - Remove particulates by centrifugation and assay (see activation procedure) immediately or aliquot and store unactivated samples at ≤ -70 °C. Avoid repeated freeze-thaw cycles.

Notes: *Animal serum used in the preparation of cell culture media may contain high levels of latent TGF- β 1. For best results, do not use animal serum for growth of cell cultures when assaying for TGF- β 1 production. If animal serum is used as a supplement in the media, precautions should be taken to prepare the appropriate control and run the control in the assay to determine the baseline concentration of TGF- β 1.*

A 10% solution of bovine serum used in cell culture can be expected to contribute typically 100-200 pg/mL of bovine TGF- β 2 upon activation. Prepare, activate, and assay an appropriate media control to determine the baseline bovine TGF- β 2 in the sample.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature. Centrifuge for 15 minutes at 1000 x g. Remove serum and assay (see activation procedure) immediately or aliquot and store unactivated samples at ≤ -70 °C. Avoid repeated freeze-thaw cycles.

Platelet-poor Plasma* - Collect plasma on ice using EDTA or heparin as an anticoagulant. Centrifuge at 2-8 °C for 15 minutes at 1000 x g within 30 minutes of collection. For more complete platelet removal, an additional centrifugation step of the separated plasma at 1500 x g for 10 minutes at 2-8 °C is recommended. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Note: *Grossly hemolyzed or lipemic samples are not suitable for the measurement of TGF- β in this assay.*

**TGF- β 1 is present in platelet granules and is released upon platelet activation. Therefore, to measure circulating levels of TGF- β 1, platelet-poor plasma should be collected for measurement. It should be noted that many protocols for plasma preparation, including procedures recommended by the Clinical and Laboratory Standards Institute (CLSI), result in incomplete removal of platelets from blood. This will cause variable and irreproducible results for assays of factors contained in platelets and released by platelet activation. The recommended plasma collection protocol is designed to minimize platelet degranulation. However, since even the best methods for plasma collection may result in some platelet degranulation on occasion, it is recommended that markers for platelet degranulation be determined in samples containing elevated TGF- β 1 levels.*

Urine - Aseptically collect the first urine of the day (mid-stream), voided directly into a sterile container. Centrifuge to remove particulate matter. Assay immediately (see activation procedure) or aliquot and store unactivated samples at ≤ -70 °C. Avoid repeated freeze/thaw cycles.

Note: *Neat unactivated urine samples exhibit a decrease in TGF- β 1 concentration in the first 24 hours of storage (frozen or refrigerated). Care should be taken that all samples are assayed under identical storage conditions and durations.*

Human Milk - Centrifuge for 15 minutes at 1000 x g at 2-8 °C. Collect the aqueous fraction and repeat this process a total of 3 times. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

ACTIVATION/NEUTRALIZATION REAGENT PREPARATION

The following solutions may be stored at room temperature for up to one month in polypropylene bottles.

Caution: *Wear protective clothing and safety glasses during preparation or use of these reagents. Refer to the appropriate SDS prior to use.*

1N HCl (100 mL) - Slowly add 8.33 mL of 12N HCl to 91.67 mL of deionized water. Mix well.

1.2N NaOH/0.5M HEPES (100 mL) - Slowly add 12 mL of 10N NaOH to 75 mL of deionized water. Mix well, and add 11.9 g of HEPES. Mix well, and bring the final volume to 100 mL with deionized water.

SAMPLE ACTIVATION PROCEDURE

To activate latent TGF- β to immunoreactive TGF- β detectable in this assay, follow the activation procedure outlined below for all sample types. Assay samples immediately after neutralization (pH 7.2-7.6). **Use polypropylene test tubes. Do not activate the TGF- β Standard Cocktail.**

1. If samples contain particulates, centrifuge for 2 minutes at 5000 x g before proceeding.
2. Add 20 μ L of 1N HCl to 100 μ L of sample. Mix well, and incubate for 10 minutes at room temperature.
3. Add 20 μ L of 1.2N NaOH/0.5M HEPES. Mix thoroughly.

SAMPLE PREPARATION

Use polypropylene tubes.

Note: *On the day of the assay, ALL fresh and previously frozen serum and plasma samples require centrifugation at 16,000 x g for 4 minutes immediately prior to use or dilution.*

Activated cell culture supernates and activated urine samples require a 1:3.57 dilution in Calibrator Diluent RD5-49 after activation. This is a 1:5 final dilution factor. A suggested 1:3.57 dilution is 112 μ L of activated/neutralized sample + 288 μ L of Calibrator Diluent RD5-49. Mix thoroughly.

Activated serum, plasma, and human milk samples require a 1:10.7 dilution in Calibrator Diluent RD6-50 after activation. This is a 1:15 final dilution factor. A suggested 1:10.7 dilution is 20 μ L of activated/neutralized sample + 194 μ L of Calibrator Diluent RD6-50. Mix thoroughly.

REAGENT PREPARATION

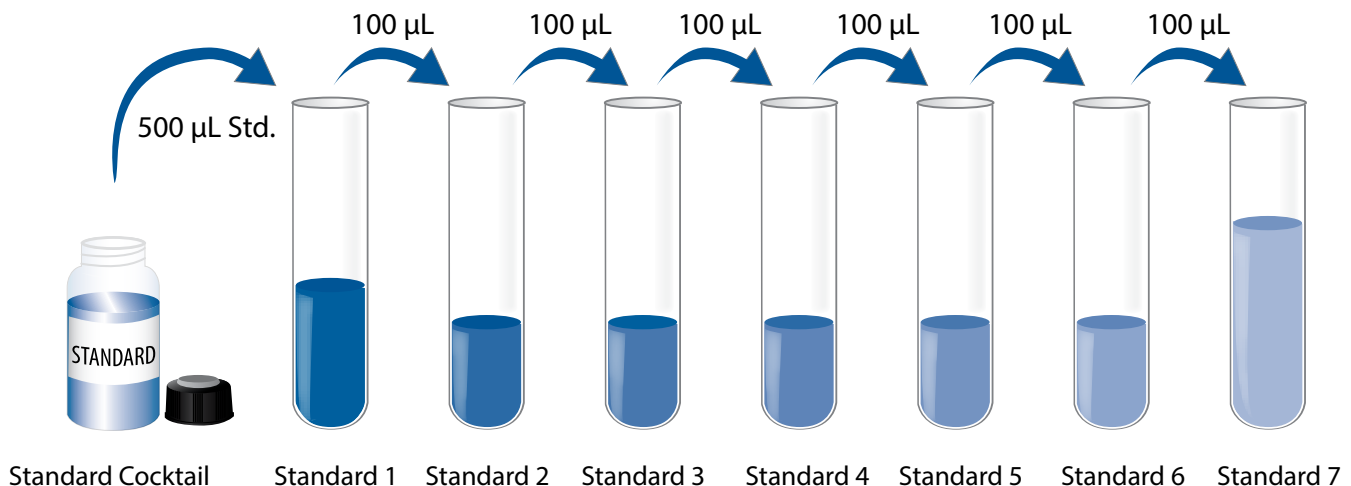
Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to 480 mL of deionized or distilled water to prepare 500 mL of Wash Buffer.

Standard - Refer to the Standard Value Card for the reconstitution volume and assigned values. Reconstitute the Standard Cocktail with Calibrator Diluent RD5-49 (*for cell culture supernate and urine samples*) or Calibrator Diluent RD6-50 (*for serum, plasma, and human milk samples*). Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

Note: Do NOT vortex standard cocktail. Gentle agitation should be initiated only after the 15-minute reconstitution step is complete.

Use polypropylene tubes. Pipette 500 μ L of the reconstituted Standard into the Standard 1 tube. Pipette 200 μ L of the appropriate calibrator diluent into the remaining tubes. Use Standard 1 to produce a 3-fold dilution series (below). Mix each tube thoroughly before the next transfer. Standard 1 serves as the high standard. The appropriate calibrator diluent serves as the blank.



DILUTED MICROPARTICLE COCKTAIL PREPARATION

1. Centrifuge the Microparticle Concentrate vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vial to resuspend the microparticles, taking precautions not to invert the vial.
3. Dilute the Microparticle Concentrate in the mixing bottle provided.

| Number of Wells Used | Microparticle Concentrate | + | Microparticle Diluent |
|----------------------|---------------------------|---|-----------------------|
| 96 | 500 µL | + | 5.00 mL |
| 72 | 375 µL | + | 3.75 mL |
| 48 | 250 µL | + | 2.50 mL |
| 24 | 125 µL | + | 1.25 mL |

Note: Protect microparticles from light during handling. Diluted microparticles cannot be stored. Prepare microparticles within 30 minutes of use.

DILUTED BIOTIN-ANTIBODY COCKTAIL PREPARATION

1. Centrifuge the Biotin-Antibody Concentrate vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vial, taking precautions not to invert the vial.
3. Dilute Biotin-Antibody Concentrate to Biotin Antibody Diluent 2. Mix gently.

| Number of Wells Used | Biotin-Antibody Concentrate | + | Biotin Antibody Diluent 2 |
|----------------------|-----------------------------|---|---------------------------|
| 96 | 500 µL | + | 5.00 mL |
| 72 | 375 µL | + | 3.75 mL |
| 48 | 250 µL | + | 2.50 mL |
| 24 | 125 µL | + | 1.25 mL |

STREPTAVIDIN-PE PREPARATION

Use a polypropylene amber bottle or a polypropylene tube wrapped with aluminum foil. Protect Streptavidin-PE from light during handling and storage.

1. Centrifuge the Streptavidin-PE vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vial, taking precautions not to invert the vial.
3. Dilute the Streptavidin-PE concentrate in Wash Buffer.

| Number of Wells Used | Streptavidin-PE Concentrate | + | Wash Buffer |
|----------------------|-----------------------------|---|-------------|
| 96 | 55.0 µL | + | 5.50 mL |
| 72 | 42.0 µL | + | 4.10 mL |
| 48 | 28.0 µL | + | 2.75 mL |
| 24 | 14.0 µL | + | 1.35 mL |

INSTRUMENT SETTINGS

Note: Adjust the probe height setting on the analyzer to avoid puncturing the plate. Calibrate the analyzer using the proper reagents for superparamagnetic microparticles (refer to instrument manual).

MAGPIX® analyzer:

- a) Sample volume: 50 µL
- b) Assign the microparticle region for each analyte being measured (see page 1)
- c) 50 count/region
- d) Collect Median Fluorescence Intensity (MFI)

Luminex® 100/200™, FLEXMAP 3D®, xMAP INTELLIFLEX®, and Bio-Rad® Bio-Plex® analyzers:

Note: Ensure that the instrument flow rate is set to the default of 60 µL/minute (fast) for all flow based analyzers.

- a) Sample volume: 50 µL
- b) Bead Type:
 - i. Luminex® 100/200™, FLEXMAP 3D®, and xMAP INTELLIFLEX select MagPlex
 - ii. Bio-Rad® Bio-Plex® Manager use Bio-Plex® MagPlex Beads (Magnetic)
- c) Doublet Discriminator gates:
 - i. Luminex® 100/200™ and FLEXMAP 3D®, set at 8000 and 16,500
 - ii. xMAP INTELLIFLEX® set at 7000 and 17,000
 - iii. Bio-Rad® Bio-Plex® Manager set at 8000 and 23,000
- d) Reporter Gain Setting:
 - i. Luminex® 100/200™ use Default setting
 - ii. FLEXMAP 3D® use Standard PMT setting
 - iii. xMAP INTELLIFLEX® use Luminex® 200™ Operating Mode on Low PMT setting
 - iv. Bio-Rad® Bio-Plex® Manager use the low RP1 target value for the CAL2 setting
- e) Assign the microparticle region for each analyte being measured (see Certificate of Analysis)
- f) 50 count/region
- g) Collect MFI

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that all standards, controls, and samples be assayed in duplicate.

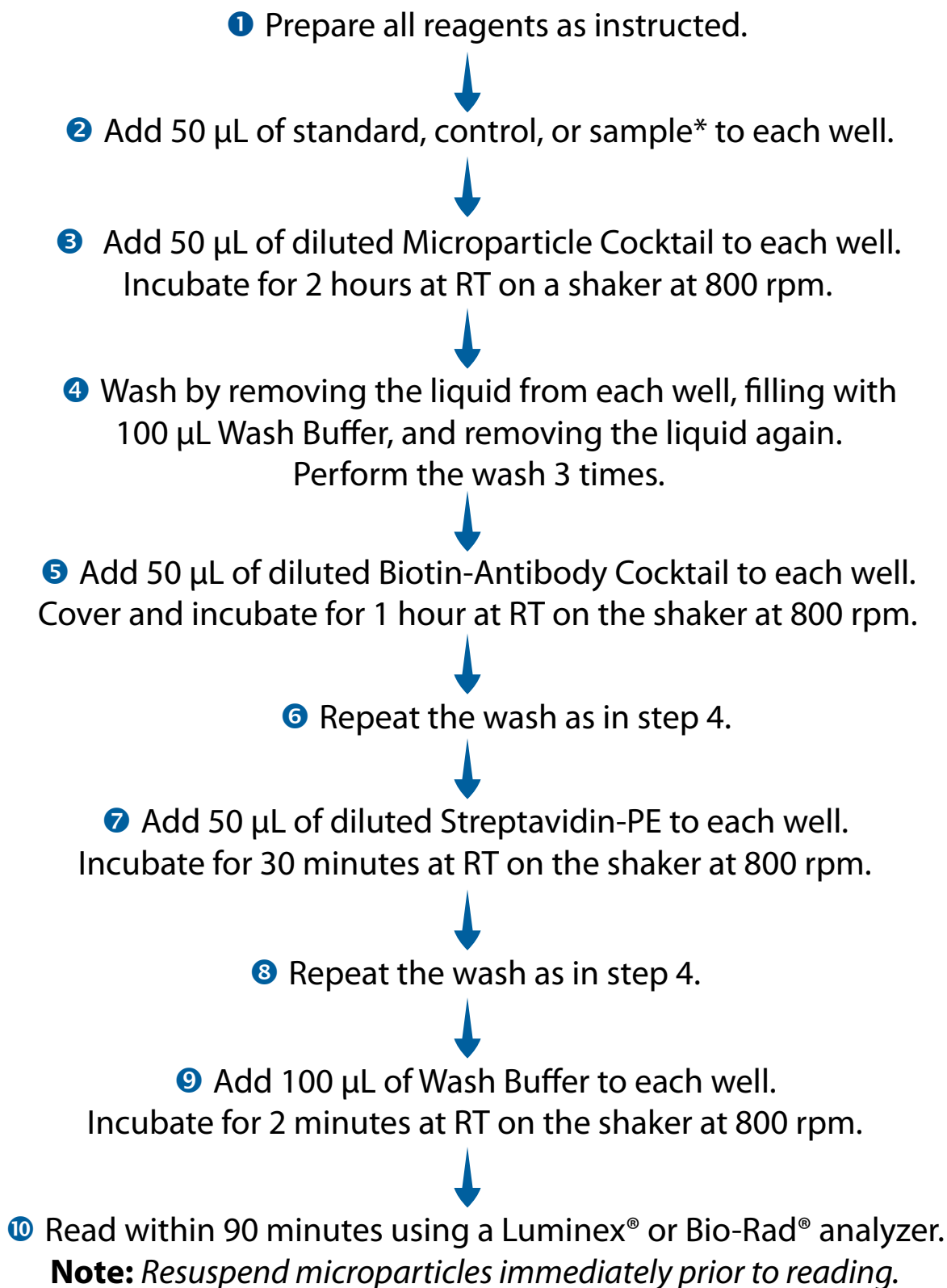
Note: *Protect microparticles and Streptavidin-PE from light at all times.*

1. Prepare all reagents, working standards, and samples as directed in the previous sections.
2. Add 50 μL of standard, control, or sample* per well. A plate layout is provided to record standards and samples assayed.
3. Resuspend the diluted Microparticle Cocktail by inversion or vortexing. Add 50 μL of the microparticle cocktail to each well of the microplate. Securely cover with a foil plate sealer. Incubate for 2 hours at room temperature on a horizontal orbital microplate shaker (0.12" orbit) set at 800 ± 50 rpm.
4. Using a magnetic device designed to accommodate a microplate, wash by applying the magnet to the bottom of the microplate, allow 1 minute before removing the liquid, filling each well with Wash Buffer (100 μL) and allow 1 minute before removing the liquid again. Uniform removal of liquid is essential for good performance. **Note: Do NOT blot; this may cause a loss of microparticles.** Perform the wash procedure three times.
5. Add 50 μL of diluted Biotin-Antibody Cocktail to each well. Securely cover with a foil plate sealer and incubate for 1 hour at room temperature on the shaker set at 800 ± 50 rpm.
6. Repeat the wash as in step 4.
7. Add 50 μL of Streptavidin-PE to each well. Securely cover with a foil plate sealer and incubate for 30 minutes at room temperature on the shaker set at 800 ± 50 rpm.
8. Repeat the wash as in step 4.
9. Resuspend the microparticles by adding 100 μL of Wash Buffer to each well. Incubate for 2 minutes at room temperature on the shaker set at 800 ± 50 rpm.
10. Read within 90 minutes using a Luminex® or Bio-Rad® Analyzer.
Note: *Resuspend microparticles immediately prior to reading by shaking the plate for 2 minutes on the plate shaker set at 800 ± 50 rpm.*

*Samples require activation and dilution. See Sample Activation and Sample Preparation sections.

ASSAY PROCEDURE SUMMARY

Note: Protect microparticles and Streptavidin-PE from light at all times.



*Samples require activation or dilution.

See Sample Activation and Sample Preparation sections.

CALCULATION OF RESULTS

Use the Standard concentrations on the Standard Value Card and calculate 3-fold dilutions for the remaining levels. Average the duplicate readings for each standard and sample and subtract the average blank Median Fluorescence Intensity (MFI).

Create a standard curve for each analyte by reducing the data using computer software, such as [Bio-Techne® Quantist™](#), capable of generating a five parameter logistic (5-PL) curve-fit.

Since samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

CALIBRATION

This assay is calibrated against highly purified recombinant human TGF- β 1, TGF- β 2, and TGF- β 3 produced at R&D Systems®.

PRECISION

Intra-Assay Precision - Generated from the mean of the %CV's from 20 reportable results across three different concentrations of analytes in a single serum assay.

Inter-Assay Precision - Generated from the mean of the %CV's across three different concentrations of analytes across 20 different serum assays.

| Analyte | Intra-Assay (%CV) | Inter-Assay (%CV) |
|----------------|-------------------|-------------------|
| TGF- β 1 | 7.3 | 13.0 |
| TGF- β 2 | 6.4 | 9.3 |
| TGF- β 3 | 7.1 | 8.3 |

ACCURACY

Linearity - The data represents mean spiked linearity in serum matrix samples.

Spiked Recovery - The data represents mean percent recovery of spiked standards ranging from low, medium, and high concentration in serum matrix samples.

| Analyte | n | Linearity Average % | n | Recovery Average % |
|----------------|---|---------------------|---|--------------------|
| TGF- β 1 | 4 | 97 | 4 | 97 |
| TGF- β 2 | 4 | 93 | 4 | 103 |
| TGF- β 3 | 4 | 108 | 4 | 76 |

SPECIFICITY

This assay recognizes natural and recombinant TGF- β s.

The factors listed below were prepared at 50 ng/mL in calibrator diluent, assayed, and measured less than 0.5% cross-reactivity unless otherwise noted. The same factors prepared at 50 ng/mL in a mid-range recombinant TGF- β control were assayed for interference. No interference was observed except as noted.

Recombinant human:

| | |
|----------------|------------------|
| 6Ckine | HCC-1 |
| Activin A | HCC-4 |
| Activin RIA | I-309 |
| Activin RIIA | IL-8 |
| Activin RII B | Inhibin A |
| BLC/BCA-1 | Inhibin B |
| BMP-2 | IP-10 |
| BMP-3 | I-TAC |
| BMP-3b | LAP |
| BMP-4 | MCP-1 |
| BMP-5 | MCP-2 |
| BMP-6 | MCP-3 |
| BMP-8b | MCP-4 |
| BMP-10 | MDC |
| BMP-15 | MIG |
| BMP RIA | MIP-1 α |
| BMP RIB | MIP-1 β |
| BMP RII | MIP-1 δ |
| CTACK | MIP-3 α |
| ENA-78 | MIP-3 β |
| Eotaxin | MPIF-1 |
| Eotaxin-2 | NAP-2 |
| Eotaxin-3 | PARC |
| Follistatin288 | RANTES |
| Follistatin300 | SDF-1 α |
| Follistatin315 | SDF-1 β |
| GCP-2 | TECK |
| GRO α | TGF- α |
| GRO β | TGF- β RII |
| GRO γ | |

Recombinant mouse:

| |
|----------------|
| 6Ckine |
| BLC/BCA-1 |
| BMP-3b |
| BMP RIA |
| BMP RIB |
| CRG-2/IP-10 |
| CTACK |
| Eotaxin |
| GCP-2 |
| JE/MCP-1 |
| KC |
| MARC |
| MCP-5 |
| MDC |
| MIG |
| MIP-1 α |
| MIP-1 β |
| MIP-1 γ |
| RANTES |
| SDF-1 α |
| TARC |
| TECK |

Recombinant rat:

| |
|----------------|
| Agrin |
| MIP-3 α |

Other recombinants:

| |
|--------------------------|
| porcine IL-8 |
| amphibian TGF- β 5 |

Recombinant human TGF- β 1.2 exhibits 4.5% cross-reactivity in this assay.

Recombinant human TGF- β RIII interferes at concentrations \geq 50 ng/mL.

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PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

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NOTES

NOTES

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