# Luminex<sup>®</sup> Performance Assay

## **Mouse Cytokine Premixed Kit**

Catalog Number FCST05

For the simultaneous quantitative determination of multiple mouse cytokine concentrations in cell culture supernates, serum, and plasma.

This package insert must be read in its entirety before using this product. For research use only. Not for use in diagnostic procedures.

#### **TABLE OF CONTENTS**

#### **SECTION**

#### PAGE

INTRODUCTION	.1
PRINCIPLE OF THE ASSAY	.2
LIMITATIONS OF THE PROCEDURE	.2
TECHNICAL HINTS	.2
MATERIALS PROVIDED & STORAGE CONDITIONS	.3
OTHER SUPPLIES REQUIRED	.4
PRECAUTIONS	.4
SAMPLE COLLECTION & STORAGE	.4
SAMPLE PREPARATION	.4
REAGENT PREPARATION	.5
DILUTED MICROPARTICLE COCKTAIL PREPARATION	.6
DILUTED BIOTIN ANTIBODY COCKTAIL PREPARATION	.6
STREPTAVIDIN-PE PREPARATION	.6
INSTRUMENT SETTINGS	.7
ASSAY PROCEDURE	.8
CALCULATION OF RESULTS	.9
CALIBRATION	9
PLATE LAYOUT	0

#### MANUFACTURED AND DISTRIBUTED BY:

#### USA & Canada | R&D Systems, Inc.

614 McKinley Place NE, Minneapolis, MN 55413, USA TEL: (800) 343-7475 (612) 379-2956 FAX: (612) 656-4400 E-MAIL: info@RnDSystems.com

#### **DISTRIBUTED BY:**

#### UK & Europe | R&D Systems Europe, Ltd.

19 Barton Lane, Abingdon Science Park, Abingdon OX14 3NB, UK TEL: +44 (0)1235 529449 FAX: +44 (0)1235 533420 E-MAIL: info@RnDSystems.co.uk

#### China | R&D Systems China Co., Ltd.

24A1 Hua Min Empire Plaza, 726 West Yan An Road, Shanghai PRC 200050 TEL: +86 (21) 52380373 FAX: +86 (21) 52371001 E-MAIL: info@RnDSystemsChina.com.cn

#### **INTRODUCTION**

Cytokines are intercellular signaling proteins released from a wide variety of cells and tissues. They play an integral role in regulating growth and cellular proliferation as well as modulating host response to infection, injury, and inflammation. Cytokines also influence reproduction and bone remodeling. A large number of cytokines are pleiotropic and share similar functions. In addition, many cytokines influence the production of other cytokines. Analysis and quantification of cytokines in biological fluids and cell culture supernates has thus become increasingly important. Methods such as bioassay, enzyme-linked immunosorbent assay (ELISA), intracellular staining, ribonuclease protection assay (RPA) and polymerase chain reaction (PCR) have all been used for quantifying cytokines; however, each of these techniques has limitations associated with it. These techniques are not capable of measuring multiple cytokines simultaneously in a limited sample volume.

This kit can be used to simultaneously assess the levels of up to 10 cytokines from the table below in a single sample. For ease of use, the microparticles and the biotinylated detection antibodies are premixed in respective vials.

Analyte	Microparticle Region	Performance Data Online (www.RnDSystems.com/pdf/)
CCL2/JE/MCP-1	30	LUM479.pdf
GM-CSF	11	LUM415B.pdf
IFN-γ	75	LUM485.pdf
IL-1β/IL-1F2	06	LUM401.pdf
IL-2	17	LUM402.pdf
IL-4	21	LUM404.pdf
IL-5	54	LUM405.pdf
IL-6	32	LUM406.pdf
IL-10	50	LUM417.pdf
IL-12 p70	66	LUM419.pdf
IL-13	36	LUM413.pdf
IL-17	39	LUM421.pdf
CXCL1/KC	26	LUM453.pdf
CXCL2/MIP-2	47	LUM452.pdf
TNF-a	79	LUM410.pdf
VEGF	43	LUM493.pdf

#### **PRINCIPLE OF THE ASSAY**

Luminex<sup>®</sup> Performance Assay multiplex kits are designed for use with the Luminex 100<sup>™</sup>, Luminex 200<sup>™</sup>, or Bio-Rad<sup>®</sup> Bio-Plex<sup>®</sup> dual laser, flow-based sorting and detection analyzers.

Analyte-specific antibodies are pre-coated onto color-coded microparticles. Microparticles, standards and samples are pipetted into wells and the immobilized antibodies bind the analytes of interest. After washing away any unbound substances, a biotinylated antibody cocktail specific to the analytes of interest is added to each well. Following a wash to remove any unbound biotinylated antibody, streptavidin-phycoerythrin (Streptavidin-PE), which binds to the biotinylated detection antibodies, is added to each well. A final wash removes unbound Streptavidin-PE. The microparticles are resuspended in buffer and read using the Luminex or Bio-Plex analyzer. One laser is microparticle-specific and determines which analyte is being detected. The other laser determines the magnitude of the phycoerythrin-derived signal, which is in direct proportion to the amount of analyte bound.

#### LIMITATIONS OF THE PROCEDURE

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples generate values higher than the highest standard, dilute the samples with the appropriate Calibrator Diluent and repeat the assay.
- Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- Variations in sample collection, processing, and storage may cause sample value differences.
- This assay is designed to eliminate interference by other factors present in biological samples. However, until these factors have been tested in the Luminex Performance Assay, the possibility of interference cannot be excluded.
- Luminex Performance Assays afford the user the benefit of multianalyte analysis of cytokines in a single complex sample. For each sample type, a single multipurpose diluent is used to optimize recovery, linearity, and reproducibility. Such a multipurpose diluent may not optimize any single analyte to the same degree that a unique diluent selected for analysis of that analyte can optimize conditions. Therefore, some performance characteristics may be more variable than those for assays designed specifically for single analyte analysis.
- Only the analytes listed on the Standard Value card can be measured with this kit. Refer to the enclosed certificate of analysis for specific analytes included in this kit.

## **TECHNICAL HINTS**

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Protect microparticles and Streptavidin-PE from light at all times to prevent photobleaching.
- For best results, adjust the vacuum strength on the plate washer to between 15 and 40 cm of mercury.

## **MATERIALS PROVIDED & STORAGE CONDITIONS**

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

PART	PART #	DESCRIPTION	STORAGE OF OPENED, DILUTED, OR RECONSTITUTED MATERIAL		
Standard Cocktail 1	895814	2 vials of recombinant mouse cytokines in a buffered protein base with preservatives; lyophilized.	Discard after use. Use fresh standards for each assay.		
Mouse Cytokine Premixed Kit Microparticle Cocktail	894107	0.6 mL of a concentrated microparticle cocktail with preservatives.	May be stored for up to 1 month at 2-8 °C.* Once diluted, any unused microparticle cocktail must be discarded.		
Mouse Cytokine Premixed Kit Biotin Antibody Cocktail	894106	0.6 mL of a concentrated biotin antibody cocktail with preservatives.			
Microparticle Diluent 2	895815	6 mL of a buffered protein base with preservative.			
Biotin Antibody Diluent	895816	5.5 mL of a buffered protein base with preservative.			
Calibrator Diluent RD5K	895119	21 mL of a 2-fold concentrated solution of a buffered protein base with preservatives. <i>For cell culture supernate</i> <i>samples. Used diluted 1:2 in this assay.</i>			
Calibrator Diluent RD6-40	895817	21 mL of a buffered protein base with preservatives. <i>For serum/plasma</i> <i>samples. May contain a precipitate. Mix</i> <i>well before and during use.</i>	May be stored for up to 1 month at 2-8 °C.*		
Streptavidin-PE	892525	0.07 mL of a 100-fold concentrated streptavidin-phycoerythrin conjugate with preservatives.			
Wash Buffer Concentrate	895003	21 mL of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>			
Microplate	640763	1 filter-bottomed 96-well microplate used as a vessel for the assay.			
Mixing Bottles	895505	2 empty 8 mL bottles used for mixing microparticles with Microparticle Diluent.			
Plate Sealers	640445	4 adhesive foil strips.			
Standard Value Card	750601	1 card listing the Standard Cocktail 1 reconstitution volume and working standard concentrations for this lot of kit.			

\*Provided this is within the expiration date of the kit.

## **OTHER SUPPLIES REQUIRED**

- Luminex 100, Luminex 200, or Bio-Rad Bio-Plex analyzer with X-Y platform.
- Microplate vacuum manifold (Millipore Multiscreen<sup>™</sup> Vacuum Manifold Catalog # MAVM096 or equivalent).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Multi-channel pipette, manifold dispenser, or automated dispensing unit.
- 50 mL and 500 mL graduated cylinders.
- Horizontal orbital microplate shaker (0.12" orbit) capable of maintaining a speed of  $500 \pm 50$  rpm.
- Microcentrifuge.
- Polypropylene test tubes for dilution of standards and samples.

## PRECAUTIONS

The Biotin Antibody Diluent in this kit contains sodium azide, which may react with lead and copper plumbing to form explosive metallic azides. Flush with large volumes of water during disposal.

Some components in this kit contain ProClin<sup>®</sup> which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Please refer to the MSDS on our website prior to use.

## **SAMPLE COLLECTION & STORAGE**

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

**Cell Culture Supernates** - Remove particulates by centrifugation and assay immediately or aliquot and store samples at  $\leq$  -20 °C. Avoid repeated freeze-thaw cycles.

**Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at  $\leq$  -20 °C. Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at  $\leq$  -20 °C. Avoid repeated freeze-thaw cycles.

**Note:** Citrate plasma has not been validated for use in this assay.

## SAMPLE PREPARATION

Serum and plasma samples require a 4-fold dilution. A suggested 4-fold dilution is 30  $\mu$ L of sample + 90  $\mu$ L of Calibrator Diluent RD6-40. Mix thoroughly.

#### **REAGENT PREPARATION**

#### Bring all reagents to room temperature before use.

**Wash Buffer** - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to deionized or distilled water to prepare 500 mL of Wash Buffer.

**Calibrator Diluent RD5K (diluted 1:2)** - Add 20 mL of Calibrator Diluent RD5K concentrate to 20 mL of deionized or distilled water to prepare 40 mL of Calibrator Diluent RD5K (diluted 1:2).

**Standard** - Reconstitute Standard Cocktail 1 with Calibrator Diluent RD5K (diluted 1:2) (*for cell culture supernate samples*) or Calibrator Diluent RD6-40 (*for serum/plasma samples*). Refer to the Standard Value Card for the reconstitution volume and the assigned values of Standard 1. Allow the standards to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

**Use polypropylene tubes.** Pipette 300  $\mu$ L of reconstituted Standard Cocktail 1 into a tube labeled Standard 1. Pipette 200  $\mu$ L of the appropriate Calibrator Diluent into the remaining tubes. Use Standard 1 to produce a 3-fold dilution series (below). Mix each tube thoroughly before the next transfer. Standard 1 serves as the high standard. The appropriate Calibrator Diluent serves as the blank.



## **DILUTED MICROPARTICLE COCKTAIL PREPARATION**

- 1. Centrifuge the Microparticle Cocktail vial for 30 seconds at 1000 x g prior to removing the cap.
- 2. Gently vortex the vial to resuspend the microparticles, taking precautions not to invert the vial.
- 3. Dilute the Microparticle Cocktail in the mixing bottle provided.

Number of Wells Used	Microparticle Cocktail	+	Microparticle Diluent 2
96	500 μL	+	5.00 mL
72	375 μL	+	3.75 mL
48	250 μL	+	2.50 mL
24	125 μL	+	1.25 mL

**Note:** *Protect microparticles from light during handling. Diluted microparticles cannot be stored. Prepare microparticles within 30 minutes of use.* 

## **DILUTED BIOTIN ANTIBODY COCKTAIL PREPARATION**

- 1. Centrifuge the Biotin Antibody Cocktail vial for 30 seconds at 1000 x g prior to removing the cap.
- 2. Gently vortex the vial, taking precautions not to invert the vial.
- 3. Dilute the Biotin Antibody Cocktail in Biotin Antibody Diluent. Mix gently.

Number of Wells Used	Biotin Antibody Cocktail	+	<b>Biotin Antibody Diluent</b>
96	500 μL	+	5.00 mL
72	375 μL	+	3.75 mL
48	250 μL	+	2.50 mL
24	125 μL	+	1.25 mL

#### **STREPTAVIDIN-PE PREPARATION**

#### Use a polypropylene amber bottle or a polypropylene tube wrapped with aluminum foil. Protect Streptavidin-PE from light during handling and storage.

- 1. Centrifuge the Streptavidin-PE vial for 30 seconds at 1000 x g prior to removing the cap.
- 2. Gently vortex the vial, taking precautions not to invert the vial.
- 3. Dilute the 100X Streptavidin-PE to a 1X concentration by adding 55  $\mu$ L of Streptavidin-PE to 5.5 mL of Wash Buffer.

#### **INSTRUMENT SETTINGS**

Adjust the probe height setting on the Luminex analyzer to avoid puncturing the membrane. Refer to the instrument manual.

- a) Assign the bead region for each analyte being measured
- b) 50 events/bead
- c) Minimum events: 0
- d) Flow rate: 60  $\mu$ L/minute (fast)
- e) Sample size: 50 µL
- f) Doublet Discriminator gates at approximately 7500 and 15,500
- g) Collect Median Fluorescence Intensity (MFI)

**Note:** For the Bio-Rad Bio-Plex analyzer, set the gates at 4300 and 10,000. The CAL2 setting for the Bio-Rad Bio-Plex analyzer should be set at the low RP1 target value.

#### **ASSAY PROCEDURE**

## Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.

#### Note: Protect microparticles and Streptavidin-PE from light at all times.

- 1. Prepare all reagents, working standards, and samples as directed in the previous sections.
- 2. Pre-wet the filter-bottomed microplate by filling each well with 100  $\mu$ L of Wash Buffer. Remove the liquid through the filter at the bottom of the plate using a vacuum manifold designed to accommodate a microplate.

**Note:** After each final wash cycle and subsequent reagent addition, blot the bottom of the microplate with a paper towel to prevent wicking.

- 3. Resuspend the diluted Microparticle Cocktail by inversion or vortexing. Add 50 µL of the microparticle mixture to each well of the pre-wet filter-bottomed microplate.
- 4. Add 50  $\mu$ L of Standard or sample\* per well. Pipette the assay within 20 minutes. Securely cover with a foil plate sealer. Incubate for 3 hours at room temperature on a horizontal orbital microplate shaker (0.12" orbit) set at 500 ± 50 rpm. A plate layout is provided to record standards and samples assayed.
- 5. Using a vacuum manifold designed to accommodate a microplate, wash by removing the liquid, filling each well with Wash Buffer (100  $\mu$ L), and removing the liquid again. All of the liquid must be removed through the filter at the bottom of the microplate to avoid any loss of microparticles. Complete removal of liquid is essential for good performance. Perform the wash procedure three times.
- 6. Add 50  $\mu$ L of diluted Biotin Antibody Cocktail to each well. Securely cover with a new foil plate sealer, and incubate for 1 hour at room temperature on the shaker set at 500  $\pm$  50 rpm.
- 7. Repeat the wash as in step 5.
- 8. Add 50  $\mu$ L of diluted Streptavidin-PE to each well. Securely cover with a new foil plate sealer, and incubate for 30 minutes at room temperature on the shaker set at 500 ± 50 rpm.
- 9. Repeat the wash as in step 5.
- 10. Resuspend the microparticles by adding 100  $\mu$ L of Wash Buffer to each well. Incubate for 2 minutes at room temperature on the shaker set at 500 ± 50 rpm.
- 11. Read within 90 minutes using the Luminex or Bio-Rad analyzer.

\*Serum/plasma samples require dilution. See the Sample Preparation section.

#### **CALCULATION OF RESULTS**

Use the Standard concentrations on the Standard Value Card and calculate 3-fold dilutions for the remaining levels. Average the duplicate readings for each standard and sample and subtract the average blank Median Fluorescence Intensity (MFI).

Create a standard curve for each analyte by reducing the data using computer software capable of generating a five parameter logistic (5-PL) curve-fit.

Since samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

#### **CALIBRATION**

This assay is calibrated against highly purified recombinant mouse cytokines produced at R&D Systems.

All trademarks and registered trademarks are the property of their respective owners.

#### **PLATE LAYOUT**

Use this plate layout to record standards and samples assayed.



©2013 R&D Systems, Inc.