

DESCRIPTION

Species Reactivity	Human
Specificity	Detects human Chemerin in ELISAs. In ELISAs, this antibody does not cross-react with recombinant mouse Chemerin.
Source	Monoclonal Mouse IgG _{2B} Clone # 365314
Purification	Protein A or G purified from hybridoma culture supernatant
Immunogen	<i>E. coli</i> -derived recombinant human Chemerin Glu21-Ser157 Accession # Q99969
Conjugate	Alexa Fluor 532 Excitation Wavelength: 534 nm Emission Wavelength: 553 nm
Formulation	Supplied 0.2mg/ml in 1X PBS with RDF1 and 0.09% Sodium Azide
*Contains <0.1% Sodium Azide, which is not hazardous at this concentration according to GHS classifications. Refer to the Safety Data Sheet (SDS) for additional information and handling instructions.	

APPLICATIONS

Please Note: Optimal dilutions should be determined by each laboratory for each application. [General Protocols](#) are available in the Technical Information section on our website.

ELISA Capture (Matched Antibody Pair)	Optimal dilution of this antibody should be experimentally determined.
ELISA Detection (Matched Antibody Pair)	Optimal dilution of this antibody should be experimentally determined.

PREPARATION AND STORAGE

Shipping	The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below.
Stability & Storage	Protect from light. Do not freeze. 12 months from date of receipt, 2 to 8 °C as supplied

BACKGROUND

Human Chemerin, also known as Tazarotene-induced Gene 2, (TIG2) is a new, but distant member of the Cystatin superfamily (1-3). Members of this superfamily contain at least two intrachain disulfide bonds and an α -helical structure over a distance of about 100 amino acids (2, 3). Chemerin is synthesized as a 163 aa precursor that contains a hydrophobic 20 aa N-terminal sequence, an intervening 137 aa Cystatin-fold containing domain, and a six aa C-terminal prosegment (1, 4). Within the cystatin-fold domain there are three intrachain disulfide bonds that contribute to the fold, and three potential sites for phosphorylation and one for myristoylation (5). The precursor molecule undergoes proteolytic processing at both termini by unknown proteases. The N-terminal residue 20 aa hydrophobic segment is described as being either a signal sequence or a transmembrane (TM) segment for a type II TM protein (1, 6). In either case, it gives rise to a soluble proform that undergoes further processing at the C-terminus. In human, the C-terminal six residues are cleaved, giving rise to a monomeric, 16 kDa heparin-binding bioactive molecule (aa 21-157) (7). A shorter 134 aa form has been described (5). Bioactivity seems to be concentrated in the nine residues preceding the prosegment (aa 149-157). Retention of the prosegment blocks activity (4). The 137 aa mature segment is known to bind to the G-protein coupled receptor termed ChemR23 (5, 7). Binding results in macrophage and immature dendritic cell chemotaxis (7). The distribution of this receptor is limited to immune APCs, and it is assumed that Chemerin is an inflammatory molecule. It is unclear which cells are actually producing Chemerin, but keratinocytes, endothelial cells and osteoclasts are potential candidates (1, 7). Mature human Chemerin shares 67% aa sequence identity with mouse Chemerin (7). There is apparently cross-species activity for the protein (8).

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