CALCULATION OF RESULTS
Average the duplicate readings for each standard and sample, then subtract the average zero standard optical density (O.D.). Results may be normalized to total protein or cell number.

Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the phospho-p38α (T180/Y182) concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

TYPICAL DATA
A standard curve should be generated for each set of samples assayed. The graph below represents typical data generated when using this Human/Mouse/Rat Phospho-p38α (T180/Y182) DuoSet® IC ELISA. The standard curve was calculated using a computer generated 4-PL curve-fit. This standard curve is for demonstration purposes only.

CALIBRATION
The Human/Mouse/Rat Phospho-p38α (T180/Y182) DuoSet® IC ELISA is calibrated against a highly purified E. coli-expressed recombinant human phospho-p38α (T180/Y182) produced at R&D Systems®. Samples containing natural phospho-p38α (T180/Y182) showed linear dilution parallel to the standard curve obtained using the Human/Mouse/Rat Phospho-p38α (T180/Y182) Standard. These results indicate that O.D. values from this DuoSet® IC ELISA can be used to determine the relative concentration of Phospho-p38α (T180/Y182) in natural samples.

SPECIFICITY
The Human/Mouse/Rat Phospho-p38α (T180/Y182) DuoSet® IC ELISA specifically recognizes p38α phosphorylated at T180/Y182. Specificity was demonstrated by Western Blot analysis of the protein bound by the capture antibody coated on the plate and cross-reactivity analysis.

TECHNICAL HINTS & LIMITATIONS
- This DuoSet® IC ELISA should not be used beyond the expiration date on the kit label.
- Individual results may vary due to differences in technique, plasticware, and water sources.
- It is important that the diluents selected for reconstitution and for dilution of the standard and sample reflect the environment of the samples being measured. The diluents suggested in this protocol should be suitable for most cell lysates.
- The type of enzyme and substrate and the concentrations of capture/detection antibodies used can be varied to create an immunooassay with a different sensitivity and dynamic range. A basic understanding of immunooassay development is required for the successful use of these reagents in immunooassays.
- A thorough and consistent wash technique is essential for proper assay performance. Wash Buffer should be dispensed forcefully and removed completely from the wells by aspiration or decanting. Remove any remaining Wash Buffer by inverting the plate and blotting it against clean paper towels.
- Use a fresh reagent reservoir and pipette tips for each step.
- It is recommended that all standards and samples be assayed in duplicate.
- Avoid microbial contamination of reagents and buffers. This may interfere with the sensitivity of the assay. Buffers containing protein should be made under aseptic conditions and stored at 2-8 °C or be prepared fresh daily.

PRECAUTIONS
The Stop Solution recommended for use with this kit is an acid solution. Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist. Color Reagent B recommended for use with this kit may cause skin, eye, and respiratory irritation. Avoid breathing fumes. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

INTENDED USE
For the development of sandwich ELISAs to measure p38α phosphorylated at T180/Y182 in cell lysates.

PRINCIPLE OF THE ASSAY
This DuoSet® IC ELISA contains the basic components required for the development of sandwich ELISAs to measure p38α phosphorylated at T180/Y182 in cell lysates. An immobilized capture antibody specific for human/mouse/rat p38α binds both phosphorylated and unphosphorylated p38α. After washing away unbound material, a biotinylated detection antibody specific for human/mouse/rat p38α dually phosphorylated at T180 and Y182 is used to detect captured protein, utilizing a standard Streptavidin-HRP format.

This package insert must be read in its entirety before using this product. For research use only. Not for use in diagnostic procedures.

Note: The reconstitution method has changed. Read this package insert in its entirety before using this product.
SOLUIONS REQUIRED

**PBS** - 137 mM NaCl, 2.7 mM KCl, 8.1 mM Na$_2$HPO$_4$, 1.5 mM KH$_2$PO$_4$, pH 7.2-7.4, 0.2 μM filtered (R&D Systems*, Catalog DY066).

**Wash Buffer** - 0.05% Tween® 20 in PBS, pH 7.2-7.4 (R&D Systems®, Catalog #WA126).

**Block Buffer** - 1% BSA, 0.05% Na$_2$PO$_4$ in PBS, pH 7.2-7.4.

**IC Diluent #1** - 1% BSA* in PBS, pH 7.2-7.4, 0.2 μM filtered.

**IC Diluent #8** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF in PBS, pH 7.2-7.4.

**Note:** IC Diluent #8 is also the base diluent for IC Diluent #3, IC Diluent #7, and Lysis Buffer #6. Approximately 50 mL of this diluent is required to run the assay on one 96 well plate.

**IC Diluent #3** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 1 M Urea in PBS, pH 7.2-7.4.

**IC Diluent #7** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M Urea in PBS, pH 7.2-7.4.

**Lysis Buffer #6** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M Urea, 1 mM activated Sodium Orthovanadate, 2.5 mM Sodium Pyrophosphate, 10 μg/mL Leupeptin, 10 μg/mL Pepstatin, 100 μM PMSF, and 3.0 μg/mL Aprotinin in PBS, pH 7.2-7.4.

**Substrate Solution** - 1:1 mixture of Color Reagent A (H$_2$O$_2$) and Color Reagent B (Tetramethylbenzidine) (R&D Systems*, Catalog #DY999).

**Stop Solution** - 2 N H$_2$SO$_4$, (R&D Systems*, Catalog #DY994).

**PREPARATION OF SAMPLES

**Cell Lysates** - Rinse cells twice with PBS, making sure to remove any remaining PBS after the second rinse. Solubilize cells at 1 x 10$^6$ cells/mL in Lysis Buffer #6 and allow samples to sit on ice for 15 minutes. Assay immediately or store at ≤ -70 °C. Before use, centrifuge samples at 2000 x g for 5 minutes and transfer the supernate to a clean test tube. Sample protein concentration may be quantified using a total protein assay. For assaying, dilute lysates 6-fold with IC Diluent #8 and make further serial dilutions in IC Diluent #3.

**Note:** The final concentration of urea in all samples and standards should be 1 M prior to addition to the plate.

**GENERAL ELISA PROTOCOL

**Plate Preparation**

1. Dilute the capture antibody to a working concentration of 6.0 μg/mL in PBS, without carrier protein. Immediately coat a 96 well microplate with 100 μL per well of the diluted capture antibody. Seal the plate and incubate overnight at room temperature.

2. Aspirate each well and wash with Wash Buffer, repeating the process two times for a total of 3 washes. Wash by filling each well with Wash Buffer (400 μL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.

3. Block plates by adding 300 μL of Block Buffer to each well. Incubate at room temperature for 1-2 hours.

4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

**Assay Procedure**

1. Add 100 μL of sample or standard in IC Diluent #3 per well. Use IC Diluent #3 as the zero standard. Cover with a plate sealer and incubate 2 hours at room temperature.

2. Repeat the aspiration/wash as in step 2 of the Plate Preparation.

3. Add 100 μL of the diluted detection antibody to each well. Cover with a new plate sealer and incubate 2 hours at room temperature.

4. Repeat the aspiration/wash as in step 2 of the Plate Preparation.

5. Add 100 μL of the diluted Streptavidin-HRP A to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.

6. Repeat the aspiration/wash as in step 2 of the Plate Preparation.

7. Add 100 μL of Substrate Solution to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.

8. Add 50 μL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.

9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.