

DuoSet[®] IC

Human Total SMAC/Diablo

Catalog Number DYC789-2

DYC789-5

For the development of sandwich ELISAs to measure SMAC/Diablo in cell lysates.

This package insert must be read in its entirety before using this product.

**FOR RESEARCH USE ONLY.
NOT FOR USE IN DIAGNOSTIC PROCEDURES.**

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PRINCIPLE OF THE ASSAY

This DuoSet[®] IC assay kit contains the basic components required for the development of sandwich ELISAs to measure SMAC/Diablo in cell lysates. An immobilized capture antibody specifically binds SMAC/Diablo. After washing away unbound material, a biotinylated detection antibody specific for SMAC/Diablo is used to detect the protein, utilizing a standard Streptavidin-HRP format.

MATERIALS PROVIDED

Store the unopened kit at 2 - 8° C. Do not use past kit expiration date.

Description	Part #	Storage Conditions	Vials Provided	
			Cat. # DYC789-2	Cat. # DYC789-5
Total SMAC/Diablo Capture Antibody	840761	2 - 8° C	1	2
Total SMAC/Diablo Detection Antibody	840762	2 - 8° C	1	2
Total SMAC/Diablo Standard	840763	2 - 8° C	3	5
Streptavidin-HRP	890803	2 - 8° C	1	1

DYC789-2 contains sufficient materials to run ELISAs on at least two 96 well plates.*
DYC789-5 contains sufficient materials to run ELISAs on at least five 96 well plates.*

*Provided the following conditions are met:

- The reagents are prepared as described in this package insert.
- The assay is run as described in the General ELISA Protocol on page 5.
- The recommended microplates, buffers, diluents, substrates, and solutions are used.

OTHER MATERIALS REQUIRED

- Aprotinin (Sigma # A6279)
- Leupeptin (Sigma # L8511)
- Pepstatin (Sigma # P4265)
- Phenylmethylsulfonylfluoride (PMSF) (Sigma # P7626)
- Sodium azide (NaN₃) (Sigma # S2002)
- Triton[®] X-100 (Sigma # T9284)
- Pipettes and pipette tips
- Deionized or distilled water
- 96 well microplates [Costar EIA Plates (Catalog # 2592 or R&D Systems Catalog # DY990) are suggested]
- Plate sealers (R&D Systems, Catalog # DY992)
- Squirt bottle, manifold dispenser, or automated microplate washer

Triton is a registered trademark of Union Carbide.

SOLUTIONS REQUIRED

PBS - 137 mM NaCl, 2.7 mM KCl, 8.1 mM Na₂HPO₄, 1.5 mM KH₂PO₄, pH 7.2 - 7.4, 0.2 µm filtered.

Wash Buffer - 0.05% Tween[®] 20 in PBS, pH 7.2 - 7.4 (R&D Systems, Catalog # WA126).

Block Buffer - 1% BSA,* 0.05% NaN₃, in PBS, pH 7.2 - 7.4.

IC Diluent #1 - 1% BSA* in PBS, pH 7.2 - 7.4, 0.2 µm filtered.

IC Diluent #9** - 1 mM EDTA, 0.5% Triton X-100, 0.5 M NaCl in PBS, pH 7.2 - 7.4.

Note: *Approximately 50 mL of this diluent is required to run the assay on one 96-well plate.*

Lysis Buffer #4** - 1 mM EDTA, 0.5% Triton X-100, 0.5 M NaCl, 10 µg/mL Leupeptin, 10 µg/mL Pepstatin, 100 µM PMSF, and 3 µg/mL Aprotinin in PBS, pH 7.2 - 7.4.

Substrate Solution - 1:1 mixture of Color Reagent A (H₂O₂) and Color Reagent B (Tetramethylbenzidine) (R&D Systems, Catalog # DY999).

Stop Solution - 2 N H₂SO₄ (R&D Systems, Catalog # DY994).

*The use of R&D Systems Reagent Diluent Concentrate 2 (Catalog # DY995) or Millipore Bovine Serum Albumin, Fraction V, Protease free (Catalog # 82-045) is recommended. All buffers containing BSA must be stored at 2 - 8° C.

**Sample Diluent Concentrate 1 (5X) (R&D Systems, Catalog # DYC001), prepared as described in the DYC001 insert.

Tween is a registered trademark of ICI Americas.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Total SMAC/Diablo Capture Antibody (Part 840761) - Each vial contains 144 µg/mL of rabbit anti-human SMAC/Diablo when reconstituted with 200 µL of PBS. After reconstitution, store at 2 - 8° C for up to 30 days or aliquot and store at ≤ -20° C in a manual defrost freezer or at ≤ -70° C for up to 3 months.*

Total SMAC/Diablo Detection Antibody (Part 840762) - Each vial contains 5.4 µg/mL of biotinylated rabbit anti-human SMAC/Diablo when reconstituted with 1.0 mL of IC Diluent #1. After reconstitution, store at 2 - 8° C for up to 30 days or aliquot and store at ≤ -20° C in a manual defrost freezer or at ≤ -70° C for up to 3 months.*

Total SMAC/Diablo Standard (Part 840763) - Each vial contains 70 ng/mL of recombinant human SMAC/Diablo when reconstituted with 500 µL of IC Diluent #9. **Use within one hour of reconstitution. A fresh standard should be used for each assay.** A seven point standard curve using 2-fold serial dilutions and a high standard of 4000 pg/mL is recommended.

Streptavidin-HRP (Part 890803) - 1.0 mL of streptavidin conjugated to horseradish-peroxidase. Store at 2 - 8° C. **DO NOT FREEZE.**

*Provided this is within the expiration date of the kit.

PREPARATION OF SAMPLES

Cell Lysates - Rinse cells two times with PBS, making sure to remove any remaining PBS after the second rinse. Solubilize cells at 1×10^7 cells/mL in Lysis Buffer #4 and allow samples to sit on ice for 15 minutes. Assay immediately or store at $\leq -70^\circ \text{C}$. Before use, centrifuge samples at $2000 \times g$ for 5 minutes and transfer the supernate to a clean test tube. Sample protein concentration may be quantified using a total protein assay. If needed, further dilutions should be made in IC Diluent #9.

PRECAUTION

The Stop Solution suggested for use with this kit is an acidic solution. Wear eye, hand, face, and clothing protection when using this material.

TECHNICAL HINTS AND LIMITATIONS

- This DuoSet IC ELISA should not be used beyond the expiration date on the label.
- Individual results may vary due to differences in technique, plasticware and water sources.
- It is important that the diluents selected for reconstitution and for dilution of the standard reflect the environment of the samples being measured. The diluent suggested in this protocol should be suitable for most cell lysates.
- The type of enzyme and substrate and the concentrations of capture/detection antibodies used can be varied to create an immunoassay with a different sensitivity and dynamic range. A basic understanding of immunoassay development is required for the successful use of these reagents in immunoassays.
- A thorough and consistent wash technique is essential for proper assay performance. Wash Buffer should be dispensed forcefully and removed completely from the wells by aspiration or decanting. Remove any remaining Wash Buffer by inverting the plate and blotting it against clean paper towels.
- Use a fresh reagent reservoir and pipette tips for each step.
- It is recommended that all standards and samples be assayed in duplicate.
- Avoid microbial contamination of reagents and buffers. This may interfere with the sensitivity of the assay. Buffers containing protein should be made under aseptic conditions and stored at $2 - 8^\circ \text{C}$ or be prepared fresh daily.

GENERAL ELISA PROTOCOL

A plate layout is provided to record standards and samples assayed.

Plate Preparation

1. Dilute the Capture Antibody to a working concentration of 0.8 $\mu\text{g}/\text{mL}$ in PBS without carrier protein. Immediately coat a 96 well microplate with 100 μL per well of the diluted Capture Antibody. Seal the plate and incubate overnight at room temperature.
2. Aspirate each well and wash with Wash Buffer, repeating the process two times for a total of 3 washes. Wash by filling each well with Wash Buffer (400 μL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.
3. Block plates by adding 300 μL of Block Buffer to each well. Incubate at room temperature for 1 - 2 hours.
4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

Assay Procedure

1. Add 100 μL of sample or standards in IC Diluent #9 per well. Use IC Diluent #9 as the zero standard. Cover with a plate sealer and incubate 2 hours at room temperature.
Note: *A seven point standard curve using 2-fold serial dilutions and a high standard of 4000 pg/mL is recommended.*
2. Repeat the aspiration/wash as in step 2 of Plate Preparation.
3. Dilute the Detection Antibody to a working concentration of 150 ng/mL in IC Diluent #1 before use. Add 100 μL of the diluted Detection Antibody to each well. Cover with a new plate sealer and incubate 2 hours at room temperature.
4. Repeat the aspiration/wash as in step 2 of Plate Preparation.
5. Immediately before use, dilute the Streptavidin-HRP to the working concentration specified on the vial label using IC Diluent #1. Add 100 μL of the diluted Streptavidin-HRP to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
6. Repeat the aspiration/wash as in step 2 of Plate Preparation.
7. Add 100 μL of Substrate Solution to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
8. Add 50 μL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

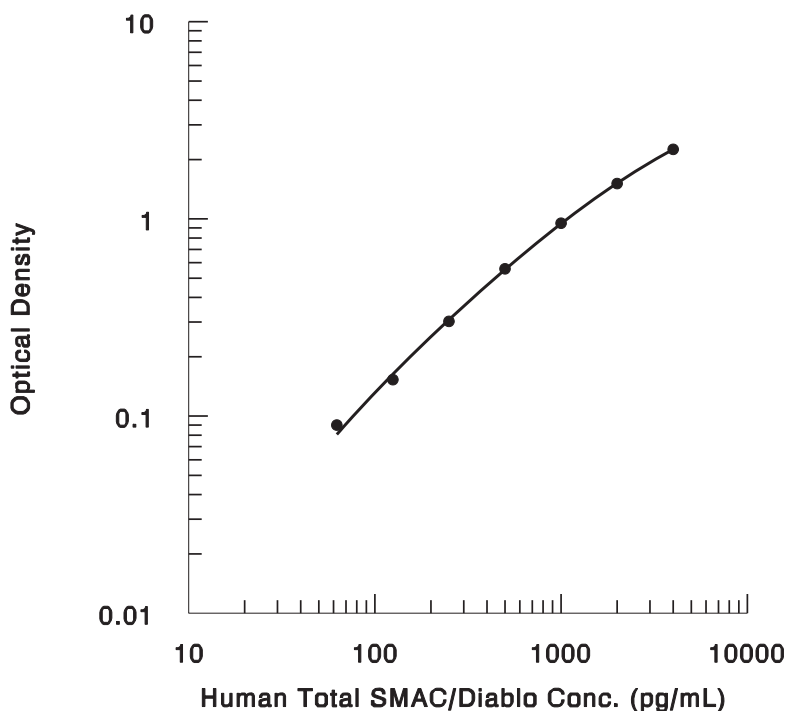
CALCULATION OF RESULTS

Average the duplicate readings for each standard and sample and then subtract the average zero standard optical density.

Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the SMAC/Diablo concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

TYPICAL DATA

A standard curve should be generated for each set of samples assayed. The graph below represents typical data generated when using this Human Total SMAC/Diablo DuoSet IC ELISA. The standard curve was calculated using a computer generated 4-PL curve-fit. This standard curve is for demonstration purposes only.



CALIBRATION

This DuoSet IC ELISA is calibrated against a highly purified *E. coli*-expressed recombinant human SMAC/Diablo produced at R&D Systems.

SPECIFICITY

The Human Total SMAC/Diablo DuoSet IC ELISA is specific for SMAC/Diablo. Specificity was demonstrated by Western blot analysis of the protein bound by the Capture Antibody supplied in the kit.

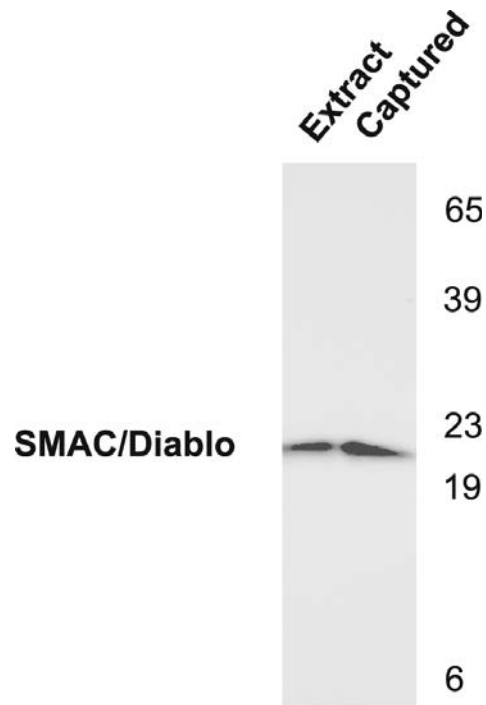


Figure 1: Lysates prepared from human CHP-100 cells were incubated in wells coated with SMAC/Diablo Capture Antibody. Unbound material was removed by washing and bound material was solubilized in SDS gel sample buffer. Captured proteins were electrophoresed, transferred to Immobilon (Millipore) membranes and immunoblotted with SMAC/Diablo Detection Antibody. Only a single band corresponding to human SMAC/Diablo was detected.

QUANTIFICATION

The amounts of SMAC/Diablo, as measured by the Human Total SMAC/Diablo DuoSet IC ELISA, are consistent with the amounts of SMAC/Diablo determined by qualitative Western blot analysis.

Quantification of human SMAC/Diablo in Jurkat and CHP-100 cells

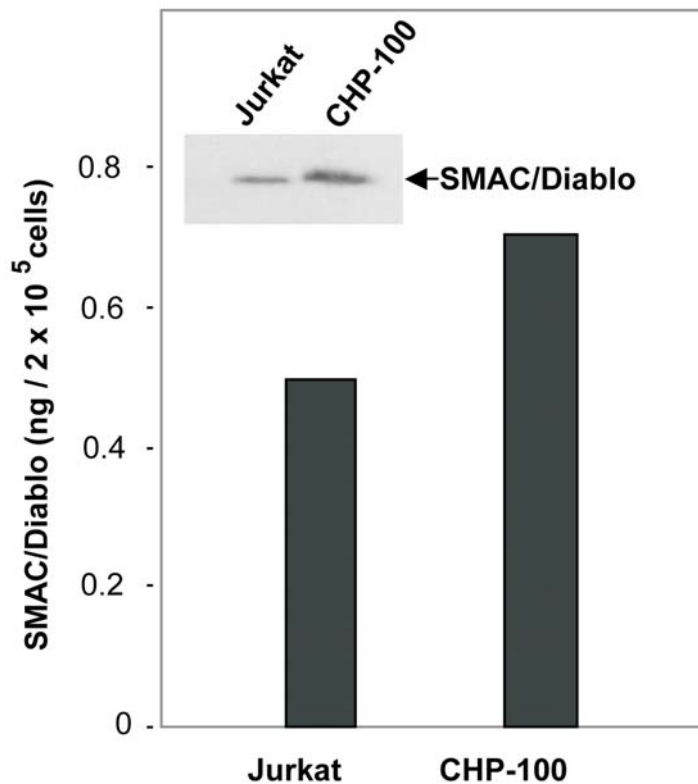


Figure 2: Jurkat and CHP-100 cells were lysed as described in the Preparation of Samples section on page 4. Human SMAC/Diablo was measured with the SMAC/Diablo DuoSet IC ELISA. The same lysates were immunoblotted (inset) with rabbit anti-human SMAC/Diablo (R&D Systems, Catalog # AF789). The DuoSet IC ELISA results correlate with SMAC/Diablo amounts detected by Western blot.

PLATE LAYOUT

Use this plate layout as a record of standards and samples assayed.

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