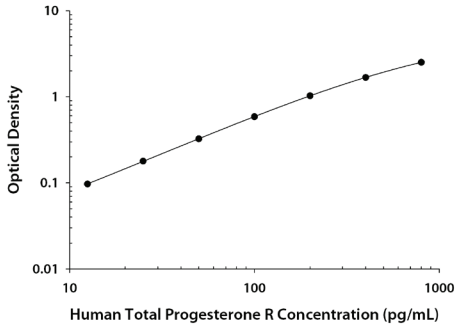
CALCULATION OF RESULTS

Average the duplicate readings for each standard and sample, then subtract the average zero standard optical density (O.D.). Results may be normalized to total protein or cell number.

Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the human Progesterone R concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

TYPICAL DATA

A standard curve should be generated for each set of samples assayed. The graph below represents typical data generated when using the Human Total Progesterone R DuoSet IC ELISA. The standard curve was calculated using a computer generated 4-PL curve-fit. This standard curve is for demonstration purposes only.



CALIBRATION

The Human Total Progesterone R/NR3C3 DuoSet IC ELISA is calibrated against a highly purified *E. coli*-expressed recombinant human Progesterone R produced at R&D Systems®. Samples containing natural Progesterone R showed linear dilution parallel to the standard curve obtained using the Human Total Progesterone R Standard. These results indicate that O.D. values from this DuoSet IC ELISA can be used to determine the concentration of Progesterone R in natural samples.

SPECIFICITY

The Human Total Progesterone R/NR3C3 DuoSet IC ELISA specifically recognizes PR. Specificity was demonstrated using cross-reactivity experiments with related nuclear hormone receptors. Recombinant human (rh) Estrogen Receptor alpha (ERα), rhERβ and rhAbdrogen receptor (AR) were each tested at 50 ng/mL and did not cross-react or interfere in the assay.

TECHNICAL HINTS & LIMITATIONS

- This DuoSet IC ELISA should not be used beyond the expiration date on the kit label.
- Individual results may vary due to differences in technique, plasticware, and water sources.
- It is important that the diluents selected for reconstitution and for dilution of the samples and standard reflect the environment of the samples being measured. The diluents suggested in this protocol should be suitable for most cell lysates.
- The type of enzyme and substrate and the concentrations of capture/detection antibodies used can be varied to create an immunoassay with a different sensitivity and dynamic range. A basic understanding of immunoassay development is required for the successful use of these reagents in immunoassays.
- A thorough and consistent wash technique is essential for proper assay performance. Wash Buffer should be dispensed forcefully and removed completely from the wells by aspiration or decanting. Remove any remaining Wash Buffer by inverting the plate and blotting it against clean paper towels.
- Use a fresh reagent reservoir and pipette tips for each step.
- It is recommended that all standards and samples be assayed in duplicate.
- Avoid microbial contamination of reagents and buffers. This may interfere with the sensitivity of the assay. Buffers containing protein should be made under aseptic conditions and stored at 2-8 °C or be prepared fresh daily.

PRECAUTIONS

The Stop Solution recommended for use with this kit is an acid solution.

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

DuoSet™ IC ELISA
INTRACELLULAR

Human Total Progesterone R/NR3C3

Catalog Number: DYC5415-2 (2 plates)
DYC5415-5 (5 plates)

INTENDED USE

For the development of sandwich ELISAs to measure human Progesterone R (PR) in cell lysates.

PRINCIPLE OF THE ASSAY

This DuoSet IC ELISA contains the basic components required for the development of sandwich ELISAs to measure human Progesterone Receptor (Progesterone R), also known as NR3C3 and PGR, in cell lysates. An immobilized capture antibody specific for the two Progesterone R isoforms, full-length Progesterone R-B and truncated Progesterone R-A, binds both phosphorylated and unphosphorylated protein. After washing away unbound material, a biotinylated detection antibody specific for human Progesterone R is used to detect the captured protein, utilizing a standard Streptavidin-HRP format.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

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MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

DESCRIPTION	PART #	CATALOG # DYC5415-2	CATALOG # DYC5415-5	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Human Total Progesterone R Capture Antibody	843529	1 vial	2 vials	Store for up to 1 month at 2-8 °C or aliquot and store at ≤ -20 °C or ≤ -70 °C for up to 3 months in a manual defrost freezer.*
Human Total Progesterone R Detection Antibody	843530	1 vial	2 vials	
Streptavidin-HRP A	890803	1 vial	1 vial	Store for up to 3 months at 2-8 °C.* **DO NOT FREEZE.**
Human Total Progesterone R Standard	843531	3 vials	5 vials	**Use within one hour of reconstitution. Use a fresh standard for each assay.**

* Provided this is within the expiration date of the kit.

DYC5415-2 contains sufficient materials to run ELISAs on at least two 96 well plates.†
DYC5415-5 contains sufficient materials to run ELISAs on at least five 96 well plates.†

† Provided the following conditions are met:

- The reagents are prepared as described in this package insert.
- The assay is run as described in the General ELISA Protocol.
- The recommended microplates, buffers, diluents, substrates, and solutions are used.

OTHER MATERIALS REQUIRED

- Aprotinin (Tocris™, # 4139)
- Leupeptin (Tocris, # 1167)
- Pepstatin (Tocris, # 1190)
- Phenylmethylsulfonyl Fluoride (PMSF) (Sigma™, # P7626)
- Sodium Azide (NaN_3) (Sigma, # S2002)
- Sodium Fluoride (NaF) (Sigma, # 201154)
- Sodium Orthovanadate (Na_3VO_4) (Sigma, # S6508), activated
- Sodium Pyrophosphate ($Na_4P_2O_7$) (Sigma, # P8010)
- Triton™ X-100 (Sigma, # T9284)
- Urea
- Pipettes and pipette tips
- Deionized or distilled water
- 96 well microplates (R&D Systems, # DY990)
- Plate sealers (R&D Systems, # DY992)
- Squirt bottle, manifold dispenser, or automated microplate washer.

SOLUTIONS REQUIRED

PBS - 137 mM NaCl, 2.7 mM KCl, 8.1 mM Na_2HPO_4, 1.5 mM KH_2PO_4, pH 7.2-7.4, 0.2 µm filtered. (R&D Systems, # DY006)

Wash Buffer - 0.05% Tween® 20 in PBS, pH 7.2-7.4 (R&D Systems, # WA126)

Block Buffer - 1% BSA*, 0.05% NaN_3, in PBS, pH 7.2-7.4

IC Diluent #1 - 1% BSA* in PBS, pH 7.2-7.4, 0.2 µm filtered

IC Diluent #8** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF in PBS, pH 7.2-7.4
Note: *IC Diluent #8 is also the base diluent for IC Diluent #3, IC Diluent #7, and Lysis Buffer #6. Approximately 50 mL of this diluent is required to run the assay on one 96 well plate.*

IC Diluent #3** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 1 M Urea in PBS, pH 7.2-7.4

IC Diluent #7** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M Urea in PBS, pH 7.2-7.4

Lysis Buffer #6** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M Urea, 1 mM activated Sodium Orthovanadate, 2.5 mM Sodium Pyrophosphate, 10 µg/mL Leupeptin, 10 µg/mL Pepstatin, 100 µM PMSF, 3 µg/mL Aprotinin in PBS, pH 7.2-7.4

Substrate Solution: ELISA TMB Substrate (R&D Systems, # DY999B)

Stop Solution - 2N H_2SO_4 (R&D Systems, # DY994)

*The use of R&D Systems Reagent Diluent Concentrate 2 (Catalog # DY995) or Millipore Bovine Serum Albumin, Fraction V, Protease free (Catalog # 82-045) is recommended. All buffers containing BSA must be stored at 2-8 °C.

**Alternatively, use Sample Diluent Concentrate 1 (5X) (Catalog # DYC001), prepared as described in the DYC001 insert.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Human Total Progesterone R Capture Antibody (Part 843529)
Each vial contains 360 µg/mL of sheep anti-human Progesterone R antibody when reconstituted with 200 µL of PBS.

Human Total Progesterone R Detection Antibody (Part 843530)
Each vial contains 18 µg/mL of biotinylated sheep anti-human Progesterone R antibody when reconstituted with 1 mL of IC Diluent #1. Immediately before use, dilute the detection antibody to a working concentration of 500 ng/mL in IC Diluent #1. Prepare only as much detection antibody as required to run each assay.

Human Total Progesterone R Standard (Part 843531) - **Reconstitute with a recommended volume of 500 µL of IC Diluent #7 to produce a stock solution. Refer to the vial label for the concentration of recombinant human Progesterone R.** Immediately before use, an initial 6-fold dilution should be made in IC Diluent #8. Additional dilutions should be made in IC Diluent #3. A seven point standard curve using 2-fold serial dilutions and a high standard of 800 pg/mL is recommended.

Streptavidin-HRP A (Part 890803) - 1 mL of Streptavidin conjugated to horseradish-peroxidase. Immediately before use, dilute the Streptavidin-HRP A to the working concentration specified on the vial label using IC Diluent #1.

PREPARATION OF SAMPLES

Cell Lysates - Rinse cells two times with PBS, making sure to remove any remaining PBS after the second rinse. Solubilize cells at 1 x 10^7 cells/mL in Lysis Buffer #6 and allow samples to sit on ice for 15 minutes. Assay immediately or store at ≤ -70 °C. Before use, centrifuge samples at 2000 x g for 5 minutes, and transfer the supernate to a clean test tube. Sample protein concentration may be quantified using a total protein assay. For assaying, dilute lysates 6-fold with IC Diluent #8 and make further serial dilutions in IC Diluent #3.
Note: *The final concentration of urea in all samples and standards should be 1 M prior to addition to the plate.*

GENERAL ELISA PROTOCOL

Plate Preparation

1. Dilute the capture antibody to a working concentration of 2 µg/mL in PBS, without carrier protein. Immediately coat a 96 well microplate with 100 µL per well of the diluted capture antibody. Seal the plate and incubate overnight at room temperature.
2. Aspirate each well and wash with Wash Buffer, repeating the process two times for a total of 3 washes. Wash by filling each well with Wash Buffer (400 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.
3. Block plates by adding 300 µL of Block Buffer to each well. Incubate at room temperature for 1-2 hours.
4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

Assay Procedure

1. Add 100 µL of sample or standard in IC Diluent #3 per well. Use IC Diluent #3 as the zero standard. Cover with a plate sealer and incubate 2 hours at room temperature.
   Note: *A seven point standard curve using 2-fold serial dilutions and a high standard of 800 pg/mL is recommended.*
2. Repeat the aspiration/wash as in step 2 of Plate Preparation.
3. Add 100 µL of the diluted detection antibody to each well. Cover with a new plate sealer and incubate 2 hours at room temperature.
4. Repeat the aspiration/wash as in step 2 of Plate Preparation.
5. Add 100 µL of the diluted Streptavidin-HRP A to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
6. Repeat the aspiration/wash as in step 2 of Plate Preparation.
7. Add 100 µL of Substrate Solution to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
8. Add 50 µL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.