Recombinant mouse:
decrease the observed reading of rhFractalkine:
The following 500 ng/mL samples of recombinant rat (rr) Fractalkine reads as 2.18 ng/mL (6.97% cross-reactivity).
A sample containing 31.25 ng/mL of recombinant mouse GCP-2 This DuoSet detects the chemokine domain of human Fractalkine.
Fractalkine (Chemokine domain aa 25-105)
Fractalkine (Chemokine domain and mucin stalk)
Fractalkine (Chemokine domain aa 22-105)
exhibited no cross-reactivity or interference.
The following factors prepared at 500 ng/mL were assayed and
TECHNICAL HINTS & LIMITATIONS
• We recommend the use of R&D Systems’ Reagent Diluent Concentrate 2 (Catalog # DY995) to prepare Reagent Diluent for use in this assay.
• The use of high quality Bovine Serum Albumin (BSA) for the Reagent Diluent is crucial for the optimum performance of the DuoSet ELISA Development kit. Impurities such as proteases, binding proteins, soluble receptors or other interfering substances can be found to varying degrees in virtually all BSA preparations and can inhibit or interfere with the detection of certain analytes. If the standard curve appears suppressed, consider evaluating a different preparation of BSA.
• It is suggested to start Reagent Diluent optimization for serum and plasma samples by using PBS supplemented with 10-50% animal serum. Do not use buffers with animal serum to reconstitute or dilute the Detection Antibody or Streptavidin-HRP.
• It is important that the Reagent Diluent selected for reconstitution and dilution of the standard reflects the environment of the samples being measured.
• Avoid microbial contamination of reagents and buffers.
• A thorough and consistent wash technique is essential for proper assay performance. Wash Buffer should be dispensed forcefully and removed completely from the wells by aspiration or decanting. Remove any remaining Wash Buffer by inverting the plate and blotting it against clean paper towels.
• Individual results may vary due to differences in technique, plasticware and water sources.
• It is recommended that all standards and samples be assayed in duplicate.
• The use of PBS from tablets may interfere in this assay.

TYPICAL DATA
This standard curve is only for demonstration purposes. A standard curve should be generated for each set of samples assayed.

SPECIFICITY
The following factors prepared at 500 ng/mL were assayed and exhibited no cross-reactivity or interference.

Recombinant mouse:
Fractalkine (Chemokine domain aa 22-105)
Fractalkine (Chemokine domain and mucin stalk)
Fractalkine (Chemokine domain aa 25-105)
This DuoSet detects the chemokine domain of human Fractalkine.
A sample containing 31.25 ng/mL of recombinant mouse GCP-2 reads as 2.18 ng/mL (6.97% cross-reactivity).
The following 500 ng/mL samples of recombinant rat (rr) Fractalkine chemokine domain did not exhibit any cross-reactivity, but did decrease the observed reading of rrFractalkine:

<table>
<thead>
<tr>
<th>rFractalkine</th>
<th>Expected rFractalkine Concentration</th>
<th>Measured rFractalkine Concentration</th>
<th>% Decrease</th>
</tr>
</thead>
<tbody>
<tr>
<td>aa 22-100</td>
<td>625 pg/mL</td>
<td>479 pg/mL</td>
<td>24</td>
</tr>
<tr>
<td>aa 25-100</td>
<td>5 ng/mL</td>
<td>4 ng/mL</td>
<td>20</td>
</tr>
</tbody>
</table>

TROUBLESHOOTING
Note: For more detailed troubleshooting, please visit: www.RnDSystems.com/ELISADevelopment

Poor Standard Curve
• Impure BSA used for Reagent Diluent preparation.
• Improper reconstitution and/or storage of standard.
• Improper dilution of highest standard and standard curve.
• Incomplete washing and/or aspiration of wells.
• Inadequate volume of substrate added to wells.
• Incomplete washing or pipetting error.

Poor Precision
• Unequal volumes added to wells/pipetting error.
• Incomplete washing and/or aspiration of wells.
• Unequal mixing of reagents.

Low or No color Development
• Inadequate volume of substrate added to wells.
• Incorrect incubation times or temperatures.
• Unequal volumes added to wells/pipetting error.
• Imprope BSA used for Reagent Diluent preparation.

This package insert must be read in its entirety before using this product. Refer to the Certificate of Analysis for component concentrations as they may vary. For research use only. Not for use in diagnostic procedures.

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www.RnDSystems.com
**Substrate Solution:**
Quality of BSA is critical (see Technical Hints).
(R&D Systems, Catalog # DY995).

**Reagent Diluent:**
1% BSA in PBS, pH 7.2-7.4, 0.2 μm filtered
(R&D Systems, Catalog # WA126).

**Wash Buffer:**
0.05% Tween® 20 in PBS, pH 7.2-7.4
pH 7.2-7.4, 0.2 μm filtered (R&D Systems, Catalog # DY006).

**2 N H₂SO₄ (R&D Systems, Catalog # DY994).**

**Stop Solution:**
Reagent B (Tetramethylbenzidine) (R&D Systems, Catalog # DY999).

**recombinant human Fractalkine produced at R&D Systems.**

**CALIBRATION**
Website prior to use.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Please refer to the MSDS on our website for more details.

**PRECAUTIONS**
Some components in this kit contain ProClin® which may cause an allergic skin reaction. Avoid breathing mist.

The Stop Solution suggested for use with this kit is an acid solution.

The Color Reagent B suggested for use with this kit may cause skin, eye, and respiratory irritation. Avoid breathing fumes.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Please refer to the MSDS on our website prior to use.

**CALIBRATION**
This DuoSet is calibrated against a highly purified N50-expressed recombinant human Fractalkine produced at R&D Systems.

**OTHER MATERIALS & SOLUTIONS REQUIRED**

**DuoSet Ancillary Reagent Kit 2 (5 plates):**
(R&D Systems, Catalog # DY008) containing 96 well microplates, plate sealers, substrate solution, stop solution, plate coating buffer (PBS), wash buffer, and Reagent Diluent Concentrate 2.

**The components listed above may be purchased separately:**
96 well microplates: (R&D Systems, Catalog # DY990).

**Plate Sealers:**
(R&D Systems, Catalog # DY992).

**PBS:**
137 mM NaCl, 2.7 mM KCl, 8.1 mM Na₂HPO₄, 1.5 mM KH₂PO₄, pH 7.2-7.4 (R&D Systems, Catalog # DY008).

**Wash Buffer:**
0.05% Tween® 20 in PBS, pH 7.2-7.4
pH 7.2-7.4, 0.2 μm filtered (R&D Systems, Catalog # DY006).

**Reagent Diluent Concentrate 2:**
1% BSA in PBS, pH 7.2-7.4, 0.2 μm filtered
(R&D Systems, Catalog # WA126).

**Reagent Diluent:**
1% BSA in PBS, pH 7.2-7.4, 0.2 μm filtered
(R&D Systems, Catalog # DY995).

**Quality of BSA is critical** (see Technical Hints).

**Substrate Solution:**
1:1 mixture of Color Reagent A (H₂O₂) and Color Reagent B (Tetramethylbenzidine) (R&D Systems, Catalog # DY999).

**Stop Solution:**
2 N H₂SO₄ (R&D Systems, Catalog # DY994).

**REAGENT PREPARATION**

**Bring all reagents to room temperature before use. Allow all components to sit for a minimum of 15 minutes with gentle agitation after initial reconstitution. Working dilutions should be prepared and used immediately.**

**Streptavidin-HRP:**
1.0 mL of streptavidin conjugated to horseradish peroxidase. Dilute to the working concentration specified on the vial label using Reagent Diluent.

**Mouse Anti-Human Fractalkine Capture Antibody:**
Refer to the lot-specific Certificate of Analysis (C of A) for storage conditions.

**Recombinant Human Fractalkine Standard:**
Refer to the lot-specific Certificate of Analysis (C of A) for storage conditions.

**Materials Provided & Storage Conditions**

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

<table>
<thead>
<tr>
<th>DESCRIPTION</th>
<th>PART #</th>
<th># VIALS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human Fractalkine Capture Antibody</td>
<td>840486</td>
<td>1 vial</td>
</tr>
<tr>
<td>Human Fractalkine Detection Antibody</td>
<td>840487</td>
<td>1 vial</td>
</tr>
<tr>
<td>Human Fractalkine Standard</td>
<td>840488</td>
<td>3 vials</td>
</tr>
<tr>
<td>Streptavidin-HRP</td>
<td>893410</td>
<td>1 vial</td>
</tr>
</tbody>
</table>

**GENERAL ELISA PROTOCOL**

**Plate Preparation**

1. Dilute the Capture Antibody to the working concentration in PBS without carrier protein. Immediately coat a 96-well microplate with 100 μL per well of the diluted Capture Antibody. Seal the plate and incubate overnight at room temperature.

2. Aspirate each well and wash with Wash Buffer, repeating the process two times for a total of three washes. Wash by filling each well with Wash Buffer (400 μL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.

3. Block plates by adding 300 μL of Reagent Diluent to each well. Incubate at room temperature for a minimum of 1 hour.

4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

**Assay Procedure**

1. Add 100 μL of sample or standards in Reagent Diluent, or an appropriate diluent, per well. Cover with an adhesive strip and incubate 2 hours at room temperature.

2. Repeat the aspiration/wash as in step 2 of Plate Preparation.

3. Add 100 μL of the Detection Antibody, diluted in Reagent Diluent, to each well. Cover with a new adhesive strip and incubate 2 hours at room temperature.

4. Repeat the aspiration/wash as in step 2 of Plate Preparation.

5. Add 100 μL of the working dilution of Streptavidin-HRP to each well. Cover the plate and incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.

6. Repeat the aspiration/wash as in step 2.

7. Add 100 μL of Substrate Solution to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.

8. Add 50 μL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.

9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

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