

Quantikine[®] ELISA

Human Total MMP-7 Immunoassay

Catalog Number DMP700

SMP700

PDMP700

For the quantitative determination of human active and pro-Matrix Metalloproteinase 7 (Total MMP-7) concentrations in cell culture supernates, serum, plasma, saliva, and urine.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

TABLE OF CONTENTS

SECTION	PAGE
INTRODUCTION	1
PRINCIPLE OF THE ASSAY.....	2
LIMITATIONS OF THE PROCEDURE	2
TECHNICAL HINTS.....	2
MATERIALS PROVIDED & STORAGE CONDITIONS	3
OTHER SUPPLIES REQUIRED	4
PRECAUTIONS.....	4
SAMPLE COLLECTION & STORAGE.....	5
SAMPLE PREPARATION.....	5
REAGENT PREPARATION	6
ASSAY PROCEDURE	7
CALCULATION OF RESULTS.....	8
TYPICAL DATA.....	8
PRECISION	9
RECOVERY.....	9
LINEARITY.....	9
SENSITIVITY	10
CALIBRATION	10
SAMPLE VALUES.....	10
SPECIFICITY.....	11
REFERENCES.....	12
PLATE LAYOUT	13

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INTRODUCTION

Matrix metalloproteinases (MMPs), also called matrixins, constitute a family of zinc and calcium dependent endopeptidases that function in the breakdown of extracellular matrix (ECM). They play an important role in many normal physiological processes such as embryonic development, morphogenesis, reproduction and tissue remodeling (1). They also participate in many pathological processes such as arthritis, cancer and cardiovascular disease (2). While the amounts of newly synthesized MMPs are regulated mainly at the levels of transcription, the proteolytic activities of existing MMPs are controlled through both the activation of proenzymes or zymogens and the inhibition of active enzymes by endogenous inhibitors, α_2 -macroglobulins and tissue inhibitors of metalloproteinases (TIMPs).

MMP-7 (matrilysin) is expressed in the epithelial cells of normal and diseased tissues (3). The protein localizes in normal tissues to secretory and ductal epithelium in the endometrium and in various exocrine glands. It is expressed in a variety of tumors ranging from breast, colon, prostate, stomach, upper aerodigestive tract, lung, and skin. The transcription of the gene is activated by Ets transcription factors of the PEA3 subfamily in intestinal tumors and by *Pseudomonas aeruginosa* flagellin in airway epithelial cells (4, 5). Knockout mice lacking the gene have suppressed intestinal tumorigenesis (6). Over-expression of the gene results in premature mammary gland differentiation and male infertility (7).

MMP-7 is capable of digesting many proteins of the extracellular matrix such as collagen IV, gelatins, laminin, aggrecan, entactin, elastin, and versican. It activates other proteinases such as urokinase plasminogen activator and pro-MMP-1, -2, and -9, and cleaves additional substrates such as osteopontin (3, 8, 9). In addition to its roles in connective tissue remodeling and cancer, MMP-7 also regulates intestinal α -defensin activation in innate host defense and releases TNF- α in a model of herniated disc resorption (10, 11). MMP-7-mediated cleavage of Fas Ligand protects tumor cells from chemotherapeutic drug cytotoxicity and potentiates epithelial cell apoptosis (12, 13).

Structurally, MMP-7 is one of the smallest MMPs, consisting of two domains, a pro-domain and a catalytic domain (8). Activation of the proenzyme involves a proteolytic removal of the N-terminal pro-region containing the cysteine switch motif conserved in MMPs (14). The resulting mature and active enzyme consists of a catalytic domain with a zinc-binding motif conserved in metzincins (15, 16). Alternatively, activation of the proenzyme can be through oxygenation of the cysteine switch motif by hypochlorous acid (17).

The Quantikine[®] Human Total MMP-7 Immunoassay is a 4.5 hour solid phase ELISA designed to measure human MMP-7 in cell culture supernates, serum, plasma, saliva, and urine. It contains NS0-expressed recombinant human MMP-7 and antibodies raised against the recombinant factor. The Quantikine[®] kit detected both pro- and active forms of recombinant human MMP-7. Natural human MMP-7 showed dose-response curves that were parallel to the standard curves obtained using the Quantikine[®] kit standards. These results indicate that this kit can be used to determine relative levels of natural human MMP-7.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for human MMP-7 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells, and MMP-7 is bound by the immobilized antibody. After washing away unbound substances, an enzyme-linked polyclonal antibody specific for human MMP-7 is added to the wells. Following a wash to remove unbound antibody-enzyme reagent, a substrate solution is added to the wells and color develops in proportion to the amount of total MMP-7 (pro and/or active) bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples generate values higher than the highest standard, further dilute the samples with calibrator diluent and repeat the assay. If cell culture supernate samples require large dilutions, perform an intermediate dilution with culture media and the final dilution with calibrator diluent.
- Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- Variations in sample collection, processing, and storage may cause sample value differences.
- This assay is designed to eliminate interference by other factors present in biological samples. Until all factors have been tested in the Quantikine® Immunoassay, the possibility of interference cannot be excluded.

TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- When using an automated plate washer, adding a 30 second soak period following the addition of Wash Buffer, and/or rotating the plate 180 degrees between wash steps may improve assay precision.
- Substrate Solution should remain colorless until added to the plate. Keep Substrate Solution protected from light. Substrate Solution should change from colorless to gradations of blue.
- Stop Solution should be added to the plate in the same order as the Substrate Solution. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.

MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

PART	PART #	CATALOG # DMP700	CATALOG # SMP700	DESCRIPTION	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Human Total MMP-7 Microplate	891132	1 plate	6 plates	96 well polystyrene microplate (12 strips of 8 wells) coated with a monoclonal antibody specific for human MMP-7.	Return unused wells to the foil pouch containing the desiccant pack. Reseal along entire edge of zip-seal. May be stored for up to 1 month at 2-8 °C.*
Human Total MMP-7 Conjugate	891133	1 vial	6 vials	21 mL/vial of a polyclonal antibody specific for human MMP-7 conjugated to horseradish peroxidase with preservatives.	May be stored for up to 1 month at 2-8 °C.*
Human Total MMP-7 Standard	891134	1 vial	6 vials	Recombinant human Pro-MMP-7 in a buffered protein base with preservatives; lyophilized. <i>Refer to the vial label for reconstitution volume.</i>	
Assay Diluent RD1-52	895343	1 vial	6 vials	11 mL/vial of a buffered protein base with preservatives. <i>May contain a precipitate. Mix well before and during use.</i>	
Calibrator Diluent RD6-28	895345	1 vial	6 vials	21 mL/vial of diluted animal serum with preservatives.	
Wash Buffer Concentrate	895003	1 vial	6 vials	21 mL/vial of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>	
Color Reagent A	895000	1 vial	6 vials	12 mL/vial of stabilized hydrogen peroxide.	
Color Reagent B	895001	1 vial	6 vials	12 mL/vial of stabilized chromogen (tetramethylbenzidine).	
Stop Solution	895032	1 vial	6 vials	6 mL/vial of 2 N sulfuric acid.	
Plate Sealers	N/A	4 strips	24 strips	Adhesive strips.	

* Provided this is within the expiration date of the kit.

DMP700 contains sufficient materials to run an ELISA on one 96 well plate.

SMP700 (SixPak) contains sufficient materials to run ELISAs on six 96 well plates.

This kit is also available in a PharmPak (R&D Systems®, Catalog # PDMP700). PharmPaks contain sufficient materials to run ELISAs on 50 microplates. Specific vial counts of each component may vary. Refer to the literature accompanying your order for specific vial counts.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- Pipettes and pipette tips.
- 500 mL graduated cylinder.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- Horizontal orbital microplate shaker (0.12" orbit) capable of maintaining a speed of 500 ± 50 rpm.
- Collection device for saliva samples which has no enzyme binding or filtering capabilities such as a Salivette® or equivalent.
- **Polypropylene** test tubes for dilution of standards and samples.
- Human Total MMP-7 Controls (optional; R&D Systems®, Catalog # QC128).

PRECAUTIONS

MMP-7 is detectable in saliva. Take precautionary measures to prevent contamination of the kit reagents while running this assay.

The Stop Solution provided with this kit is an acid solution.

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Color Reagent B may cause skin, eye, and respiratory irritation. Avoid breathing fumes.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the MSDS on our website prior to use.

SAMPLE COLLECTION & STORAGE

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Note: *EDTA and citrate plasma are not recommended for use in this assay due to their chelating properties.*

Saliva - Collect saliva using a collection device such as a Salivette or equivalent. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Note: *Saliva collector cannot have any enzyme binding or filtering capabilities.*

Urine - Aseptically collect the first urine of the day (mid-stream), voided directly into a sterile container. Centrifuge to remove particulate matter. Assay immediately or aliquot and store at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

SAMPLE PREPARATION

Use polypropylene tubes.

Cell culture supernates, serum, plasma, and saliva samples require at least a 2-fold dilution prior to assay. A suggested 2-fold dilution is 100 μ L of sample + 100 μ L of Calibrator Diluent RD6-28.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Note: High concentrations of MMP-7 are found in saliva. The use of a face mask and gloves is recommended to protect kit reagents from contamination.

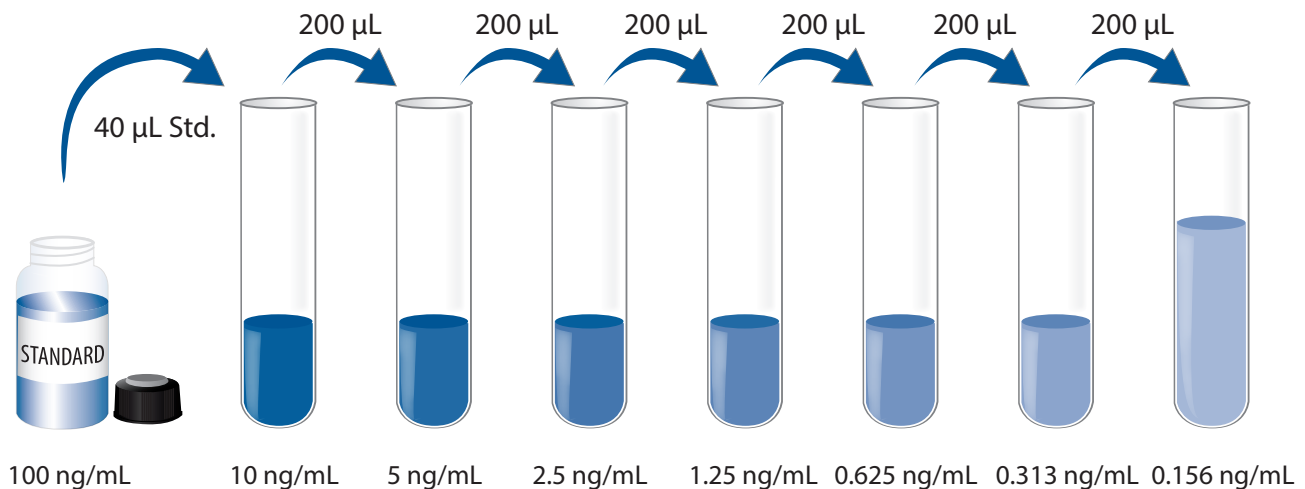
Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to deionized or distilled water to prepare 500 mL of Wash Buffer.

Substrate Solution - Color Reagents A and B should be mixed together in equal volumes within 15 minutes of use. Protect from light. 200 μ L of the resultant mixture is required per well.

Human Total MMP-7 Standard - Refer to the vial label for reconstitution volume.

Reconstitute the Human Total MMP-7 Standard with deionized or distilled water. This reconstitution produces a stock solution of 100 ng/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

Use polypropylene tubes. Pipette 360 μ L of Calibrator Diluent RD6-28 into the 10 ng/mL tube. Pipette 200 μ L of Calibrator Diluent RD6-28 into the remaining tubes. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 10 ng/mL standard serves as the high standard. Calibrator Diluent RD6-28 serves as the zero standard (0 ng/mL).



ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that all standards, samples, and controls be assayed in duplicate.

1. Prepare all reagents, working standards, and samples as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 100 μL of Assay Diluent RD1-52 to each well. *Assay Diluent RD1-52 may contain a precipitate. Mix well before and during use.*
4. Add 50 μL of standard, control, or sample* per well. Cover with the adhesive strip provided. Incubate for 2 hours at room temperature on a horizontal orbital microplate shaker (0.12" orbit) set at 500 ± 50 rpm. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (400 μL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 200 μL of Human Total MMP-7 Conjugate to each well. Cover with a new adhesive strip. Incubate for 2 hours at room temperature on the shaker.
7. Repeat the aspiration/wash as in step 5.
8. Add 200 μL of Substrate Solution to each well. Incubate for 30 minutes at room temperature **on the benchtop. Protect from light.**
9. Add 50 μL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green or the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
10. Determine the optical density of each well within 30 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

*Samples may require dilution. See the Sample Preparation section.

CALCULATION OF RESULTS

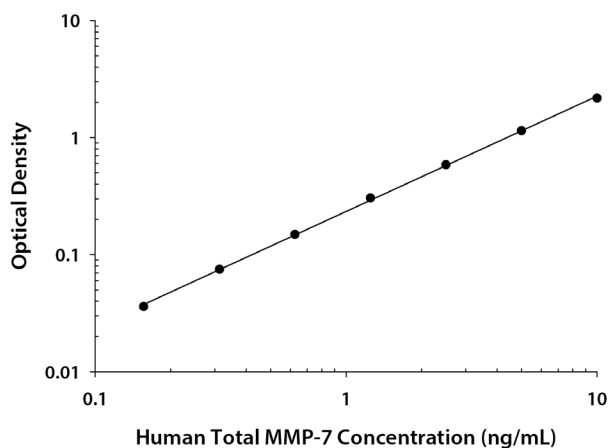
Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density (O.D.).

Create a standard curve by reducing the data using computer software capable of generating a log/log curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on a log/log graph. The data may be linearized by plotting the log of the human MMP-7 concentrations versus the log of the O.D. on a linear scale, and the best fit line can be determined by regression analysis.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

TYPICAL DATA

This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.



(ng/mL)	O.D.	Average	Corrected
0	0.008 0.010	0.009	—
0.156	0.044 0.046	0.045	0.036
0.313	0.083 0.085	0.084	0.075
0.625	0.157 0.158	0.158	0.149
1.25	0.309 0.319	0.314	0.305
2.5	0.583 0.607	0.595	0.586
5	1.132 1.173	1.152	1.143
10	2.121 2.240	2.180	2.171

PRECISION

Intra-assay Precision (Precision within an assay)

Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

Three samples of known concentration were tested in forty separate assays to assess inter-assay precision. Assays were performed by at least three technicians using two lots of components.

Sample	Intra-Assay Precision			Inter-Assay Precision		
	1	2	3	1	2	3
n	20	20	20	40	40	40
Mean (ng/mL)	1.26	4.58	6.29	1.23	4.82	6.76
Standard deviation	0.064	0.168	0.217	0.054	0.198	0.309
CV (%)	5.1	3.7	3.4	4.4	4.1	4.6

RECOVERY

The recovery of human MMP-7 spiked to levels throughout the range of the assay in various matrices was evaluated.

Sample Type	Average % Recovery	Range
Cell culture media (n=4)	101	96-108%
Urine* (n=2)	99	89-109%

*Samples were diluted prior to assay.

LINEARITY

To assess the linearity of the assay, samples containing and/or spiked with high concentrations of human MMP-7 in various matrices were diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

		Cell culture media* (n=4)	Serum* (n=5)	Heparin plasma* (n=5)	Saliva* (n=2)	Urine (n=2)
1:2	Average % of Expected	103	105	105	103	100
	Range (%)	99-108	103-108	102-108	99-107	97-102
1:4	Average % of Expected	101	106	108	98	99
	Range (%)	97-105	102-113	104-111	88-107	93-104
1:8	Average % of Expected	99	100	102	99	100
	Range (%)	94-106	93-106	97-106	92-106	96-103
1:16	Average % of Expected	97	—	—	—	93
	Range (%)	90-108	—	—	—	88-97

*Samples were diluted prior to assay as directed in the Sample Preparation section.

SENSITIVITY

Forty assays were evaluated and the minimum detectable dose (MDD) of human MMP-7 ranged from 0.005-0.094 ng/mL. The mean MDD was 0.016 ng/mL.

The MDD was determined by adding two standard deviations to the mean O.D. value of twenty zero standard replicates and calculating the corresponding concentration.

CALIBRATION

This immunoassay is calibrated against a highly purified NS0-expressed recombinant human Pro-MMP-7 produced at R&D Systems®.

SAMPLE VALUES

Serum/Plasma/Saliva/Urine - Samples from apparently healthy volunteers were evaluated for the presence of human MMP-7 in this assay. No medical histories were available for the donors used in this study.

Sample Type	Mean (ng/mL)	Range (ng/mL)	Standard Deviation (ng/mL)
Serum (n=62)	2.29	1.07-4.40	0.71
Heparin plasma (n=36)	2.38	1.10-4.59	0.76
Saliva (n=6)	14.7	3.80-28.3	10.7
Urine (n=7)	2.89	0.17-7.01	2.62

Cell Culture Supernates - Human peripheral blood mononuclear cells (5×10^6 cells/mL) were cultured in DMEM supplemented with 5% fetal bovine serum, 50 μ M β -mercaptoethanol, 2 mM L-glutamine, 100 U/mL penicillin, and 100 μ g/mL streptomycin sulfate. The cells were cultured unstimulated or stimulated with 10 μ g/mL PHA. Aliquots of the cell culture supernates were removed on days 1 and 5 and assayed for levels of human MMP-7.

Condition	Day 1 (ng/mL)	Day 5 (ng/mL)
Unstimulated	ND	21.8
Stimulated	ND	0.93

ND=Non-detectable

SPECIFICITY

This assay recognizes natural and recombinant human MMP-7.

The factors listed below were prepared at 200 ng/mL in calibrator diluent and assayed for cross-reactivity. Preparations of the following factors prepared at 200 ng/mL in a mid-range recombinant human MMP-7 control were assayed for interference. No significant cross-reactivity or interference was observed.

Recombinant human:

MMP-1
MMP-2
MMP-3
MMP-8
MMP-9
MMP-10
MMP-13
TIMP-1
TIMP-2
TIMP-3
TIMP-4

Recombinant mouse:

MMP-9

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PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

12								
11								
10								
9								
8								
7								
6								
5								
4								
3								
2								
1								
	A	B	C	D	E	F	G	H

NOTES

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