

Quantikine[®] ELISA

Human CXCL1/GRO α Immunoassay

Catalog Number DGR00

SGR00

PDGR00

For the quantitative determination of human Growth-Regulated protein alpha (GRO α) concentrations in cell culture supernates, serum, and plasma.

Note: The standard reconstitution method has changed. Please read this package insert in its entirety before using this product.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

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INTRODUCTION

The term chemokine refers to a large superfamily of small (8-10 kDa), inducible, secreted, pro-inflammatory cytokines. These cytokines are characterized by four conserved cysteine residues and nearly all of these cytokines have now been shown to chemoattract and activate different leukocyte subsets. The chemokine family of cytokines can be divided into four subfamilies based on the positioning of the cysteine residues. In the chemokine α or "CXC" subfamily, the first two cysteine residues are separated by a variable amino acid (aa). In the chemokine β or "CC" subfamily, the first two cysteine residues are adjacent. Some of the better known chemokine α family members are IL-8, GRO, platelet factor 4, β -thromboglobulin and IP-10. Examples of chemokine β family members include RANTES, MIP-1 α , MIP-1 β , MCP-1, and I-309. The genes for all of the human chemokine α family members have been mapped to chromosome 4q while those for chemokine β family members are closely linked on human chromosome 17q. Chemokines act primarily on leukocytes, regulating their trafficking. In general, chemokine α family members preferentially chemoattract and activate neutrophils, whereas chemokine β family members are chemotactic for monocytes and T cells (1-3).

GRO α , or CXCL1, was initially discovered as a growth-regulated gene that is overexpressed constitutively in tumorigenic cells (4) and transcribed in normal cells only during growth stimulation (5). Independently, a melanoma growth-stimulating activity (MGSA) was cloned from a human melanoma cell line, Hs294T, and found to be identical to human GRO α (6). In addition to the initially cloned GRO α , two additional human GRO genes, named GRO β or MIP-2 α and GRO γ or MIP-2 β , which share 90% and 86% amino acid (aa) sequence homology, respectively, with GRO α have been identified (7, 8). GRO α expression is inducible by PDGF and/or by a variety of inflammatory mediators, such as IL-1 and TNF, in monocytes, fibroblasts, melanocytes, mammary epithelial cells, and umbilical vein endothelial cells (4). GRO α is also one of several neutrophil-activating factors extracted from human psoriatic scales (9).

The GRO α cDNA encodes a 107 aa precursor protein from which the N-terminal 34 aa residues are cleaved to generate the 73 aa residue mature GRO α . There are no potential N-linked glycosylation sites in the aa sequence nor does the mature protein appear to undergo any post-translational modifications. GRO α exhibits growth stimulatory activity for the human Hs294T melanoma cell line from which it was initially isolated. As a chemokine α member, GRO α , like IL-8, is a potent chemoattractant for human neutrophils and stimulates neutrophil degranulation and enzyme release from cytochalasin B-treated human neutrophils (4). It has been demonstrated that GRO α can cause a decrease in the expression of interstitial collagens by rheumatoid synovial fibroblasts (10). In *in vivo* experiments, GRO α has been shown to induce an acute inflammatory reaction when injected into the foot-pads of endotoxin resistant mice (4).

GRO α can bind with high affinity to the type II IL-8 receptor, also called IL-8 RB. However, it will only bind with low affinity to the type I IL-8 receptor (IL-8 RA) (11). Both IL-8 RA and IL-8 RB, like other chemokine receptors, are members of the superfamily of rhodopsin-like, G-protein-coupled receptors containing seven transmembrane domains. A novel GRO α -specific receptor that does not bind IL-8 has also been identified in synovial fibroblasts and Hs294T cells (10, 12).

The Quantikine Human CXCL1/GRO α Immunoassay is a 2.75-4.5 hour solid phase ELISA designed to measure human GRO α in cell culture supernates, serum, and plasma. It contains *E. coli*-expressed recombinant human GRO α and antibodies raised against the recombinant protein. This immunoassay has been shown to accurately quantitate the recombinant factor. Results obtained using natural human GRO α showed linear curves that were parallel to the standard curves obtained using the Quantikine kit standards. These results indicate that this kit can be used to determine relative mass values for natural human GRO α .

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for human GRO α has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any GRO α present is bound by the immobilized antibody. After washing away any unbound substances, an enzyme-linked polyclonal antibody specific for human GRO α is added to the wells. Following a wash to remove any unbound antibody-enzyme reagent, a substrate solution is added to the wells and color develops in proportion to the amount of GRO α bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- It is important that the Calibrator Diluent selected for the standard curve be consistent with the samples being assayed.
- If samples generate values higher than the highest standard, dilute the samples with the appropriate Calibrator Diluent and repeat the assay.
- Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- Variations in sample collection, processing, and storage may cause sample value differences.
- This assay is designed to eliminate interference by other factors present in biological samples. Until all factors have been tested in the Quantikine Immunoassay, the possibility of interference cannot be excluded.

TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- When using an automated plate washer, adding a 30 second soak period following the addition of Wash Buffer, and/or rotating the plate 180 degrees between wash steps may improve assay precision.
- Substrate Solution should remain colorless until added to the plate. Keep Substrate Solution protected from light. Substrate Solution should change from colorless to gradations of blue.
- Stop Solution should be added to the plate in the same order as the Substrate Solution. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.

MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

PART	PART #	CATALOG # DGR00	CATALOG # SGR00	DESCRIPTION	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Human GRO α Microplate	890150	1 plate	6 plates	96 well polystyrene microplate (12 strips of 8 wells) coated with a monoclonal antibody specific for human GRO α .	Return unused wells to the foil pouch containing the desiccant pack. Reseal along entire edge of zip-seal. May be stored for up to 1 month at 2-8 °C.*
Human GRO α Standard	890152	1 vial	6 vials	Recombinant human GRO α in a buffered protein base with preservatives; lyophilized. <i>Refer to the vial label for reconstitution volume.</i>	Aliquot and store for up to 1 month at ≤ -20 °C in a manual defrost freezer.* Avoid repeated freeze-thaw cycles.
Human GRO α Conjugate	890151	1 vial	6 vials	21 mL/vial of a polyclonal antibody specific for human GRO α conjugated to horseradish peroxidase with preservatives.	May be stored for up to 1 month at 2-8 °C.*
Assay Diluent RD1U	895138	1 vial	6 vials	6 mL/vial of a buffered protein base with preservatives. <i>May appear cloudy and contain a precipitate. Mix well before and during use. For serum/plasma samples.</i>	
Calibrator Diluent RD5K	895119	1 vial	6 vials	21 mL/vial of a buffered protein base with preservatives. <i>For cell culture supernate samples.</i>	
Calibrator Diluent RD6-69	896012	1 vial	6 vials	21 mL/vial of diluted animal serum with preservatives. <i>For serum/plasma samples.</i>	
Wash Buffer Concentrate	895003	1 vial	6 vials	21 mL/vial of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>	
Color Reagent A	895000	1 vial	6 vials	12 mL/vial of stabilized hydrogen peroxide.	
Color Reagent B	895001	1 vial	6 vials	12 mL/vial of stabilized chromogen (tetramethylbenzidine).	
Stop Solution	895032	1 vial	6 vials	6 mL/vial of 2 N sulfuric acid.	
Plate Sealers	N/A	4 strips	24 strips	Adhesive strips.	

* Provided this is within the expiration date of the kit.

DGR00 contains sufficient materials to run an ELISA on one 96 well plate.

SGR00 (SixPak) contains sufficient materials to run ELISAs on six 96 well plates.

This kit is also available in a PharmPak (R&D Systems, Catalog # PDGR00). PharmPaks contain sufficient materials to run ELISAs on 50 microplates. Specific vial counts of each component may vary. Please refer to the literature accompanying your order for specific vial counts.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 2-8 °C centrifuge for plasma preparation.
- 2-8 °C incubator for assay procedure.
- 500 mL graduated cylinder.
- Test tubes for dilution of standards.
- Human GRO α Controls (optional; R&D Systems, Catalog # QC101).

PRECAUTIONS

The Stop Solution provided with this kit is an acid solution.

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Color Reagent B may cause skin, eye, and respiratory irritation. Avoid breathing fumes.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Please refer to the MSDS on our website prior to use.

SAMPLE COLLECTION & STORAGE

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Caution: *Normal human serum added to cell culture media may contain GRO α . For best results, do not use human serum for growth of cell cultures if assaying for GRO α production. Because there is no species cross-reactivity of this kit, human GRO α levels in culture media containing 10% bovine or fetal bovine serum can be assayed without interference.*

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Platelet-poor Plasma - Collect plasma on ice using EDTA, heparin, or citrate as an anticoagulant. Centrifuge at 1000 x g within 30 minutes of collection. An additional centrifugation step of the plasma at 10,000 x g for 10 minutes at 2-8 °C is recommended for complete platelet removal. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

GRO α appears to be present in platelet granules and released upon platelet activation. Therefore, to measure circulating levels of GRO α , platelet-poor plasma should be collected for measurement. It should be noted that many protocols for plasma preparation, including procedures recommended by the Clinical and Laboratory Standards Institute (CLSI), result in incomplete removal of platelets from blood.

REAGENT PREPARATION

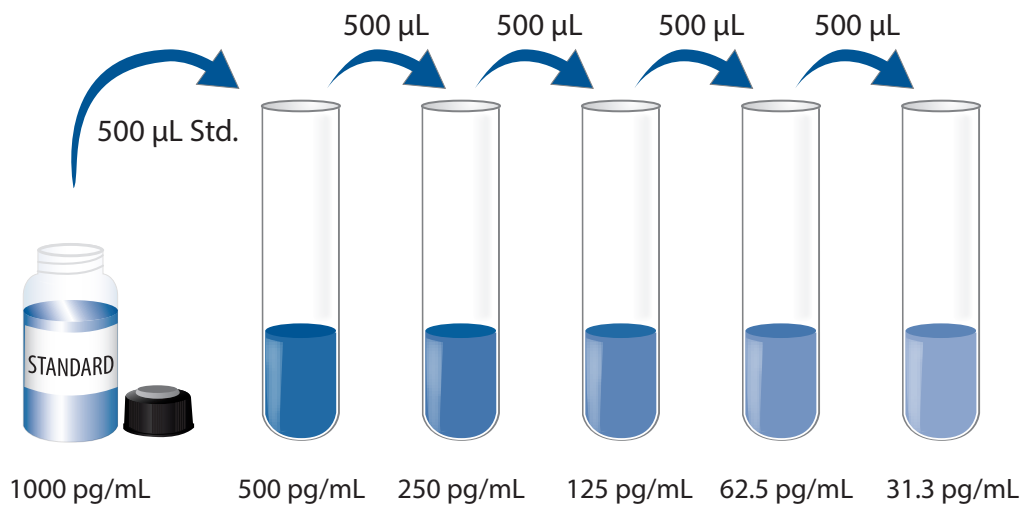
Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to deionized or distilled water to prepare 500 mL of Wash Buffer.

Substrate Solution - Color Reagents A and B should be mixed together in equal volumes within 15 minutes of use. Protect from light. 200 μ L of the resultant mixture is required per well.

Human GRO α Standard - Refer to the vial label for reconstitution volume. Reconstitute the Human GRO α Standard with Calibrator Diluent RD5K (*for cell culture supernate samples*) or Calibrator Diluent RD6-69 (*for serum/plasma samples*). This reconstitution produces a stock solution of 1000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

Pipette 500 μ L of the appropriate Calibrator Diluent into each tube. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The undiluted GRO α Standard (1000 pg/mL) serves as the high standard. The appropriate Calibrator Diluent serves as the zero standard (0 pg/mL).



ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that all standards, samples, and controls be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. **For Serum/Plasma Samples Only:** Add 50 μL of Assay Diluent RD1U to each well. *Contains a precipitate. Mix well before and during use.*
4. Add 200 μL of Standard, sample, or control per well. Cover with the adhesive strip provided.
For Cell Culture Supernate Samples: Incubate for 1.5 hours at room temperature.
For Serum/Plasma Samples: Incubate for 2 hours at room temperature.
5. Aspirate each well and wash, repeating the process twice for a total of three washes. Wash by filling each well with Wash Buffer (400 μL) using a squirt bottle, manifold dispenser, or autowasher. **After 1 minute, empty wells by aspiration or decanting.** Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper toweling.
6. Add 200 μL of Human GRO α Conjugate to each well. Cover with a new adhesive strip.
For Cell Culture Supernate Samples: Incubate for 1 hour at **2-8 °C**.
For Serum/Plasma Samples: Incubate for 2 hours at **2-8 °C**.
7. Repeat the aspiration/wash as in step 5.
8. Add 200 μL of Substrate Solution to each well. **Protect from light.**
For Cell Culture Supernate Samples: Incubate for 15 minutes at room temperature.
For Serum/Plasma Samples: Incubate for 30 minutes at room temperature.
9. Add 50 μL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green or the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
10. Determine the optical density of each well within 30 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

CALCULATION OF RESULTS

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density (O.D.).

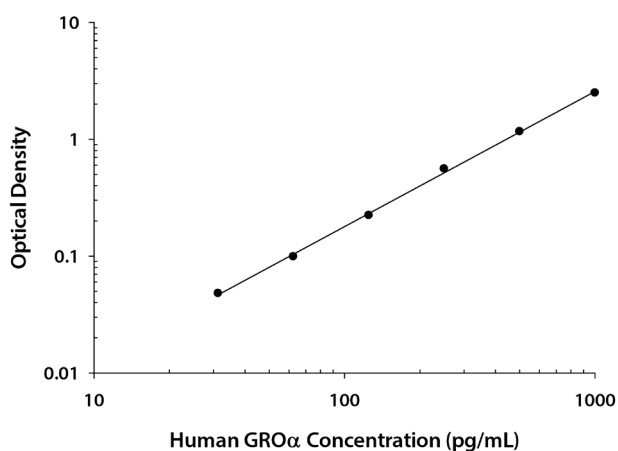
Create a standard curve by reducing the data using computer software capable of generating a log/log curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on a log/log graph. The data may be linearized by plotting the log of the human GRO α concentrations versus the log of the O.D. on a linear scale, and the best fit line can be determined by regression analysis.

If the samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

TYPICAL DATA

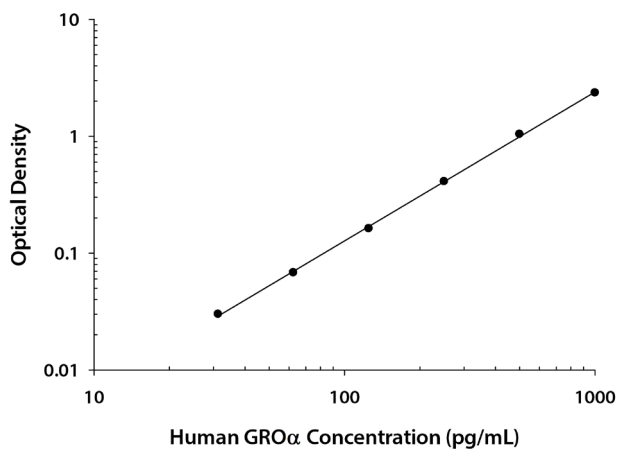
These standard curves are provided for demonstration only. A standard curve should be generated for each set of samples assayed.

CELL CULTURE SUPERNATE ASSAY



(pg/mL)	O.D.	Average	Corrected
0	0.009 0.013	0.011	—
31.3	0.059 0.059	0.059	0.048
62.5	0.108 0.111	0.110	0.099
125	0.231 0.237	0.234	0.223
250	0.568 0.570	0.569	0.558
500	1.162 1.191	1.176	1.165
1000	2.465 2.546	2.506	2.495

SERUM/PLASMA ASSAY



(pg/mL)	O.D.	Average	Corrected
0	0.020 0.023	0.022	—
31.3	0.051 0.052	0.052	0.030
62.5	0.090 0.090	0.090	0.068
125	0.184 0.184	0.184	0.162
250	0.428 0.435	0.432	0.410
500	1.055 1.071	1.063	1.041
1000	2.352 2.394	2.373	2.351

PRECISION

Intra-assay Precision (Precision within an assay)

Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

Three samples of known concentration were tested in twenty separate assays to assess inter-assay precision. Assays were performed by at least three technicians using two lots of components.

CELL CULTURE SUPERNATE ASSAY

Sample	Intra-Assay Precision			Inter-Assay Precision		
	1	2	3	1	2	3
n	20	20	20	20	20	20
Mean (pg/mL)	81.4	318	734	87.4	320	758
Standard deviation	3.7	7.4	55.0	11.9	27.6	62.7
CV (%)	4.5	2.3	7.5	13.6	8.6	8.3

SERUM/PLASMA ASSAY

Sample	Intra-Assay Precision			Inter-Assay Precision		
	1	2	3	1	2	3
n	20	20	20	20	20	20
Mean (pg/mL)	61.4	251	736	62.8	252	734
Standard deviation	2.2	6.0	20.0	3.6	11.9	38.9
CV (%)	3.6	2.4	2.7	5.7	4.7	5.3

RECOVERY

The recovery of human GRO α spiked to levels throughout the range of the assay in various matrices was evaluated.

Sample Type	Average % Recovery	Range
Cell culture media	97	81-108%
Serum	98	87-108%
EDTA plasma	93	86-102%
Heparin plasma	95	83-109%
Citrate plasma	90	80-98%

SENSITIVITY

The minimum detectable dose (MDD) of human GRO α is typically less than 10 pg/mL.

The MDD was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

LINEARITY

To assess linearity of the assay, samples containing and/or spiked with high concentrations of human GRO α were diluted with Calibrator Diluent to produce samples with values within the dynamic range of the assay.

		Cell culture media (n=4)	Serum (n=4)	EDTA plasma (n=4)	Heparin plasma (n=4)	Citrate plasma (n=4)
1:2	Average % of Expected	107	103	95	112	99
	Range (%)	99-112	97-107	93-100	110-116	97-103
1:4	Average % of Expected	109	103	92	115	96
	Range (%)	98-117	92-111	87-100	108-122	92-101
1:8	Average % of Expected	106	102	88	115	92
	Range (%)	93-113	90-112	82-96	105-125	83-99
1:16	Average % of Expected	111	104	90	116	95
	Range (%)	96-119	93-119	83-101	105-129	87-101

CALIBRATION

This immunoassay is calibrated against a highly purified *E. coli*-expressed recombinant human GRO α produced at R&D Systems.

The NIBSC/WHO GRO α interim reference preparation 92/722 which was intended as a potency standard, was evaluated in this kit. The interim reference material parallels the Quantikine standard curve. To convert sample values obtained with the Quantikine Human CXCL1/GRO α kit to approximate NIBSC nominally assigned mass values use the equation below.

NIBSC (92/722) approximate value (pg/mL) = 0.32 x Quantikine Human GRO α value (pg/mL)

SAMPLE VALUES

Serum/Platelet-poor Plasma - Serum and platelet-poor plasma samples from apparently healthy volunteers were evaluated for the presence of human GRO α in this assay. No medical histories were available for the donors used in this study. Forty serum samples measured between 32-180 pg/mL with a mean of 93 pg/mL. Ten platelet-poor plasma samples were tested, and all measured less than the lowest Human GRO α Standard, 31.3 pg/mL.

Cell Culture Supernates - Human peripheral blood mononuclear cells (1×10^6 cells/mL) were cultured in RPMI supplemented with 10% fetal calf serum, 50 μ M β -mercaptoethanol, 2 mM L-glutamine, 100 U/mL penicillin, and 100 μ g/mL streptomycin sulfate and stimulated with 10 μ g/mL PHA. Aliquots of the culture supernates were removed on days 1, 3, and 5 and assayed for levels of human GRO α .

Condition	Day 1 (pg/mL)	Day 3 (pg/mL)	Day 5 (pg/mL)
Unstimulated	885	1611	1869
Stimulated	18,440	50,810	46,090

It is important to note that sample collection and handling procedures will have a significant impact on measured GRO α levels. Refer to the Sample Collection and Storage section for the recommended sample collection procedure.

SPECIFICITY

This assay recognizes natural and recombinant human GRO α .

The factors listed below were prepared at 50 ng/mL in Calibrator Diluent and assayed for cross-reactivity. Preparations of the following factors at 50 ng/mL in a mid-range recombinant human GRO α control were assayed for interference. No significant cross-reactivity or interference was observed.

Recombinant human:

CXCL2/GRO β
CXCL5/ENA-78
CXCL7/NAP-2
CXCL8/IL-8
CCL2/MCP-1
CCL3/MIP-1 α
CCL4/MIP-1 β
CCL5/RANTES

Recombinant mouse:

CCL3/MIP-1 α
CCL4/MIP-1 β
CXCL1/KC

Recombinant rat:

CXCL1/CINC-1

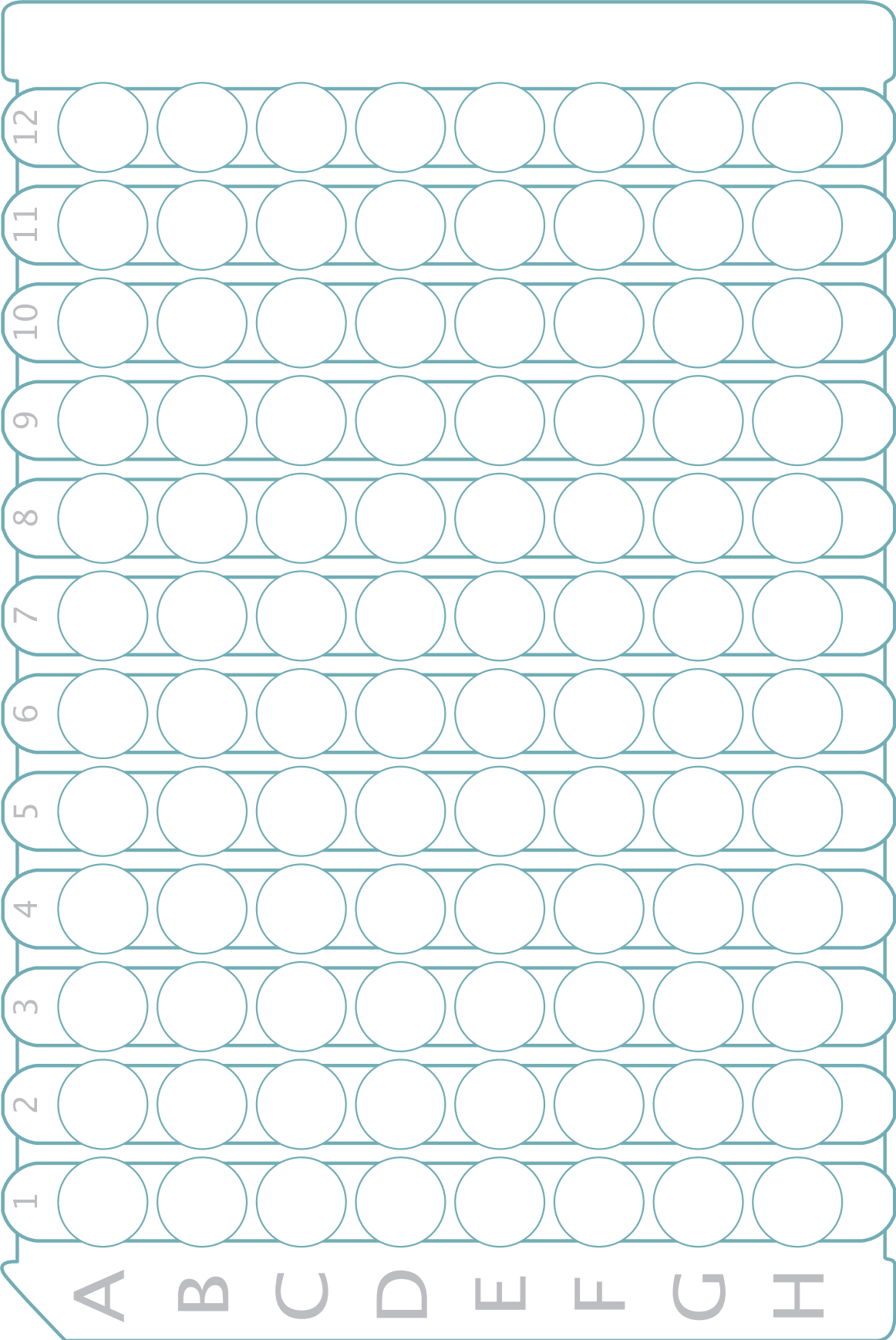
Recombinant human GRO γ cross-reacts approximately 0.4% in this assay.

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PLATE LAYOUT

Use this plate layout to record standards and samples assayed.



NOTES

NOTES

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