

Proteome Profiler™ Array

Human Soluble Receptor Array Kit Non-Hematopoietic Panel

Catalog Number ARY012

For the parallel determination of the relative levels of soluble receptors and related proteins in non-hematopoietic cells.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

TABLE OF CONTENTS

SECTION	PAGE
INTRODUCTION	1
PRINCIPLE OF THE ASSAY	1
TECHNICAL HINTS	1
MATERIALS PROVIDED & STORAGE CONDITIONS	2
OTHER SUPPLIES REQUIRED	3
PRECAUTIONS.....	3
SAMPLE COLLECTION & STORAGE	4
REAGENT PREPARATION	5
ARRAY PROCEDURE	6
DATA ANALYSIS	8
PROFILING SOLUBLE RECEPTORS IN CELL CULTURE SUPERNATES	9
PROFILING SOLUBLE RECEPTORS IN CELL LYSATES	11
PROFILING SOLUBLE RECEPTORS IN SERUM	13
APPENDIX	14

Manufactured and Distributed by:

USA R&D Systems, Inc.

614 McKinley Place NE, Minneapolis, MN 55413

TEL: 800 343 7475 612 379 2956

FAX: 612 656 4400

E-MAIL: info@bio-techne.com

Distributed by:

Europe | Middle East | Africa Bio-Techne Ltd.

19 Barton Lane, Abingdon Science Park

Abingdon OX14 3NB, UK

TEL: +44 (0)1235 529449

FAX: +44 (0)1235 533420

E-MAIL: info.emea@bio-techne.com

China Bio-Techne China Co., Ltd.

Unit 1901, Tower 3, Raffles City Changning Office,

1193 Changning Road, Shanghai PRC 200051

TEL: +86 (21) 52380373 (400) 821-3475

FAX: +86 (21) 52371001

E-MAIL: info.cn@bio-techne.com

INTRODUCTION

Analyzing the expression profiles of soluble receptors expressed and released by non-hematopoietic cells is essential for understanding the roles these molecules play in activation of endothelial cells. The Human Soluble Receptor Array Kit is a rapid, sensitive, and economical tool to simultaneously detect changes in proteins between samples. The relative expression levels of 119 soluble receptors and related proteins can be determined without performing numerous immunoprecipitations or Western Blots. Each capture and detection antibody, directed to the extracellular domain, was carefully selected using both natural and recombinant proteins.

PRINCIPLE OF THE ASSAY

Capture and control antibodies have been spotted in duplicate on nitrocellulose membranes. Cell culture supernate, serum, or cell lysate samples are diluted and incubated overnight with each array part of the Non-Hematopoietic Panel. This panel is comprised of the Non-Hematopoietic Array and the Common Analytes Array ([R&D Systems®](#), # ARY011). The membranes are washed to remove unbound material followed by incubation with their specific cocktail of biotinylated detection antibodies. Streptavidin-HRP and chemiluminescent detection reagents are applied, and a signal is produced at each capture spot corresponding to the amount of protein bound. Refer to the Appendix for the coordinates of analytes and controls.

TECHNICAL HINTS

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- This kit should not be used beyond the expiration date on the kit label.
- **Do not mix or substitute reagents with those from other lots or sources. Substitution of some high intensity chemiluminescent reagents for Chemi Reagents 1 and 2 may cause either increased background or diminished signal depending on the reagent.**
- Any variation in sample handling, buffers, operator, pipetting technique, washing technique, and incubation time or temperature can alter the performance of the kit.
- The Human Soluble Receptor Array membranes are validated for single use only.
- Always use gloved hands and flat-tipped tweezers to handle the membranes.
- Pick up the membranes from the edge on the side with the identification number avoiding the area with the printed antibodies.
- A thorough and consistent wash technique is essential for proper assay performance. Individual arrays should be washed in separate containers to minimize background. Wash Buffer should be removed completely from the membranes before proceeding to the next step.
- Do not allow the membranes to dry out. This will cause high background.
- Avoid microbial contamination of reagents and buffers.
- For a procedure demonstration video, visit: www.RnDSystems.com/ProteomeProfilerVideo.

MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

PART	PART #	DESCRIPTION	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Human Soluble Receptor Array Non-Hematopoietic Panel	893367	4 nitrocellulose membranes: 2 Part N each containing 62 different antibodies; and 2 Part C each containing 57 different antibodies printed in duplicate.	Return unused membranes to the foil pouch containing the desiccant pack. Reseal along entire edge of the zip-seal. May be stored for up to 3 months at 2-8 °C.*
Lysis Buffer 17	895943	21 mL of a non-denaturing buffered solution	Prepare fresh for each use.
Array Buffer 1	895477	2 vials (21 mL/vial) of a buffered protein base with preservatives.	May be stored for up to 3 months at 2-8 °C.*
Array Buffer 8	895050	21 mL of a buffered protein base with preservatives.	
Wash Buffer Concentrate	895003	2 vials (21 mL/vial) of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>	
Detection Antibody Cocktail N	893366	1 vial of a biotinylated antibody cocktail; lyophilized; red cap.	
Detection Antibody Cocktail C	893369	1 vial of a biotinylated antibody cocktail; lyophilized; blue cap.	
Streptavidin-HRP	893019	200 µL of streptavidin conjugated to horseradish-peroxidase.	
Chemi Reagent 1	894287	1 vial (2.5 mL)	
Chemi Reagent 2	894288	1 vial (2.5 mL)	Store at room temperature.
4-Well Rectangular Multi-dish	607544	Clear 4-well rectangular multi-dish.	
Transparency Overlay Template	607677	1 transparency overlay template for coordinate reference.	

* Provided this is within the expiration date of the kit.

Note: Additional wash buffer is available for purchase ([R&D Systems®](#), # [WA126](#)).

OTHER SUPPLIES REQUIRED

- Aprotinin ([Tocris™, # 4139](#))
- Leupeptin hemisulfate ([Tocris, # 1167](#))
- Pepstatin A ([Tocris, # 1190](#))
- Pipettes and pipette tips
- Gloves
- Phosphate-Buffered Saline (PBS) ([R&D Systems®, # RB01](#))
- Deionized or distilled water
- Rocking platform shaker
- Microcentrifuge
- A plastic container with the capacity to hold 50 mL (for washing the arrays)
- Plastic transparent sheet protector (trimmed to 10 cm x 12 cm and open on three sides)
- Plastic wrap
- Paper towels
- Absorbent lab wipes
- Autoradiography cassette
- Film developer
- X-ray film
- Flat-tipped tweezers
- Flatbed scanner with transparency adapter capable of transmission mode
- Computer capable of running image analysis software and Microsoft® Excel

PRECAUTIONS

High levels of some array analytes are found in saliva. It is recommended that a mask and gloves be used to protect kit reagents from contamination.

Chemi Reagents 1 and 2 contain Boric Acid which is suspected of damaging fertility or the unborn child.

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

SAMPLE COLLECTION & STORAGE

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Since the Human Soluble Receptor Array detects relative expression levels of individual analytes, it is important to include appropriate control samples.

Note: Sample amount may be empirically adjusted to attain optimal sensitivity with minimal background. Suggested starting ranges to use for each array part are: 200-500 μ L for cell culture supernates, 100-300 μ g for cell lysates, and 10-50 μ L for serum samples.

Cell Culture Supernates - Remove particulates by centrifugation. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Cell Lysates - Rinse cells with PBS, making sure to remove any remaining PBS before adding lysis buffer. Solubilize cells at 1×10^7 cells/mL in Lysis Buffer 17 (prepared as described in the Reagent Preparation section). Pipette up and down to resuspend and rock the lysates gently at 2-8 °C for 30 minutes. Microcentrifuge at 14,000 x g for 5 minutes, and transfer the supernate into a clean test tube. Quantitation of sample protein concentrations using a total protein assay is recommended. Assay immediately or aliquot and store at ≤ -70 °C. Avoid repeated freeze-thaw cycles.

Serum - Allow blood samples to clot for 30 minutes at room temperature before centrifuging for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Note: *High levels of some proteins are found in saliva. It is recommended that a mask and gloves be used to protect kit reagents from contamination.*

Human Soluble Receptor Array: Non-Hematopoietic Panel - Four nitrocellulose membranes; the Non-Hematopoietic Arrays (Part N; array numbers begin with N) contain 62 antibodies printed in duplicate. The Common Analyte Arrays (Part C; array numbers begin with C) contain 57 antibodies printed in duplicate. Both arrays contain four sample control antibodies. The two array parts should be used in parallel to generate a complete profile in one experiment.

Handle membranes only with gloved hands and flat-tipped tweezers.

Detection Antibody Cocktail N (red cap) - Before use, reconstitute Detection Antibody Cocktail N in 100 μ L of deionized or distilled water.

Detection Antibody Cocktail C (blue cap) - Before use, reconstitute Detection Antibody Cocktail C in 100 μ L of deionized or distilled water.

1X Array Buffer 8/1 - Dilute 1 mL of Array Buffer 8 into 9 mL of Array Buffer 1.

Lysis Buffer 17 - Add 10 μ g/mL Aprotinin, 10 μ g/mL Leupeptin hemisulfate, and 10 μ g/mL Pepstatin A to the volume of lysis buffer required for cell lysate preparation.

1X Wash Buffer - If crystals have formed in the concentrate, warm the bottles to room temperature and mix gently until the crystals have completely dissolved. Add 40 mL of Wash Buffer Concentrate to 960 mL of deionized or distilled water to prepare 1000 mL of 1X Wash Buffer.

Chemi Reagent Mix - Chemi Reagents 1 and 2 should be mixed in equal volumes within 15 minutes of use. **Protect from light. 1 mL of the resultant mixture is required for each membrane.**

ARRAY PROCEDURE

Bring all reagents to room temperature before use. Keep samples on ice. To avoid contamination, wear gloves while performing the procedures.

Note: *High levels of some proteins are found in saliva. It is recommended that a mask and gloves be used to protect kit reagents from contamination.*

1. Prepare all reagents and samples as directed in the previous sections.
2. The Human Soluble Receptor Array: Non-Hematopoietic Panel is divided into two parts (N and C). For best results, incubate Parts N and C in aliquots of the same sample but in separate wells of the 4-Well Multi-dish.
3. Pipette 2 mL of 1X Array Buffer 8/1 into each well of the 4-Well Multi-dish to be used. 1X Array Buffer 8/1 serves as a block buffer.
4. Using flat-tip tweezers, remove each membrane to be used from between the protective sheets and place in a well of the 4-Well Multi-dish. The number on the membrane should be facing upward.

Note: *Upon contact with 1X Array Buffer 8/1, the blue dye from the spots will disappear, but the capture antibodies are retained in their specific locations.*

5. Incubate for one hour on a rocking platform shaker. Orient the tray so that each membrane rocks end to end in its well.
6. While arrays are blocking, prepare samples for both parts of the array (N and C) by adding the desired quantity of sample (up to 500 μ L for cell lysates or 1 mL for all other sample types) to 300 μ L of Array Buffer 8. Adjust to a final volume of 3 mL with Array Buffer 1.
7. Aspirate 1X Array Buffer 8/1 from the wells of the 4-Well Multi-dish and add 1.5 mL of the prepared sample to the Part N array and 1.5 mL to the Part C array. Place the lid on the 4-Well Multi-dish.
8. Incubate overnight at 2-8 °C on a rocking platform shaker.

Note: *A shorter incubation time may be used if optimal sensitivity is not required.*

9. Carefully remove each array and place into separate plastic containers with 20 mL of 1X Wash Buffer. The recommended container size for washing is approximately 11 x 8 x 2 cm (L x W x H). Rinse the 4-Well Multi-dish with deionized or distilled water and dry thoroughly.
10. Wash each array with 1X Wash Buffer for 10 minutes on a rocking platform shaker. Repeat two times for a total of three washes.
11. For each Part N array, dilute 30 μ L of reconstituted Detection Antibody Cocktail N (red cap) to 1.5 mL with 1X Array Buffer 8/1. Pipette 1.5 mL per well of diluted Detection Antibody Cocktail N into the 4-Well Multi-dish.
12. Carefully remove each Part N array from its wash container. Allow excess Wash Buffer to drain from the array. Return the array to the 4-Well Multi-dish containing the diluted Detection Antibody Cocktail N.
13. For each Part C array, dilute 30 μ L of reconstituted Detection Antibody Cocktail C (blue cap) to 1.5 mL with 1X Array Buffer 8/1. Pipette 1.5 mL per well of diluted Detection Antibody Cocktail C into the 4-Well Multi-dish.

ARRAY PROCEDURE *CONTINUED*

14. Carefully remove each Part C array from its wash container. Allow excess Wash Buffer to drain from the array. Return the array to the 4-Well Multi-dish containing the diluted Detection Antibody Cocktail C, and cover with the lid.
15. Incubate for 2 hours at room temperature on a rocking platform shaker.
16. Wash the array as described in steps 9 and 10.
17. Dilute the Streptavidin-HRP in 1X Array Buffer 8/1 using the dilution factor on the vial label. Pipette 2 mL of diluted Streptavidin-HRP into each well of the 4-Well Multi-dish.
18. Carefully remove each membrane from its wash container. Allow excess Wash Buffer to drain from the membrane. Return the membrane to the 4-Well Multi-dish containing the diluted Streptavidin-HRP. Cover the wells with the lid.
19. Incubate for 30 minutes at room temperature on a rocking platform shaker.
20. Wash each array as described in steps 9 and 10.

Note: *Complete the remaining steps without interruption.*

21. Carefully remove each membrane from its wash container. Allow excess Wash Buffer to drain from the membrane by blotting the lower edge onto paper towels. Place each membrane on the bottom sheet of the plastic sheet protector with the identification number facing up.
22. Pipette 1 mL of the prepared Chemi Reagent Mix evenly onto each set of membranes.
Note: *Using less than 1 mL of Chemi Reagent Mix per membrane may result in incomplete membrane coverage.*
23. Carefully cover with the top sheet of the plastic sheet protector. Gently smooth out any air bubbles and ensure Chemi Reagent Mix is spread evenly to all corners of each membrane. Incubate for 1 minute.
24. Position paper towels on the top and sides of the plastic sheet protector containing the membranes and carefully squeeze out excess Chemi Reagent Mix.
25. Remove the top plastic sheet protector and carefully lay an absorbent lab wipe on top of the membranes to blot off any remaining Chemi Reagent Mix.
26. Leaving membranes on the bottom plastic sheet protector, cover the membranes with plastic wrap taking care to gently smooth out any air bubbles. Wrap the excess plastic wrap around the back of the sheet protector so that the membranes and sheet protector are completely wrapped.
27. Place the membranes with the identification numbers facing up in an autoradiography film cassette.
Note: *Use an autoradiography cassette that is not used with radioactive isotope detection.*
28. Expose membranes to X-ray film for 1-10 minutes. Multiple exposure times are recommended.

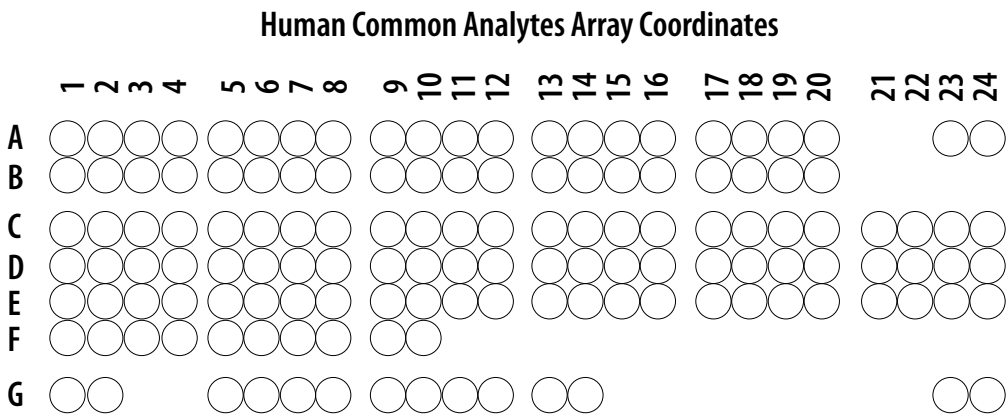
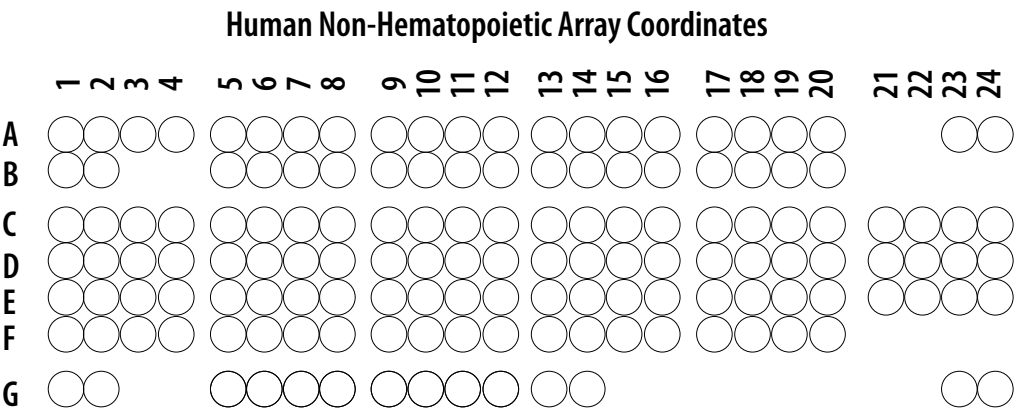
DATA ANALYSIS

The positive signals seen on developed film can be quickly identified by placing the transparency overlay template on the array image and aligning it with the pairs of reference spots in the corners of each array. The stamped identification number on the array should be placed on the left hand side. The Human Non-Hematopoietic Array membranes contain 6 reference spots in the upper left hand corner and the Human Common Analytes Array membranes contain 8 reference spots in the upper left hand corner. The location of reference spots and capture antibodies is listed in the Appendix.

Note: Reference spots are included to align the transparency overlay template and to demonstrate that the array has been incubated with Streptavidin-HRP during the assay procedure.

Pixel densities on developed X-ray film can be collected and analyzed using a transmission-mode scanner and image analysis software.

- 1. Create a template to analyze pixel density in each spot of the array.
- 2. Export signal values to a spreadsheet file for manipulation in a program such as Microsoft Excel.
- 3. Determine the average signal (pixel density) of the pair of duplicate spots representing each protein.
- 4. Subtract an averaged background signal from each spot. Use a signal from a clear area of the array or negative control spots as a background value.
- 5. Compare corresponding signals on different arrays to determine the relative change in protein levels between samples.



These images are not to scale. They are for coordinate reference only.
Please use the transparency overlay for analyte identification.

PROFILING SOLUBLE RECEPTORS IN CELL CULTURE SUPERNATES

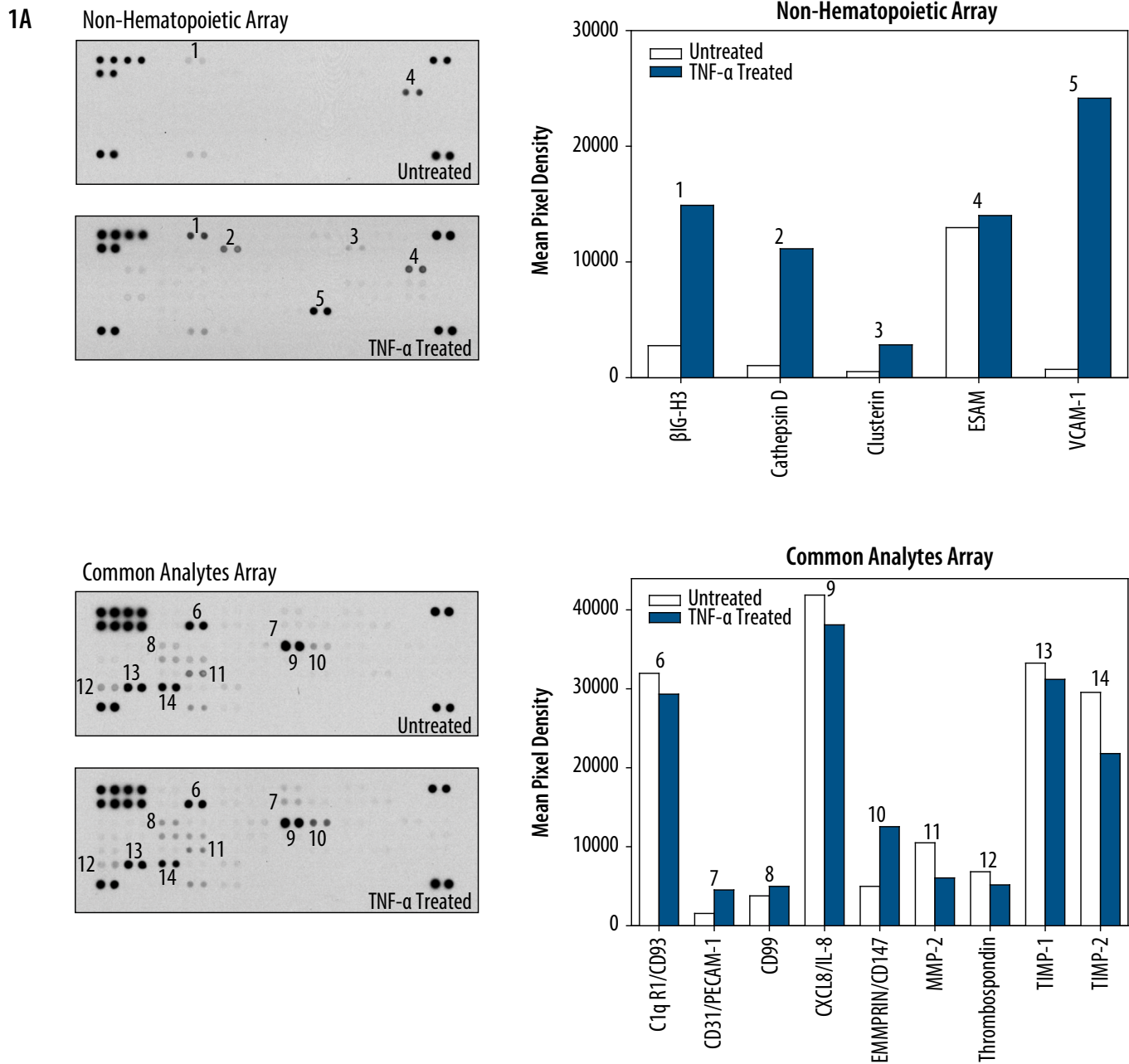


Figure 1A: The Human Soluble Receptor Array detects multiple analytes in cell culture supernates. HUVEC human umbilical vein endothelial cells were untreated or treated with 100 µg/mL of Recombinant Human TNF-α ([R&D Systems®, # 210-TA](#)) for 24 hours. 500 µL of cell culture supernate was run on each array. Data shown are from a 10 minute exposure to X-ray film.

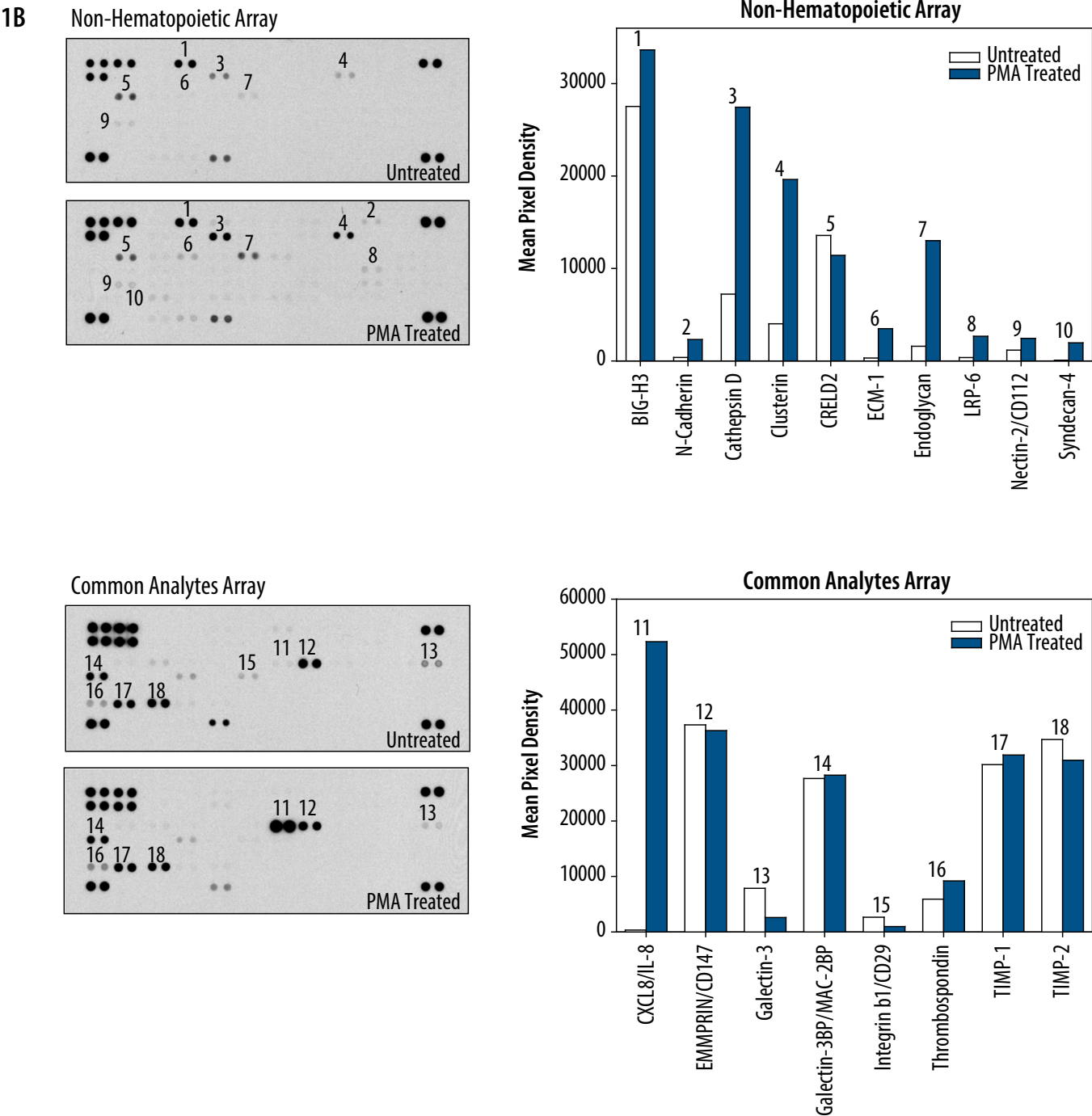


Figure 1B: HeLa human cervical epithelial carcinoma cells were untreated or treated with 0.5 mM PMA for 18 hours. 500 μ L of cell culture supernate was run on each array. Data shown are from a 10 minute exposure to X-ray film.

PROFILING SOLUBLE RECEPTORS IN CELL LYSATES

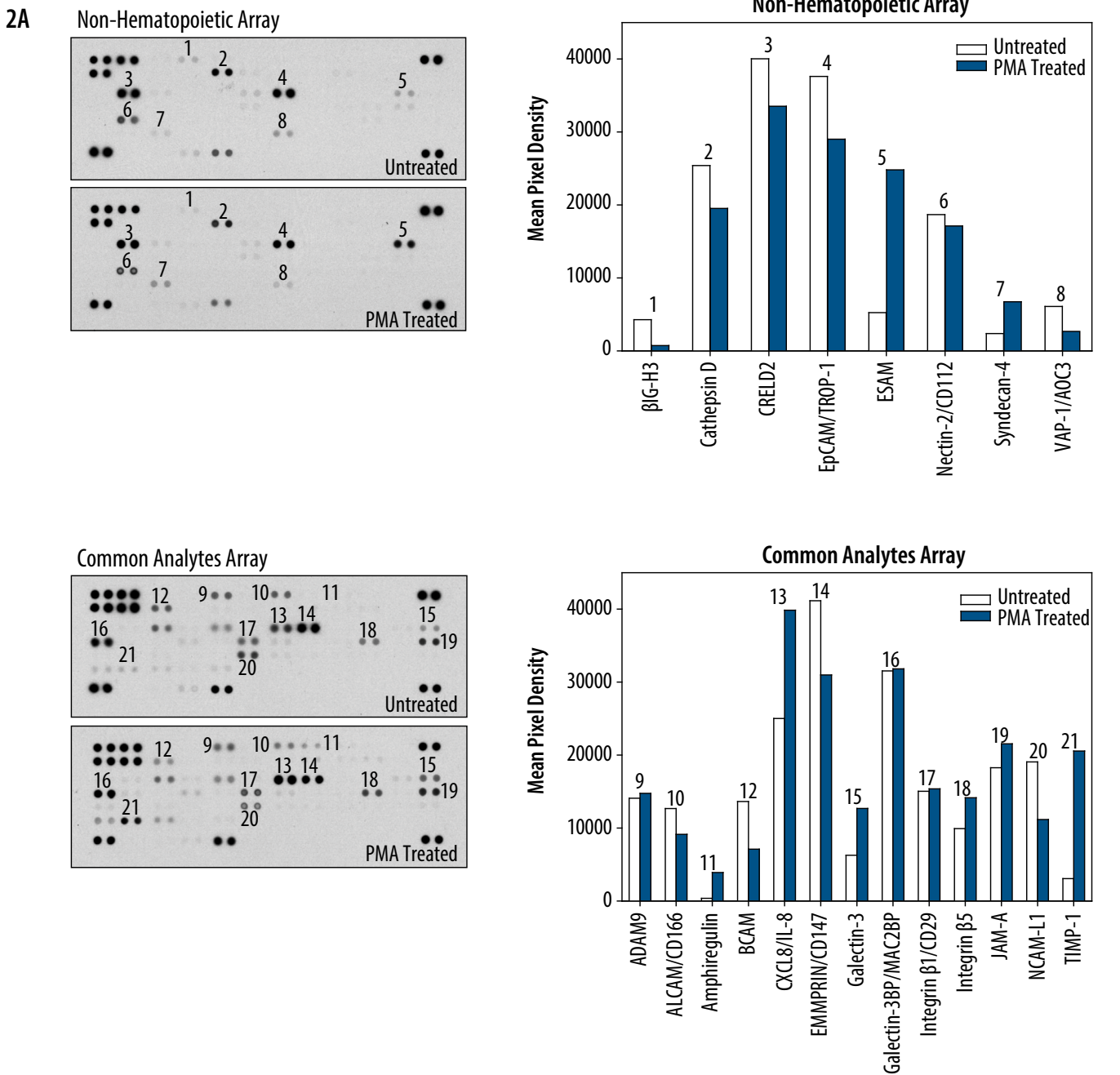


Figure 2A: The Human Soluble Receptor Array detects multiple analytes in cell lysates. HepG2 human hepatocellular carcinoma cells were untreated or treated with 80 nM of PMA for 24 hours. 200 µg of cell lysate was run on each array. Data shown are from a 10 minute exposure to X-ray film

PROFILING SOLUBLE RECEPTORS IN CELL LYSATES *CONTINUED*

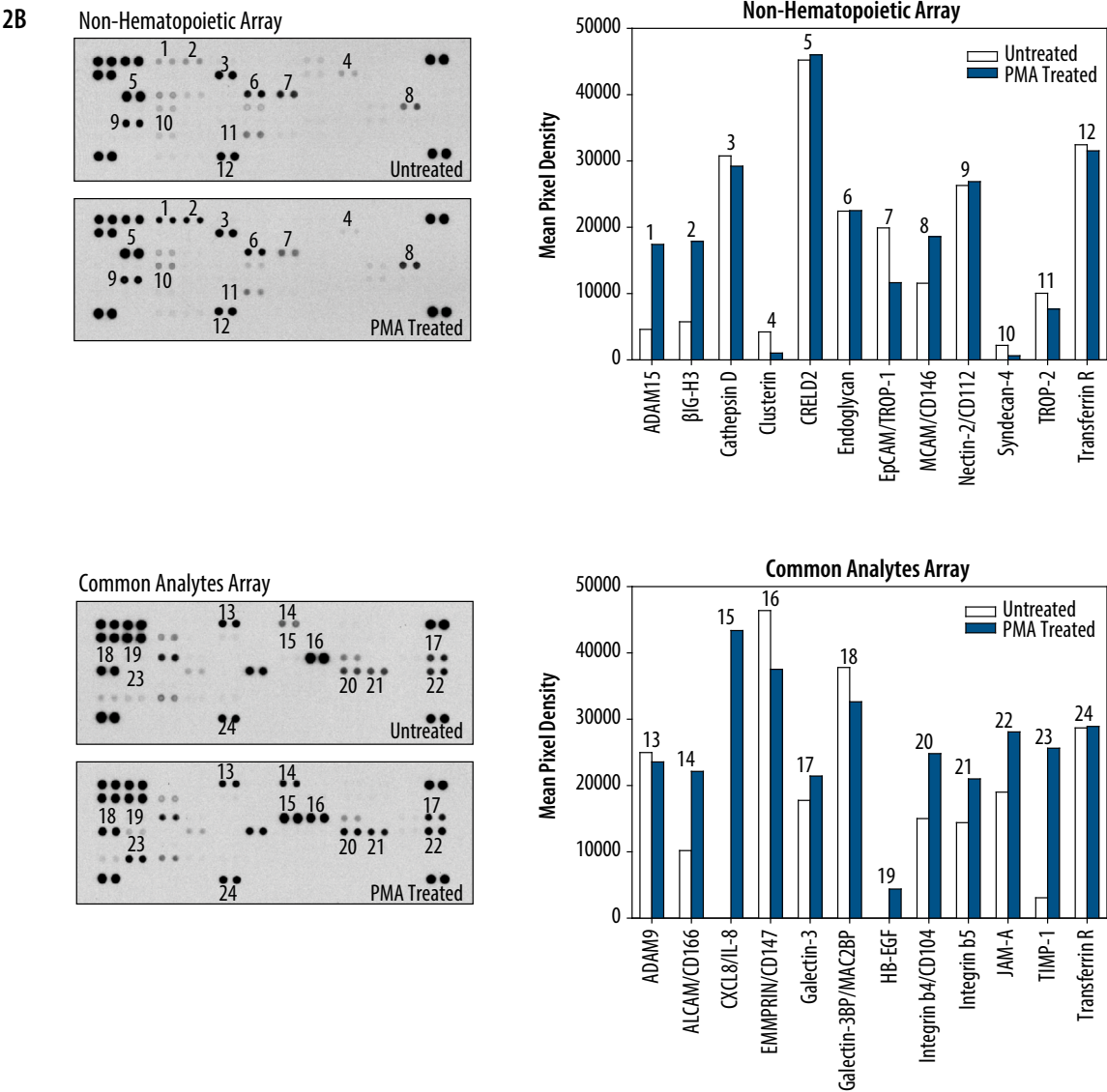


Figure 2B: HeLa human cervical epithelial carcinoma cells were untreated or treated with 0.5 mM PMA for 18 hours. 200 μ g of cell lysate was run on each array. Data shown are from a 5 minute exposure to X-ray film.

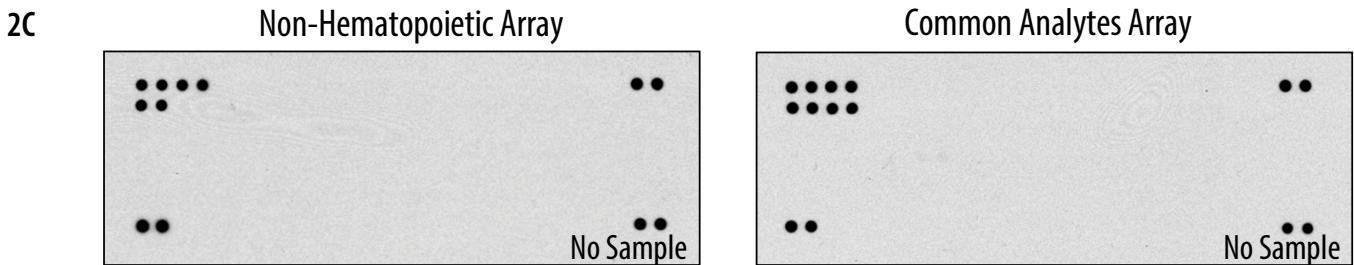


Figure 2C: Arrays incubated with buffer only (no sample) are shown. Images are from a 5 minute exposure to X-ray film.

PROFILING SOLUBLE RECEPTORS IN SERUM

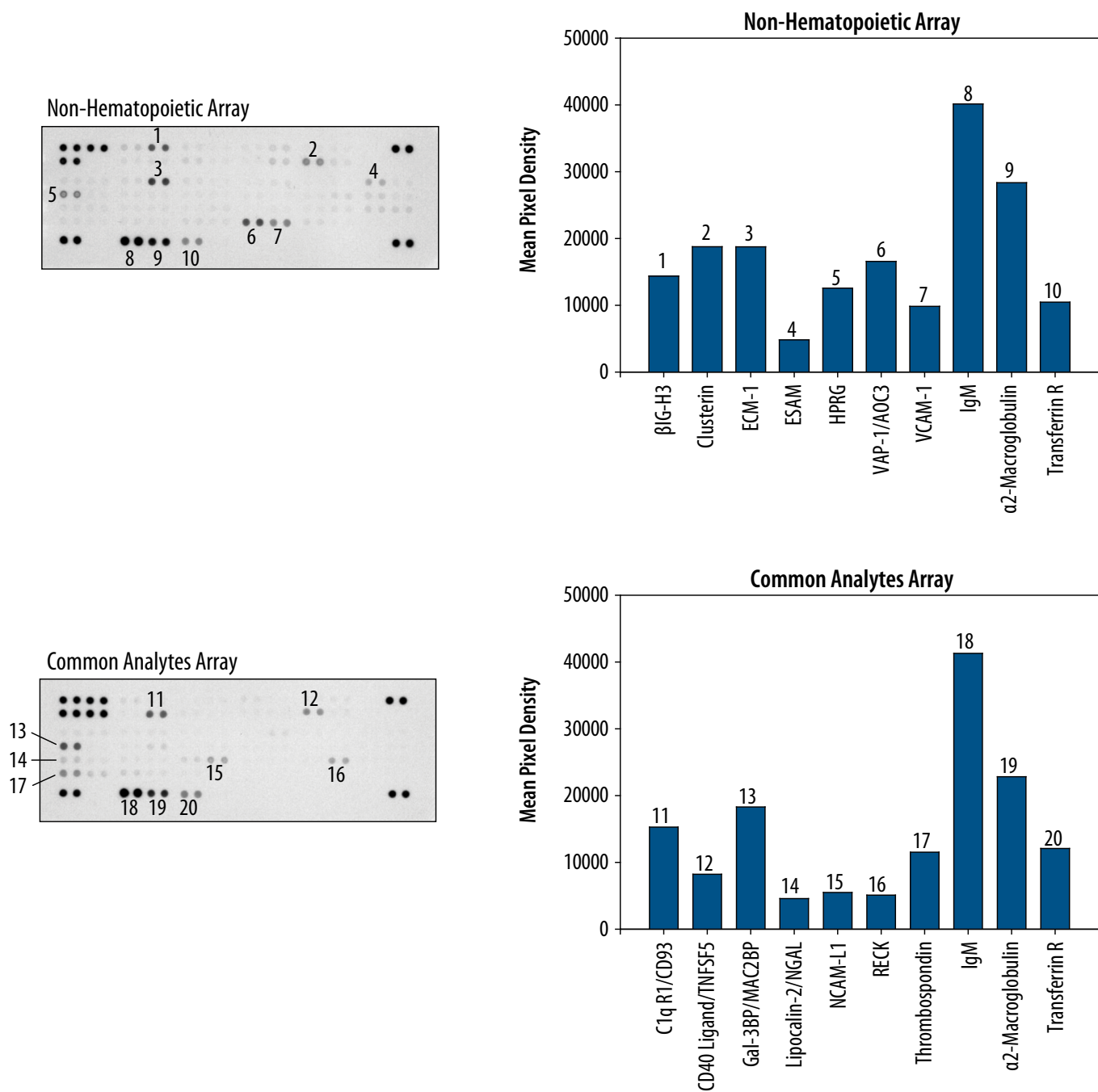


Figure 3: The Human Soluble Receptor Array detects multiple analytes in serum. 25 μ L of serum from a normal donor was run on each array. Data shown are from a 1 minute exposure to X-ray film.

APPENDIX

Refer to the table below for the Human Non-Hematopoietic Array (Part N) coordinates.

Coordinate	Analyte/Control	Alternate Nomenclature	Entrez Gene ID
A1, A2, A3, A4	Reference Spots	—	—
A5, A6	ADAM15	—	8751
A7, A8	βIG-H3	—	7045
A9, A10	BMPR-IB/ALK-6	CDw293	658
A11, A12	Cadherin-4/R-Cadherin	CAD4	1002
A13, A14	Cadherin-11	CAD11	1009
A15, A16	Cadherin-13	CDHH	1012
A17, A18	E-Cadherin	CD324, ECAD	999
A19, A20	N-Cadherin	CD325, NCAD	1000
A23, A24	Reference Spots	—	—
B1, B2	Reference Spots	—	—
B5, B6	P-Cadherin	PCAD	1001
B7, B8	VE-Cadherin	CD144, CDH5	1003
B9, B10	Cathepsin D	—	1509
B11, B12	CD40/TNFRSF5	—	958
B13, B14	CEACAM-5/CD66e	CEA	1048
B15, B16	CHL-1/L1CAM-2	CALL	10752
B17, B18	Clusterin	—	1191
B19, B20	Coagulation Factor II/Thrombin	—	2147
C1, C2	COMP/Thrombospondin-5	—	1311
C3, C4	CRELD2	—	79174
C5, C6	Desmoglein 2	—	1829
C7, C8	ECM-1	—	1893
C9, C10	EGF R/ErbB1	HER1	1956
C11, C12	Endoglycan	PODXL2	50512
C13, C14	EpCAM/TROP-1	—	4072
C15, C16	ErbB2/HER2	—	2064
C17, C18	ErbB3/HER3	—	2065
C19, C20	ErbB4/HER4	—	2066
C21, C22	ESAM	—	90952
C23, C24	Galectin-2	—	3957
D1, D2	HPRG	—	3273
D3, D4	Integrin α3/CD49c	ITGA3	3675
D5, D6	Integrin α5/CD49e	ITGA5	3678
D7, D8	Integrin α6/CD49f	ITGA6	3655
D9, D10	Integrin α9	ITGA9	3680
D11, D12	Integrin αV/CD51	ITGAV	3685

continued on next page....

APPENDIX CONTINUED

Coordinate	Analyte/Control	Alternate Nomenclature	Entrez Gene ID
D13, D14	Jagged 1	JAG1, CD339	182
D15, D16	JAM-B/VE-JAM	CD322, JAM2	58494
D17, D18	JAM-C	JAM3	83700
D19, D20	LRP-6	—	4040
D21, D22	MCAM/CD146	MUC18	4162
D23, D24	MEPE	—	56955
E1, E2	MUCDHL	—	53841
E3, E4	Nectin-2/CD112	PVRR2, PVRL2	5819
E5, E6	Nectin-4	PVRL4	81607
E7, E8	Neurotrimin	IGLON2	50863
E9, E10	Notch-1	—	4851
E11, E12	NrCAM	—	4897
E13, E14	Periostin/OSF-2	—	10631
E15, E16	Podocalyxin	PODXL	5420
E17, E18	E-Selectin/CD62e	CD62E, ELAM	6401
E19, E20	Semaphorin 3A	SEMA3A	10371
E21, E22	SREC-I/SR-F1	SCARF1	8578
E23, E24	SREC-II	SCARF2	91179
F1, F2	Stanniocalcin 1	STC1	6781
F3, F4	Syndecan-1/CD138	SDC1	6382
F5, F6	Syndecan-4	SDC4	6385
F7, F8	Thrombospondin-2	TSP2, THBS2	7058
F9, F10	TIMP-4	—	7079
F11, F12	TROP-2	—	4070
F13, F14	VAP-1/AOC3	SSAO	8639
F15, F16	VCAM-1	CD106	7412
F17, F18	VEGF R1/Flt-1	—	2321
F19, F20	VEGF R2/KDR/Flk-1	CD309	3791
G1, G2	Reference Spots	—	—
G5, G6	IgM (Sample Control)*	—	3507
G7, G8	α 2-Macroglobulin (Sample Control)*	A2M	2
G9, G10	Transferrin R (Sample Control)*	CD71	7037
G11, G12	Vimentin (Sample Control)*	VIM	7431
G13, G14	PBS (Negative Control)	—	—
G23, G24	Reference Spots	—	—

*Sample controls are included to allow for the detection of proteins commonly present in cell culture supernates, cell lysates, and serum. If these endogenous proteins are present in a particular sample, positive signals indicate that the sample has been incubated with the array and the assay procedure has been performed correctly.

APPENDIX CONTINUED

Refer to the table below for the Human Common Analytes Array (Part C) coordinates.

Coordinate	Analyte/Control	Alternate Nomenclature	Entrez Gene ID
A1, A2, A3, A4	Reference Spots	—	—
A5, A6	ACE	CD143	1636
A7, A8	ADAM8	CD156	101
A9, A10	ADAM9	—	8754
A11, A12	ADAM10	CD156c	102
A13, A14	ALCAM/CD166	—	214
A15, A16	Amphiregulin	AR	374
A17, A18	APP (pan)	—	351
A19, A20	BACE-1	—	23621
A23, A24	Reference Spots	—	—
B1, B2, B3, B4	Reference Spots	—	—
B5, B6	BCAM	CD239	4059
B7, B8	C1q R1/CD93	—	22918
B9, B10	CD9	—	928
B11, B12	CD23/Fc ϵ RII	—	2208
B13, B14	CD31/PECAM-1	—	5175
B15, B16	CD36/SR-B3	FAT	948
B17, B18	CD40 Ligand/TNFSF5	CD154	959
B19, B20	CD44H	—	960
C1, C2	CD58/LFA-3	—	965
C3, C4	CD90/Thy1	—	7070
C5, C6	CD99	—	4267
C7, C8	CD155/PVR	—	5817
C9, C10	CEACAM-1/CD66a	—	634
C11, C12	CX3CL1/Fractalkine	Neurotactin	6376
C13, C14	CXCL8/IL-8	NAP-1	3576
C15, C16	EMMPRIN/CD147	BSG	682
C17, C18	Endoglin/CD105	—	2022
C19, C20	Epiregulin	—	2069
C21, C22	Galectin-1	GAL1	3956
C23, C24	Galectin-3	GAL3	3958
D1, D2	Galectin-3BP/MAC-2BP	—	3959
D3, D4	HB-EGF	—	1839
D5, D6	ICAM-2/CD102	—	3384
D7, D8	IL-1 RII	CD121b	7850
D9, D10	IL-15 Ra	—	3601
D11, D12	Integrin β 1/CD29	ITGB1	3688

continued on next page....

APPENDIX CONTINUED

Coordinate	Analyte/Control	Alternate Nomenclature	Entrez Gene ID
D13, D14	Integrin β 2/CD18	ITGB2	3689
D15, D16	Integrin β 3/CD61	ITGB3	3690
D17, D18	Integrin β 4/CD104	ITGB4	3691
D19, D20	Integrin β 5	ITGB5	3693
D21, D22	Integrin β 6	ITGB6	3694
D23, D24	JAM-A	CD321	50848
E1, E2	Lipocalin-2/NGAL	—	3934
E3, E4	LOX-1/SR-E1	CLEC8A	4973
E5, E6	MD-1	LY86	9450
E7, E8	MMP-2 (total)	—	4313
E9, E10	NCAM-1/CD56	—	4684
E11, E12	NCAM-L1	L1CAM, CD171	3897
E13, E14	Osteopontin	OPN	6696
E15, E16	PAR1	—	2149
E17, E18	Pref-1/DLK-1/FA1	—	8878
E19, E20	RECK	—	8434
E21, E22	Stabilin-1	CLEVER-1, FEEL-1	23166
E23, E24	TACE/ADAM17	CD156b	6868
F1, F2	Thrombospondin	THBS, TSP	7057
F3, F4	TIMP-1	—	7076
F5, F6	TIMP-2	—	7077
F7, F8	TIMP-3	—	7078
F9, F10	TNF RII/TNFRSF1B	CD120b	7133
G1, G2	Reference Spots	—	—
G5, G6	IgM (Sample Control)	—	3507
G7, G8	α 2-Macroglobulin (Sample Control)*	A2M	2
G9, G10	Transferrin R (Sample Control)*	CD71	7037
G11, G12	Vimentin (Sample Control)*	VIM	7431
G13, G14	PBS (Negative Control)	—	—
G23, G24	Reference Spots	—	—

*Sample controls are included to allow for the detection of proteins commonly present in cell culture supernates, cell lysates, and serum. If these endogenous proteins are present in a particular sample, positive signals indicate that the sample has been incubated with the array and the assay procedure has been performed correctly.

All trademarks and registered trademarks are the property of their respective owners.