

biotechne[®]

R&D SYSTEMS

Proteome Profiler™ Array

Rat Cytokine Array Panel A

Catalog Number ARY008

For the parallel determination of the relative levels of selected rat cytokines and chemokines.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

TABLE OF CONTENTS

SECTION	PAGE
INTRODUCTION	1
PRINCIPLE OF THE ASSAY.....	1
TECHNICAL HINTS.....	2
PRECAUTIONS.....	2
MATERIALS PROVIDED & STORAGE CONDITIONS	3
OTHER SUPPLIES REQUIRED	4
SAMPLE COLLECTION & STORAGE.....	5
REAGENT PREPARATION.....	6
ARRAY PROCEDURE	7
DATA ANALYSIS	9
PROFILING PROTEINS IN CELL CULTURE SUPERNATES.....	10
PROFILING PROTEINS IN CELL LYSATES	11
PROFILING PROTEINS IN TISSUE LYSATES AND SERUM	12
APPENDIX.....	14

Manufactured and Distributed by:

USA R&D Systems, Inc.

614 McKinley Place NE, Minneapolis, MN 55413

TEL: 800 343 7475 612 379 2956

FAX: 612 656 4400

E-MAIL: info@bio-techne.com

Distributed by:

Europe | Middle East | Africa Bio-Techne Ltd.

19 Barton Lane, Abingdon Science Park

Abingdon OX14 3NB, UK

TEL: +44 (0)1235 529449

FAX: +44 (0)1235 533420

E-MAIL: info.emea@bio-techne.com

China Bio-Techne China Co., Ltd.

Unit 1901, Tower 3, Raffles City Changning Office,

1193 Changning Road, Shanghai PRC 200051

TEL: +86 (21) 52380373 (400) 821-3475

FAX: +86 (21) 52371001

E-MAIL: info.cn@bio-techne.com

INTRODUCTION

Cytokines and chemokines are extracellular signaling molecules that mediate cell-cell communication. They are released from cells and have critical roles in many biological processes such as cellular growth, differentiation, gene expression, migration, immunity and inflammation. In most biological processes, multiple cytokines operate in a large network, where the action of one cytokine is regulated by the presence or absence of other cytokines. Measuring cytokines present in biological samples one at a time is tedious and requires large sample volumes. The Proteome Profiler™ Rat Cytokine Array Panel A Kit is a rapid, sensitive, and economical tool for simultaneously profiling the relative levels of multiple cytokines between samples.

PRINCIPLE OF THE ASSAY

Capture and control antibodies have been spotted in duplicate on nitrocellulose membranes. Cell culture supernates, cell lysates, serum, plasma, or tissue lysates are diluted and mixed with a cocktail of biotinylated detection antibodies. The sample/antibody mixture is then incubated with the Rat Cytokine Array Panel A membrane. Any cytokine/detection antibody complex present is bound by its cognate immobilized capture antibody on the membrane. Following a wash to remove unbound material, Streptavidin-HRP and chemiluminescent detection reagents are applied and a signal is produced at each spot corresponding to the amount of cytokine bound. Refer to the Appendix for a list and coordinates of analytes and controls.

TECHNICAL HINTS

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- This kit should not be used beyond the expiration date on the kit label.
- **Do not mix or substitute reagents with those from other lots or sources. Substitution of some high intensity chemiluminescent reagents for Chemi Reagents 1 and 2 may cause either increased background or diminished signal depending on the reagent.**
- If using a LI-COR, additional reagents and protocol modifications are required. Refer to <https://www.rndsystems.com/resources/technical/use-proteome-profiler-arrays-li-cor-detection> for more details.
- Any variation in sample handling, buffers, operator, pipetting technique, washing technique, and incubation time or temperature can alter the performance of the kit.
- The Rat Cytokine Array membranes are validated for single use only.
- Always use gloved hands and flat-tipped tweezers to handle the membranes.
- Pick up the membranes from the edge on the side with the identification number avoiding the area with the printed antibodies.
- A thorough and consistent wash technique is essential for proper assay performance. Individual arrays should be washed in separate containers to minimize background. Wash Buffer should be removed completely from the membrane before proceeding to the next step.
- Do not allow the membrane to dry out. This will cause high background.
- Avoid microbial contamination of reagents and buffers.
- Soluble receptors and other proteins present in biological samples do not necessarily interfere with the measurement of cytokines in samples. Until these proteins have been tested with the Rat Cytokine Array Panel A, the possibility of interference cannot be excluded.
- For a procedure demonstration video, visit: www.RnDSystems.com/ProteomeProfilerVideo.

PRECAUTIONS

Chemi Reagents 1 and 2 contain Boric Acid which is suspected of damaging fertility or the unborn child.

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Refer to the SDS on our website prior to use.

MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

PART	PART #	DESCRIPTION	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Rat Cytokine Array Panel A	893587	4 nitrocellulose membranes each containing 29 different capture antibodies printed in duplicate.	Return unused membranes to the foil pouch containing the desiccant pack. Reseal along entire edge of the zip-seal. May be stored for up to 3 months at 2-8 °C.*
Array Buffer 4	895022	21 mL of a buffered protein base with preservatives. <i>May contain a precipitate. Mix well before and during use.</i>	May be stored for up to 3 months at 2-8 °C.*
Array Buffer 6	893573	2 vials (21 mL/vial) of a buffered protein base with preservatives.	May be stored for up to 3 months at 2-8 °C.*
Wash Buffer Concentrate	895003	2 vials (21 mL/vial) of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>	
Detection Antibody Cocktail, Rat Cytokine Array Panel A	893586	1 vial of a biotinylated antibody cocktail; lyophilized.	
Streptavidin-HRP	893019	200 µL of streptavidin conjugated to horseradish-peroxidase.	
Chemi Reagent 1	894287	1 vial (2.5 mL/vial)	
Chemi Reagent 2	894288	1 vial (2.5 mL/vial)	Store at room temperature.
4-Well Rectangular Multi-dish	607544	Clear 4-well rectangular multi-dish.	
Transparency Overlay Template	607590	1 transparency overlay template for coordinate reference.	

* Provided this is within the expiration date of the kit.

Note: Additional wash buffer is available for purchase ([R&D Systems®](#), # WA126).

OTHER SUPPLIES REQUIRED

- Aprotinin ([Tocris™, # 4139](#))
- Leupeptin hemisulfate ([Tocris, # 1167](#))
- Pepstatin A ([Tocris, # 1190](#))
- Igepal® CA-630 ([Sigma™, # I3021](#))
- Pipettes and pipette tips
- Gloves
- Phosphate-Buffered Saline (PBS) ([R&D Systems®, # RB01](#))
- Deionized or distilled water
- Rocking platform shaker
- Microcentrifuge
- A plastic container with the capacity to hold 50 mL (for washing the arrays)
- Plastic transparent sheet protector (trimmed to 10 cm x 12 cm and open on three sides)
- Plastic wrap
- Paper towels
- Absorbent lab wipes
- Autoradiography cassette
- Film developer
- X-ray film
- Flat-tipped tweezers
- Flatbed scanner with transparency adapter capable of transmission mode
- Computer capable of running image analysis software and Microsoft® Excel

If using cell lysate samples, the following buffers are also required:

- Phosphate-Buffered Saline (PBS) ([R&D Systems®, # RB01](#))
- Lysis Buffer (1% Igepal CA-630, 20 mM Tris-HCl (pH 8.0), 137 mM NaCl, 10% Glycerol, 2 mM EDTA, 10 µg/mL Aprotinin, 10 µg/mL Leupeptin hemisulfate, and 10 µg/mL Pepstatin A)

If using tissue lysate samples, the following are also required:

- PBS with protease inhibitors (10 µg/mL Aprotinin, 10 µg/mL Leupeptin hemisulfate, and 10 µg/mL Pepstatin A)
- Triton™ X-100 ([Sigma, # T9284](#))

SAMPLE COLLECTION & STORAGE

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Since the Rat Cytokine Array detects relative expression levels of individual analytes, it is important to include appropriate control samples.

Note: Sample amount may be empirically adjusted to attain optimal sensitivity with minimal background. Suggested starting ranges are: 200-700 μ L for cell culture supernates, 100-400 μ g for cell and tissue lysates, and 50-200 μ L for serum and plasma samples.

Cell Culture Supernates - Remove particulates by centrifugation. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Cell Lysates - Rinse cells with PBS, making sure to remove any remaining PBS before adding lysis buffer. Solubilize cells at 1×10^7 cells/mL in lysis buffer. Pipette up and down to resuspend and rock the lysates gently at 2-8 °C for 30 minutes. Microcentrifuge at 14,000 x g for 5 minutes, and transfer the supernate into a clean test tube. Quantitation of sample protein concentration using a total protein assay is recommended. Assay immediately or aliquot and store at ≤ -70 °C. Avoid repeated freeze-thaw cycles. Thawed lysates should be kept on ice prior to use.

Serum - Allow blood samples to clot for 2 hours at room temperature before centrifuging for 20 minutes at approximately 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 20 minutes at approximately 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Tissue Lysates - Excise tissue and homogenize in PBS with protease inhibitors. After homogenization, add Triton X-100 to a final concentration of 1%. Freeze samples at ≤ -70 °C, thaw, and centrifuge at 10,000 x g for 5 minutes to remove cellular debris. Quantitation of sample protein concentrations using a total protein assay is recommended. Assay immediately or aliquot and store samples at ≤ -70 °C. Avoid repeated freeze-thaw cycles. Thawed lysates should be kept on ice prior to use.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Rat Cytokine Array Panel A - Four nitrocellulose membranes each containing 29 different anti-cytokine antibodies printed in duplicate. **Handle membranes only with gloved hands and flat-tipped tweezers.**

Detection Antibody Cocktail - One vial of lyophilized biotinylated antibodies. Before use, reconstitute the Detection Antibody Cocktail with 100 μ L of deionized or distilled water.

1X Wash Buffer - If crystals have formed in the concentrate, warm the bottles to room temperature and mix gently until the crystals have completely dissolved. Add 40 mL of 25X Wash Buffer Concentrate to 960 mL of deionized or distilled water to prepare 1000 mL of 1X Wash Buffer.

Chemi Reagent Mix - Chemi Reagents 1 and 2 should be mixed in equal volumes within 15 minutes of use. **Protect from light. 1 mL of the resultant mixture is required per membrane.** Discard any remaining after use.

ARRAY PROCEDURE

Bring all reagents to room temperature before use. Keep samples on ice. To avoid contamination, wear gloves while performing the procedures.

1. Prepare all reagents and samples as directed in the previous sections.
2. Pipette 2 mL of Array Buffer 6 into each well of the 4-Well Multi-dish to be used. Array Buffer 6 serves as a block buffer.
3. Using flat-tip tweezers, remove each membrane to be used from between the protective sheets and place in a well of the 4-Well Multi-dish. The number on the membrane should be facing upward.

Note: *Upon contact with Array Buffer 6, the blue dye from the spots will disappear, but the capture antibodies are retained in their specific locations.*

4. Incubate for one hour on a rocking platform shaker. Orient the tray so that each membrane rocks end to end in its well.
5. While the membranes are blocking, prepare samples by adding up to 1 mL of each sample to 0.5 mL of Array Buffer 4 in separate tubes. Adjust to a final volume of 1.5 mL with Array Buffer 6 as necessary.
6. Add 15 μ L of reconstituted Detection Antibody Cocktail to each prepared sample. Mix and incubate at room temperature for one hour.
7. Aspirate Array Buffer 6 from the wells of the 4-Well Multi-dish and add sample/antibody mixtures prepared in steps 5 and 6. Place the lid on the 4-Well Multi-dish.
8. Incubate overnight at 2-8 °C on a rocking platform shaker.

Note: *A shorter incubation time may be used if optimal sensitivity is not required.*

9. Carefully remove each membrane and place into individual plastic containers with 20 mL of 1X Wash Buffer. Rinse the 4-Well Multi-dish with deionized or distilled water and dry thoroughly.
10. Wash each membrane with 1X Wash Buffer for 10 minutes on a rocking platform shaker. Repeat two times for a total of three washes.
11. Dilute the Streptavidin-HRP in Array Buffer 6 using the dilution factor on the vial label. Pipette 2 mL of diluted Streptavidin-HRP into each well of the 4-Well Multi-dish.
12. Carefully remove each membrane from its wash container. Allow excess buffer to drain from the membrane. Return the membrane to the 4-Well Multi-dish containing the diluted Streptavidin-HRP. Cover the wells with the lid.
13. Incubate for 30 minutes at room temperature on a rocking platform shaker.

ARRAY PROCEDURE *CONTINUED*

14. Wash each array as described in steps 9 and 10.

Note: *Complete the remaining steps without interruption.*

15. Carefully remove each membrane from its wash container. Allow excess Wash Buffer to drain from the membrane by blotting the lower edge onto paper towels. Place each membrane on the bottom sheet of the plastic sheet protector with the identification number facing up.

16. Pipette 1 mL of the prepared Chemi Reagent Mix evenly onto each membrane.

Note: *Using less than 1 mL of Chemi Reagent Mix per membrane may result in incomplete membrane coverage.*

17. Carefully cover with the top sheet of the plastic sheet protector. Gently smooth out any air bubbles and ensure Chemi Reagent Mix is spread evenly to all corners of each membrane. Incubate for 1 minute.

18. Position paper towels on top and sides of plastic sheet protector containing the membranes and carefully squeeze out excess Chemi Reagent Mix.

19. Remove the top plastic sheet protector and carefully lay an absorbent lab wipe on top of the membranes to blot off any remaining Chemi Reagent Mix.

20. Leaving the membranes on the bottom plastic sheet protector, cover the membranes with plastic wrap taking care to gently smooth out any air bubbles. Wrap the excess plastic wrap around the back of the sheet protector so that the membranes and sheet protector are completely wrapped.

21. Place the membranes with the identification numbers facing up in an autoradiography film cassette.

Note: *Use an autoradiography cassette that is not used with radioactive isotope detection.*

22. Expose membranes to X-ray film for 1-10 minutes. Multiple exposure times are recommended.

DATA ANALYSIS

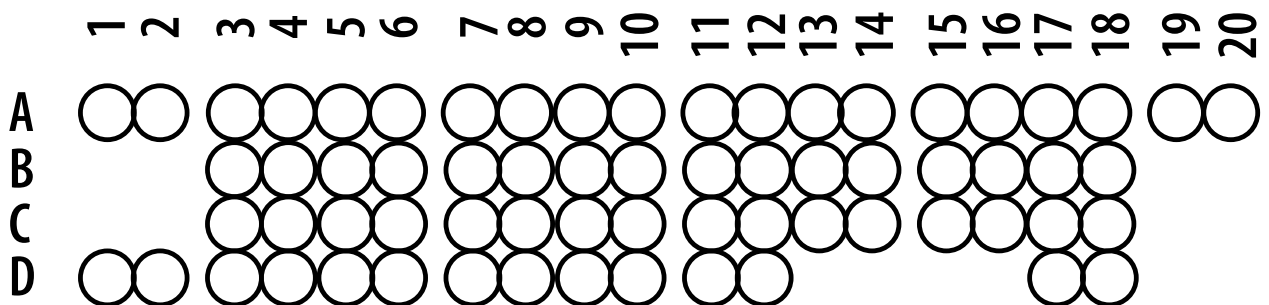
The positive signals seen on developed film can be quickly identified by placing the transparency overlay template on the array image and aligning it with the pairs of reference spots in three corners of each array. The stamped identification number on the array should be placed on the left hand side. The location of controls and cytokine capture antibodies is listed in the Appendix.

Note: Reference spots are included to align the transparency overlay template and to demonstrate that the array has been incubated with Streptavidin-HRP during the assay procedure.

Pixel densities on developed X-ray film can be collected and analyzed using a transmission-mode scanner and image analysis software.

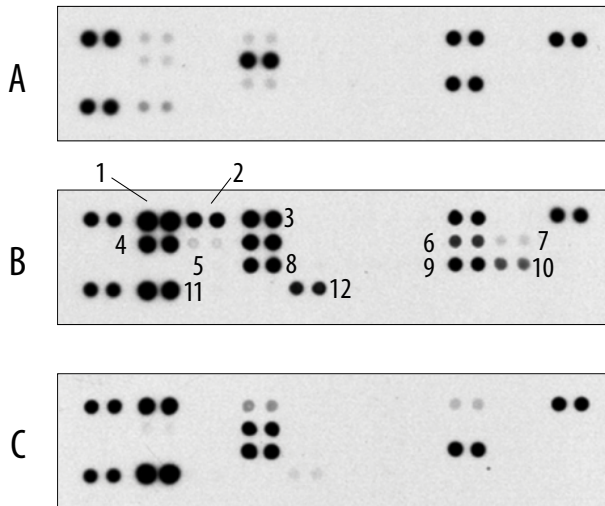
1. Create a template to analyze pixel density in each spot of the array.
2. Export signal values to a spreadsheet file for manipulation in a program such as Microsoft Excel.
3. Determine the average signal (pixel density) of the pair of duplicate spots representing each cytokine.
4. Subtract an averaged background signal from each spot. Use a signal from a clear area of the array or negative control spots as a background value.
5. Compare corresponding signals on different arrays to determine the relative change in cytokine levels between samples.

Rat Cytokine Array Panel A Coordinates



This image is not to scale; it is for coordinate reference only.
Please use the transparency overlay for analyte identification.

PROFILING PROTEINS IN CELL CULTURE SUPERNATES



		MEAN PIXEL DENSITY		
		A	B	C
1	CINC-1	10,132	53,770	50,001
2	CINC-2α/β	794	45,826	721
3	CINC-3	9175	51,769	18,158
4	IL-1α/IL-1F1	7883	50,730	4734
5	IL-1β/IL-1F2	694	6987	905
6	IL-6	535	28,269	820
7	IL-10	453	7630	915
8	IP-10	10,642	45,370	46,015
9	MIP-1α/CCL3	42,556	37,520	41,834
10	MIP-3α	731	26,063	1446
11	RANTES/CCL5	16,491	51,258	53,724
12	TNF-α	206	36,230	6739

NR8383 Cell Supernates

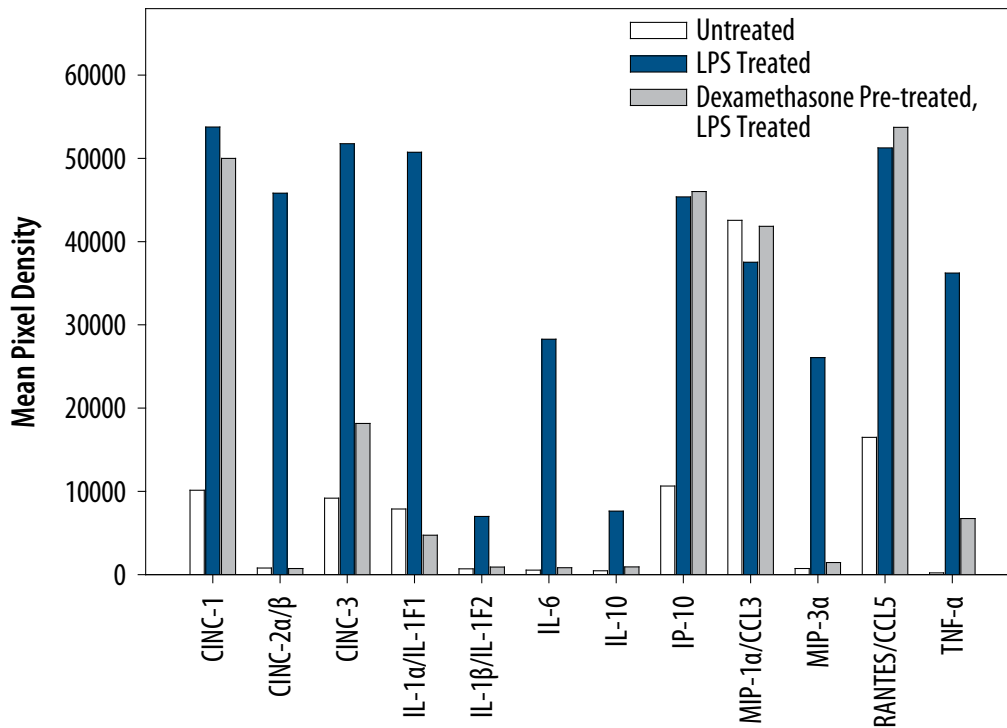


Figure 1: The Rat Cytokine Array detects multiple analytes in cell culture supernates.

NR8383 rat alveolar macrophage cells were either untreated or treated as below. Data shown are from a two minute exposure to X-ray film.

A. Untreated

B. Treated with 100 ng/mL LPS for 24 hours.

C. Pre-treated with 100 nM Dexamethasone for 72 hours followed by treatment with 100 ng/mL LPS for 24 hours.

PROFILING PROTEINS IN CELL LYSATES

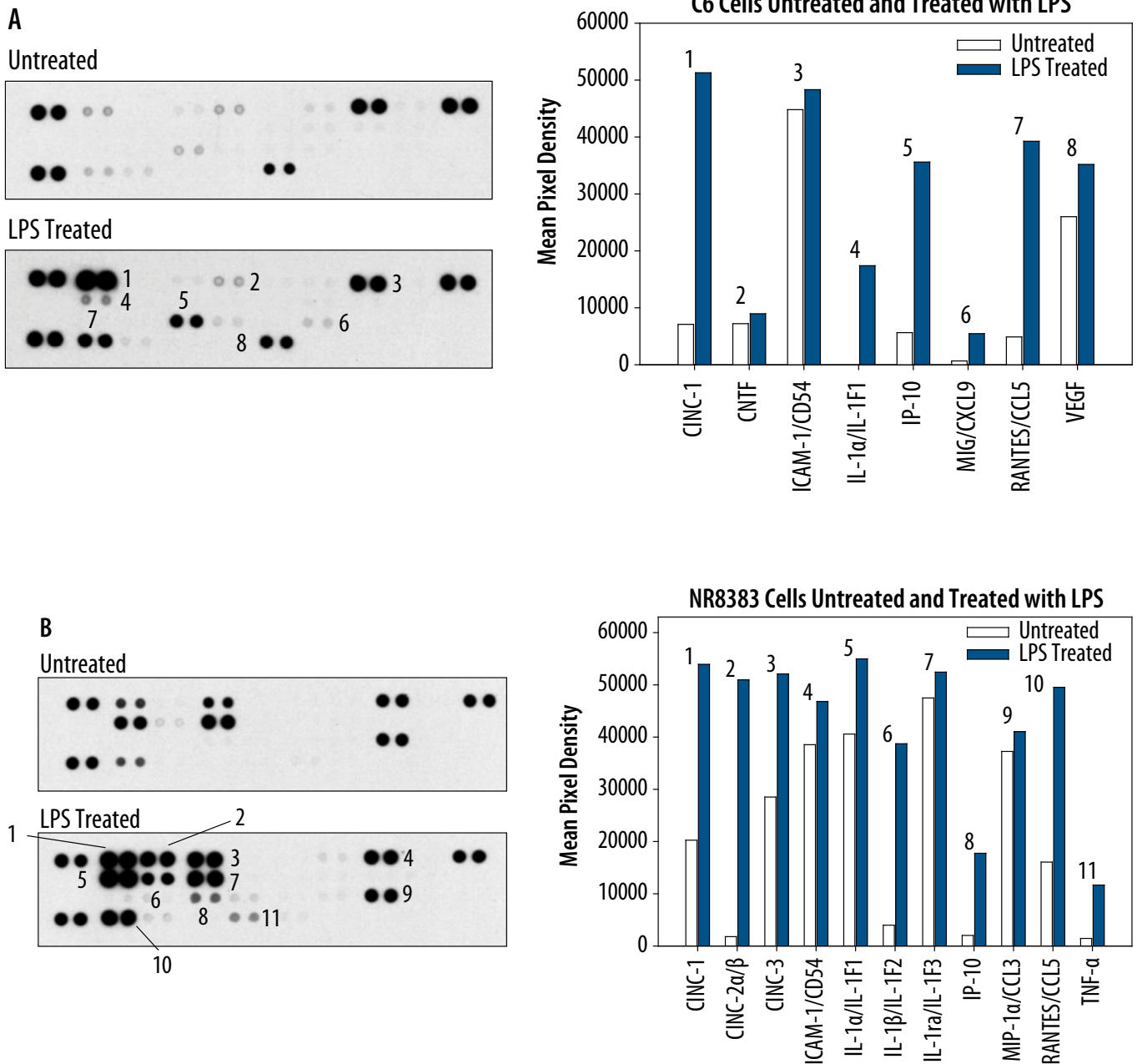


Figure 2: The Rat Cytokine Array detects multiple analytes in untreated and treated cell lysates. Data shown are from a 5 minute exposure to X-ray film.

A. C6 rat glioma cells were either untreated or treated with 100 ng/mL LPS for 24 hours. 200 μ g of lysate was run on each array.

B. NR8383 rat alveolar macrophage cells were either untreated or treated with 50 ng/mL LPS for 24 hours. 200 μ g of lysate was run on each array.

PROFILING PROTEINS IN TISSUE LYSATES AND SERUM

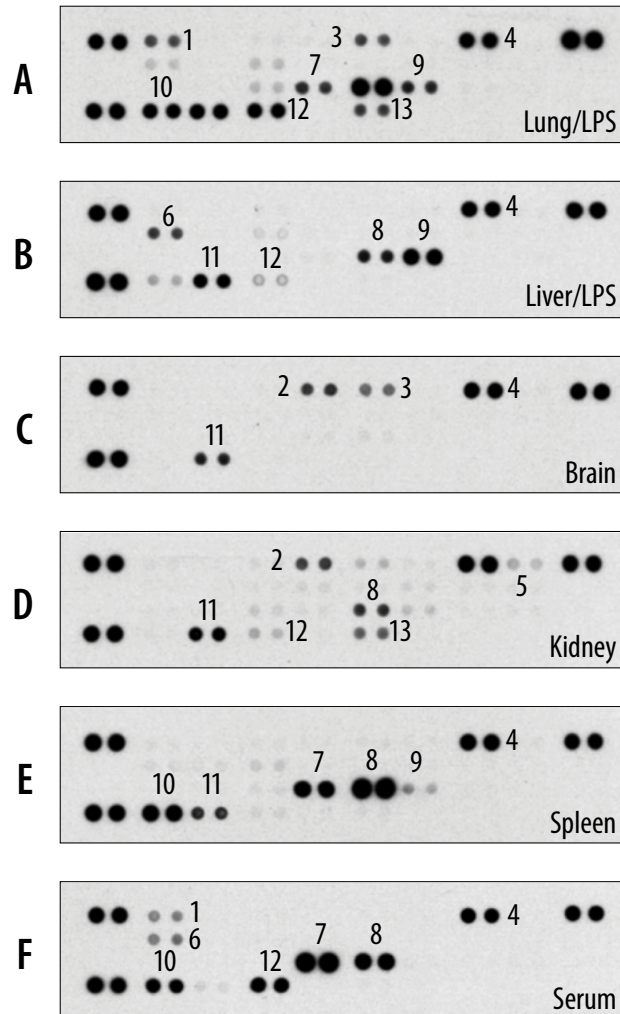


Figure 3: The Rat Cytokine Array detects multiple analytes in tissue lysates and serum.

A-B. A rat was injected with 0.1 mg/kg LPS for 24 hours. Tissues were excised and prepared as described in the Sample Collection and Storage section. 400 µg of lysate was run on each array. Data shown are from a 7 minute exposure to X-ray film.

C-E. Tissues were excised from untreated rats and prepared as described in the Sample Collection and Storage section. 400 µg of lysate was run on each array. Data shown are from a 7 minute exposure to X-ray film.

F. Serum samples from 15 week old male rats were prepared as described in the Sample Collection and Storage section. 100 µL of serum was run on the array. Data shown are from a 5 minute exposure to X-ray film.

PROFILING PROTEINS IN TISSUE LYSATES AND SERUM *CONTINUED*

		MEAN PIXEL DENSITY					
		A	B	C	D	E	F
		Lung/LPS	Liver/LPS	Brain	Kidney	Spleen	Serum
1	CINC-1	16,812	1096	406	2284	844	10,792
2	CNTF	399	287	19,226	19,226	1183	1226
3	Fractalkine	15,631	450	13,245	4998	1733	1544
4	ICAM-1/CD54	38,203	38,243	38,416	42,109	42,834	36,505
5	IFN- γ	590	1049	1277	5614	1936	1190
6	IL-1 α /IL-1F1	3695	15,835	768	1343	1624	12,050
7	LIX	19,682	984	1118	2172	43,605	51,272
8	L-Selectin	46,309	22,458	1568	19,403	52,406	44,565
9	MIG/CXCL9	20,979	41,455	723	4224	6818	1966
10	RANTES/CCL5	34,020	6041	635	1873	41,514	33,135
11	Thymus Chemokine	36,064	30,758	20,324	30,315	21,319	2709
12	TIMP-1	33,931	5988	661	6003	2562	37,417
13	VEGF	17,487	1012	801	14,013	2739	1892

APPENDIX

Refer to the table below for the Rat Cytokine Array coordinates.

Coordinate	Target/Control	Alternate Nomenclature
A1, A2	Reference Spots	—
A3, A4	CINC-1	—
A5, A6	CINC-2 α / β	—
A7, A8	CINC-3	—
A9, A10	CNTF	—
A11, A12	Fractalkine	CX3CL1
A13, A14	GM-CSF	—
A15, A16	ICAM-1	CD54
A17, A18	IFN- γ	—
A19, A20	Reference Spots	—
B3, B4	IL-1 α	IL-1F1
B5, B6	IL-1 β	IL-1F2
B7, B8	IL-1ra	IL-1F3
B9, B10	IL-2	—
B11, B12	IL-3	—
B13, B14	IL-4	—
B15, B16	IL-6	—
B17, B18	IL-10	—
C3, C4	IL-13	—
C5, C6	IL-17	—
C7, C8	IP-10	CXCL10
C9, C10	LIX	—
C11, C12	L-Selectin	CD62L/LECAM-1
C13, C14	MIG	CXCL9
C15, C16	MIP-1 α	CCL3
C17, C18	MIP-3 α	CCL20
D1, D2	Reference Spots	—
D3, D4	RANTES	CCL5
D5, D6	Thymus Chemokine	CXCL7
D7, D8	TIMP-1	—
D9, D10	TNF- α	TNFSF1A
D11, D12	VEGF	VEGF-A/Vasculotropin
D17, D18	Negative Control	Control (-)

All trademarks and registered trademarks are the property of their respective owners.

©2024 R&D Systems®, Inc.