

## DESCRIPTION

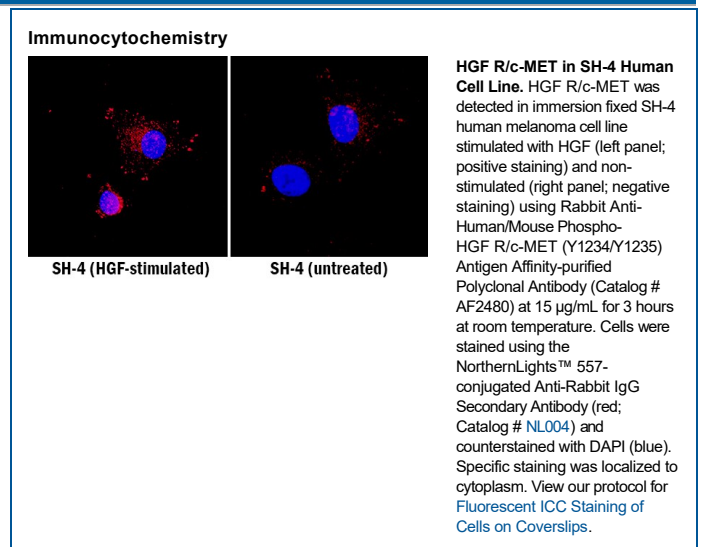
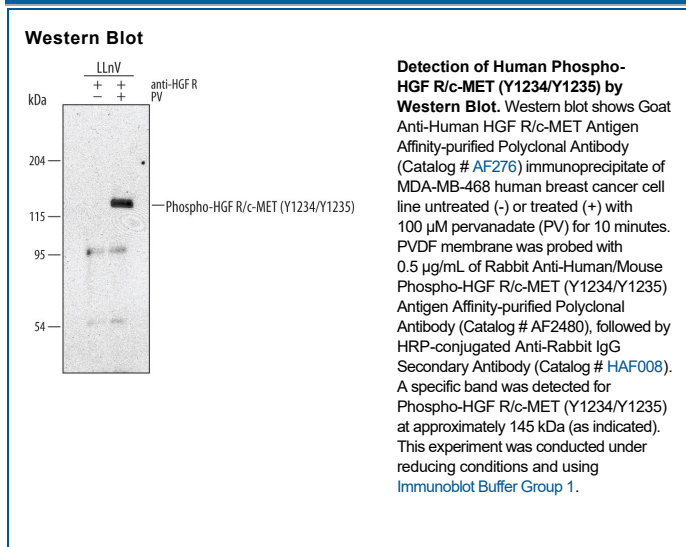
|                           |   |
|---------------------------|---|
| <b>Species Reactivity</b> | Human/Mouse   |
| <b>Specificity</b>        | Detects human and mouse HGF R/c-MET when phosphorylated at Y1234/Y1235 in Western blots.  |
| <b>Source</b>             | Polyclonal Rabbit IgG   |
| <b>Purification</b>       | Antigen Affinity-purified   |
| <b>Immunogen</b>          | Phosphopeptide containing human HGF R/c-MET Y1234/1235 sites  |
| <b>Formulation</b>        | Lyophilized from a 0.2 µm filtered solution in PBS with Trehalose. See Certificate of Analysis for details.<br>*Small pack size (-SP) is supplied either lyophilized or as a 0.2 µm filtered solution in PBS. |

## APPLICATIONS

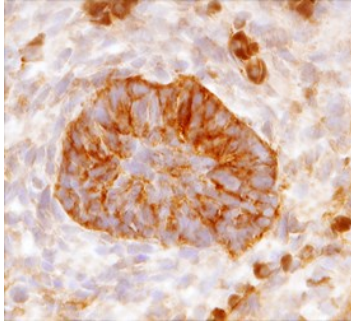
**Please Note:** Optimal dilutions should be determined by each laboratory for each application. *General Protocols* are available in the *Technical Information* section on our website.

|   | Recommended Concentration   | Sample    |
|---|---|-----------|
| <b>Western Blot</b>                             | 0.5 µg/mL   | See Below |
| <b>Immunocytochemistry</b>                      | 5-15 µg/mL  | See Below |
| <b>Immunohistochemistry</b>                     | 5-15 µg/mL  | See Below |
| <b>Intracellular Staining by Flow Cytometry</b> | 2.5 µg/10 <sup>6</sup> cells  | See Below |
| <b>Simple Western</b>                           | 5 µg/mL   | See Below |
| <b>CyTOF-reported</b>                           | Brodie, T.M. <i>et al.</i> (2018) <i>Cytometry Part A</i> . <b>93</b> : 406. Ready to be labeled using established conjugation methods. No BSA or other carrier proteins that could interfere with conjugation. |           |

## DATA

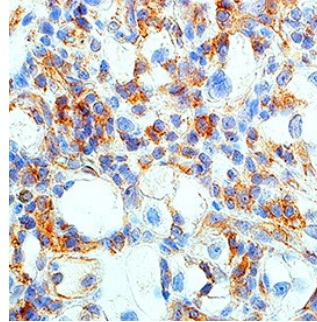


## Immunohistochemistry



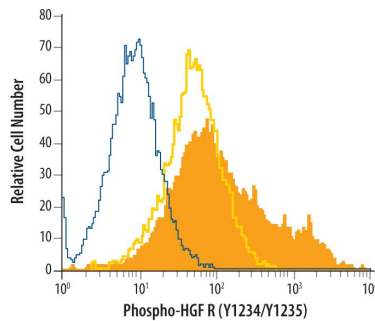
**HGF R/c-MET in Mouse Embryo.** HGF R/c-MET was detected in immersion fixed frozen sections of mouse embryo (13 d.p.c.) using Rabbit Anti-Human/Mouse Phospho-HGF R/c-MET (Y1234/Y1235) Antigen Affinity-purified Polyclonal Antibody (Catalog # AF2480) at 15 µg/mL overnight at 4 °C. Tissue was stained using the Anti-Rabbit HRP-DAB Cell & Tissue Staining Kit (brown; Catalog # CTS005) and counterstained with hematoxylin (blue). View our protocol for [Chromogenic IHC Staining of Frozen Tissue Sections](#).

## Immunohistochemistry



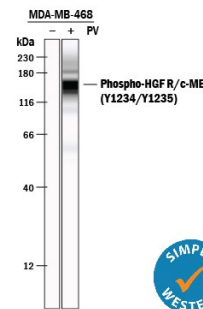
**HGF R/c-MET in Human Renal Cell Carcinoma Tissue.** HGF R/c-MET was detected in immersion fixed paraffin-embedded sections of human renal cell carcinoma tissue using Rabbit Anti-Human/Mouse Phospho-HGF R/c-MET (Y1234/Y1235) Antigen Affinity-purified Polyclonal Antibody (Catalog # AF2480) at 15 µg/mL for 1 hour at room temperature followed by incubation with the Anti-Rabbit IgG VisUCyte™ HRP Polymer Antibody (Catalog # VC003). Before incubation with the primary antibody, tissue was subjected to heat-induced epitope retrieval using Antigen Retrieval Reagent-Basic (Catalog # CTS013). Tissue was stained using DAB (brown) and counterstained with hematoxylin (blue). Specific staining was localized to cytoplasm. View our protocol for [IHC Staining with VisUCyte HRP Polymer Detection Reagents](#).

## Intracellular Staining by Flow Cytometry



**Detection of HGF R/c-MET in pervanadate-treated MCF-7 Human Cell Line by Flow Cytometry.** MCF-7 human breast cancer cell line was unstimulated (light orange open histogram) or treated with 100 µM pervanadate for 10 minutes (dark orange filled histogram), then stained with Rabbit Anti-Human/Mouse Phospho-HGF R/c-MET (Y1234/Y1235) Antigen Affinity-purified Polyclonal Antibody (Catalog # AF2480), or control antibody (Catalog # AB-105-C, blue open histogram), followed by Phycoerythrin-conjugated Anti-Rabbit IgG Secondary Antibody (Catalog # F0110). To facilitate intracellular staining, cells were fixed with paraformaldehyde and permeabilized with methanol.

## Simple Western



**Detection of Human Phospho-HGF R/c-MET (Y1234/Y1235) by Simple Western™.** Simple Western lane view shows lysates of MDA-MB-468 human breast cancer cell line untreated (-) or treated (+) with 100 µM Pervanadate (PV) for 10 minutes, loaded at 0.2 mg/mL. A specific band was detected for HGF R/c-MET at approximately 156 kDa (as indicated) using 5 µg/mL of Rabbit Anti-Human/Mouse Phospho-HGF R/c-MET (Y1234/Y1235) Antigen Affinity-purified Polyclonal Antibody (Catalog # AF2480). This experiment was conducted under reducing conditions and using the 12-230 kDa separation system.



## PREPARATION AND STORAGE

|                                |   |
|--------------------------------|---|
| <b>Reconstitution</b>          | Reconstitute at 0.2 mg/mL in sterile PBS.   |
| <b>Shipping</b>                | The product is shipped at ambient temperature. Upon receipt, store it immediately at the temperature recommended below.<br>*Small pack size (-SP) is shipped with polar packs. Upon receipt, store it immediately at -20 to -70 °C  |
| <b>Stability &amp; Storage</b> | Use a manual defrost freezer and avoid repeated freeze-thaw cycles. <ul style="list-style-type: none"> <li>• 12 months from date of receipt, -20 to -70 °C as supplied.</li> <li>• 1 month, 2 to 8 °C under sterile conditions after reconstitution.</li> <li>• 6 months, -20 to -70 °C under sterile conditions after reconstitution.</li> </ul> |

**BACKGROUND**

HGF R, also known as Met (from *N*-methyl-*N*-nitro-*N*-nitrosoguanidine induced), is a glycosylated receptor tyrosine kinase that plays a central role in epithelial morphogenesis and cancer development. HGF R is synthesized as a single chain precursor which undergoes cotranslational proteolytic cleavage. This generates a mature HGF R that is a disulfide-linked dimer composed of a 50 kDa extracellular  $\alpha$  chain and a 145 kDa transmembrane  $\beta$  chain (1, 2). The extracellular domain (ECD) contains a seven bladed  $\beta$ -propeller sema domain, a cysteine-rich PSI/MRS, and four Ig-like E-set domains, while the cytoplasmic region includes the tyrosine kinase domain (3, 4). Proteolysis and alternate splicing generate additional forms of human HGF R which either lack of the kinase domain, consist of secreted extracellular domains, or are deficient in proteolytic separation of the  $\alpha$  and  $\beta$  chains (5-7). The sema domain, which is formed by both the  $\alpha$  and  $\beta$  chains of HGF R, mediates both ligand binding and receptor dimerization (3, 8). Ligand-induced tyrosine phosphorylation in the cytoplasmic region activates the kinase domain and provides docking sites for multiple SH2-containing molecules (9, 10). HGF stimulation induces HGF R downregulation *via* internalization and proteasome-dependent degradation (11). In the absence of ligand, HGF R forms noncovalent complexes with a variety of membrane proteins including CD44v6, CD151, EGF R, Fas, Integrin  $\alpha 6/\beta 4$ , Plexins B1, 2, 3, and MSP R/Ron (12-19). Ligation of one complex component triggers activation of the other, followed by cooperative signaling effects (12-19). Formation of some of these heteromeric complexes is a requirement for epithelial cell morphogenesis and tumor cell invasion (12, 16, 17). Paracrine induction of epithelial cell scattering and branching tubulogenesis results from the stimulation of HGF R on undifferentiated epithelium by HGF released from neighboring mesenchymal cells (20). Genetic polymorphisms, chromosomal translocation, overexpression, and additional splicing and proteolytic cleavage of HGF R have been described in a wide range of cancers (1). Within the ECD, human HGF R shares 86%-88% aa sequence identity with canine, mouse, and rat HGF R.

**References:**

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