

## DESCRIPTION

<b>Species Reactivity</b>	Mouse
<b>Specificity</b>	Detects mouse Crossveinless-2/CV-2 in direct ELISAs and Western blots. In Western blots, approximately 50% cross-reactivity with recombinant human CV-2 is observed.
<b>Source</b>	Polyclonal Goat IgG
<b>Purification</b>	Antigen Affinity-purified
<b>Immunogen</b>	Mouse myeloma cell line NS0-derived recombinant mouse Crossveinless-2/CV-2 (R&D Systems, Catalog # 2299-CV) Ala39-Arg685 Accession # AAH66153
<b>Formulation</b>	Lyophilized from a 0.2 µm filtered solution in PBS with Trehalose.

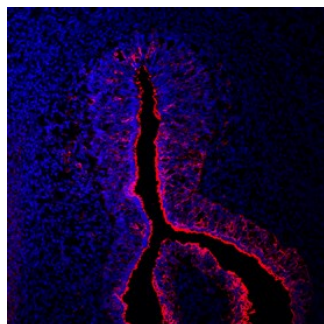
## APPLICATIONS

**Please Note:** Optimal dilutions should be determined by each laboratory for each application. [General Protocols](#) are available in the Technical Information section on our website.

	<b>Recommended Concentration</b>	<b>Sample</b>
<b>Western Blot</b>	0.1 µg/mL	Recombinant Mouse Crossveinless-2/CV-2 (Catalog # <a href="#">2299-CV</a> )
<b>Immunohistochemistry</b>	5-15 µg/mL	See Below

## DATA

### Immunohistochemistry



**Crossveinless-2/CV-2 in Mouse Embryo.** Crossveinless-2/CV-2 was detected in immersion fixed frozen sections of mouse embryo (E13.5) using Mouse Crossveinless-2/CV-2 Antigen Affinity-purified Polyclonal Antibody (Catalog # AF2299) at 10 µg/mL overnight at 4 °C. Tissue was stained using the NorthernLights™ 557-conjugated Anti-Goat IgG Secondary Antibody (red; Catalog # [NL001](#)) and counterstained with DAPI (blue). Specific staining was localized to the epithelium surrounding the nasal cavity. View our protocol for [Fluorescent IHC Staining of Frozen Tissue Sections](#).

## PREPARATION AND STORAGE

<b>Reconstitution</b>	Reconstitute at 0.2 mg/mL in sterile PBS.
<b>Shipping</b>	The product is shipped at ambient temperature. Upon receipt, store it immediately at the temperature recommended below.
<b>Stability &amp; Storage</b>	<p><b>Use a manual defrost freezer and avoid repeated freeze-thaw cycles.</b></p> <ul style="list-style-type: none"> <li>• 12 months from date of receipt, -20 to -70 °C as supplied.</li> <li>• 1 month, 2 to 8 °C under sterile conditions after reconstitution.</li> <li>• 6 months, -20 to -70 °C under sterile conditions after reconstitution.</li> </ul>

## BACKGROUND

Crossveinless-2 (CV-2), also known as bone morphogenetic protein-binding endothelial cell precursor-derived regulator (BMPER), is a secreted chordin-like protein that modulates the BMP signaling pathway (1-3). Mouse CV-2 is synthesized as a 685 amino acid (aa) residue precursor protein with a putative 39 aa signal peptide, five tandem chordin-like cysteine-rich (CR) domains, a partial von Willebrand factor type D domain (vWD), and a carboxyl trypsin inhibitor-like cysteine-rich domain (TIL) (1, 2, 4). Secreted CV-2 is reported to be proteolytically cleaved to generate two fragments that are disulfide-linked (1, 2). The GDPH sequence is conserved in CV-2 from other species. It is also found in multiple proteins that undergo a similar type of cleavage (5). Mouse CV-2 message is detected in many tissues, with the highest expression detected in the heart, lungs, and skin (2). It is also expressed in flk-1<sup>+</sup> endothelial cell precursors and in primary chondrocytes (2). During embryonic development, CV-2 is expressed in the dorsal midline, regions of the telencephalon, migrating cells of the branchial neural crest and endothelial cells in the yolk sac (2). Mouse CV-2 shares 92% and 34% aa sequence identity with the human and *Drosophila* homologs, respectively (1, 4). Results from biochemical experiments using recombinant CV-2 show that CV-2 directly interacts with BMP-2, -4, and -6 to antagonize BMP signaling, which can regulate a wide range of differentiation processes (1, 2). In contrast, genetic data from *Drosophila* suggest that CV-2 potentiates BMP-signaling (6). It is possible that like TSG, CV-2 can positively and negatively modulate BMP signal transduction depending on the cell context (7).

## References:

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2. Moser, M. *et al.* (2003) *Mol Cell Biol.* **23**:5664.
3. Garcia-Abreu, J. *et al.* (2002) *Gene* **287**:39.
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5. Lidell, M.E. *et al.* (2003) *J. Biol. Chem.* **278**:13944.
6. Conley, C.A. *et al.* (2000) *Development* **127**:3947.
7. Kamimura, M. *et al.* (2004) *Developmental Dynamics* **230**:434.