

**DESCRIPTION**

<b>Source</b>	Chinese Hamster Ovary cell line, CHO-derived human C1GalT1 protein Asp45-Pro363, with N-terminal 6-His tag (C1GalT1); Ile28-Asp318, with an N-terminal HA tag (C1GalT1C1) Accession # Q9NS00 (C1GalT1) & Q96EU7 (C1GalT1C1)
<b>N-terminal Sequence Analysis</b>	His (C1GalT1) & Tyr (C1GalT1C1)
<b>Predicted Molecular Mass</b>	38 kDa (C1GalT1) & 35 kDa (C1GalT1C1)

**SPECIFICATIONS**

<b>SDS-PAGE</b>	25-42 kDa, reducing conditions
<b>Activity</b>	Measured by its ability to transfer galactose from UDP-galactose to 4-nitrophenyl- $\alpha$ -D-galactosaminide. The specific activity is >2,750 pmol/min/ $\mu$ g, as measured under the described conditions.
<b>Endotoxin Level</b>	<1.0 EU per 1 $\mu$ g of the protein by the LAL method.
<b>Purity</b>	>90%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.
<b>Formulation</b>	Supplied as a 0.2 $\mu$ m filtered solution in Tris and NaCl. See Certificate of Analysis for details.

**Activity Assay Protocol**

<b>Materials</b>	<ul style="list-style-type: none"> <li>Glycosyltransferase Activity Kit (Catalog # EA001)</li> <li>Assay Buffer: 25 mM Tris, 5 mM CaCl<sub>2</sub>, 10 mM MnCl<sub>2</sub>, pH 7.5</li> <li>Recombinant Human C1GalT1 (rhC1GalT1) (Catalog # 8659-GT)</li> <li>Uridine 5'-diphosphogalactose (UDP-Gal) (Sigma, Catalog # U4500), 10 mM stock in deionized water</li> <li>4-Nitrophenyl N-acetyl-<math>\alpha</math>-D-galactosaminide (4-NP-GalNAc) (Sigma, Catalog # N4264), 15 mM stock in DMSO</li> <li>96-well Clear Plate (Catalog # DY990)</li> <li>Plate Reader (Model: SpectraMax Plus by Molecular Devices) or equivalent</li> </ul>
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<b>Assay</b>	<ol style="list-style-type: none"> <li>Dilute 1 mM Phosphate Standard provided by the Glycosyltransferase Kit by adding 40 <math>\mu</math>L of the 1 mM Phosphate Standard to 360 <math>\mu</math>L of Assay Buffer for a 100 <math>\mu</math>M stock. This is the first point of the standard curve.</li> <li>Complete the standard curve by performing six one-half serial dilutions of the 100 <math>\mu</math>M Phosphate stock using Assay Buffer. The standard curve has a range of 0.078 to 5 nmol per well.</li> <li>Prepare reaction mixture containing 1.2 mM UDP-Gal, 0.6 mM 4-NP-GalNAc, and 4 <math>\mu</math>g/mL Coupling Phosphatase 1 in Assay Buffer.</li> <li>Dilute rhC1GALT1 to 0.6 <math>\mu</math>g/mL in Assay Buffer.</li> <li>Load 50 <math>\mu</math>L of each dilution of the standard curve into a plate. Include a curve blank containing 50 <math>\mu</math>L of Assay Buffer.</li> <li>Load 25 <math>\mu</math>L of 0.6 <math>\mu</math>g/mL rhC1GALT1 into empty wells of the same plate as the curve. Include a Control containing 25 <math>\mu</math>L of Assay Buffer.</li> <li>Add 25 <math>\mu</math>L of the reaction mixture to all wells, excluding the standard curve.</li> <li>Seal plate and incubate at 37 °C for 20 minutes.</li> <li>Add 30 <math>\mu</math>L of the Malachite Green Reagent A to all wells. Mix briefly.</li> <li>Add 100 <math>\mu</math>L of deionized water to all wells. Mix briefly.</li> <li>Add 30 <math>\mu</math>L of the Malachite Green Reagent B to all wells. Mix and incubate sealed plate for 20 minutes at room temperature.</li> <li>Read plate at 620 nm (absorbance) in endpoint mode.</li> <li>Calculate specific activity:</li> </ol>
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$$\text{Specific Activity (pmol/min/}\mu\text{g)} = \frac{\text{Phosphate released* (nmol)} \times (1000 \text{ pmol/nmol})}{\text{Incubation time (min)} \times \text{amount of enzyme (}\mu\text{g)}}$$

\*Derived from the phosphate standard curve using linear or 4-parameter fitting and adjusted for Control.

<b>Final Assay Conditions</b>	<p>Per Well:</p> <ul style="list-style-type: none"> <li>rhC1GALT1: 0.015 <math>\mu</math>g</li> <li>Coupling Phosphatase 1: 0.1 <math>\mu</math>g</li> <li>UDP-Gal: 0.6 mM</li> <li>4-NP-GalNAc: 0.3 mM</li> </ul>
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**PREPARATION AND STORAGE**

<b>Shipping</b>	The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below.
<b>Stability &amp; Storage</b>	<p>Use a manual defrost freezer and avoid repeated freeze-thaw cycles.</p> <ul style="list-style-type: none"> <li>6 months from date of receipt, -20 to -70 °C as supplied.</li> <li>3 months, -20 to -70 °C under sterile conditions after opening.</li> </ul>

**BACKGROUND**

O-glycosylation is a ubiquitous post-translational modification of secreted and membrane bound proteins (1). The synthesis of mucin-type O-glycans is initiated by the addition of GalNAc to threonine or serine residues on proteins by polypeptide N-acetylgalactosaminyltransferases (GALNTs) (2). The GalNAc $\alpha$ 1-O-Ser/Thr structure is then extended by other glycosyltransferases to form eight types of core O-glycans (3). Core 1  $\beta$ -3-galactosyltransferase (C1GalT1), in particular, synthesizes Gal- $\beta$ 1-3GalNAc $\alpha$ 1-O-Ser/Thr, a precursor for all core 1 and core 2 based mucin-type O-glycans (4). These glycans play central roles in many processes, such as angiogenesis, thrombopoiesis, and kidney homeostasis (5). C1GalT1 forms a stable, non-covalent complex with Cosmc chaperone, C1GalT1C1, which is required for the full activity of C1GalT1 (6). Defective C1GalT1 causes a rare autoimmune disease called Tn syndrome (4) as well as susceptibility to IgA nephropathy (7). The recombinant C1GalT1 is co-purified with C1GalT1C1. The enzymatic activity of recombinant human C1GalT1 was determined using a phosphatase-coupled assay (8).

**References:**

1. Gerken, T.A. *et al.* (2011) *J. Biol. Chem.* **286**:14493.
2. Bergstrom, K.S.B. and Xia, L. (2013) *Glycobiology* **23**:1026.
3. Brockhausen, I. *et al.* (1999) *Biochim. Biophys. Acta.* **1473**:67.
4. Ju, T. and Cummings, R.D. (2005) *Nature* **437**:1252.
5. Fukuda, M. *et al.* (2002) *Biochim. Biophys. Acta.* **1573**:394.
6. Aryal, R.P. *et al.* (2012) *J. Biol. Chem.* **287**:15317.
7. Pirulli, D. *et al.* (2009) *J. Nephrol.* **22**:152.
8. Wu, Z.L. *et al.* (2011) *Glycobiology* **21**:727.