

DESCRIPTION

# Recombinant *F. meningosepticum*PNGase F

Catalog Number: 7695-GH

Source	E. coli-derived Ala41-Asn354 with N-terminal Met and 6-His tag Accession # AAA85323
N-terminal Sequence Analysis	Met
Predicted Molecular Mass	36 kDa
SPECIFICATIONS	
SDS-PAGE	33-35 kDa, reducing conditions
Activity	Measured by its ability to deglycosylate ribonuclease B under denatured conditions.  >50% ribonuclease B (10 μg) is deglycosylated by 10 ng of rfmPNGase F within 30 minutes, as measured under the described conditions. See Activity Assay Protocol on www.RnDSystems.com
Endotoxin Level	<1.0 EU per 1 µg of the protein by the LAL method.
Purity	>90%, by SDS-PAGE under reducing conditions and visualized by Colloidal Coomassie® Blue stain at 5 μg per lane.
Formulation	Supplied as a 0.2 µm filtered solution in Tris and NaCl. See Certificate of Analysis for details.
Activity Assay Protoco	
Materials	<ul> <li>Assay Buffer: 0.1 M Tris, pH 7.5</li> <li>Denaturing Buffer (10X): 5% SDS, 0.8 M β-Mercaptoethanol</li> <li>Recombinant <i>F. meningosepticum</i> PNGase F (<i>rFm</i>PNGaseF) (Catalog # 7695-GH)</li> <li>Ribonuclease B, from bovine pancreas (RNase B) (Sigma, Catalog # R7884), 2.5 mg/mL stock in 25 mM Tris, pH 7.5</li> <li>10% (v/v) Triton® X-100 in deionized water</li> <li>Reducing SDS-PAGE Sample Buffer</li> <li>SDS-PAGE or Western Blot</li> </ul>
Assay	<ol> <li>Dilute Denaturing Buffer to 5X in deionized water.</li> <li>Create a Substrate Mixture containing 0.8 mg/mL RNAse B and 1X denaturing buffer in deionized water.</li> <li>Heat Substrate Mixture at 100 °C for 10 minutes. Cool to room temperature and microcentrifuge briefly.</li> <li>Add Triton X-100 to a final concentration of 1.67%.</li> <li>Dilute rFmPNGaseF to 0.67 ng/μL in Assay buffer.</li> <li>Combine 15 μL of Substrate Mixture and 15 μL of 0.67 ng/μL rFmPNGaseF. Include a control containing 15 μL of Substrate Mixture and 15 μL of Assay buffer.</li> <li>Incubate at 37 °C for 30 minutes.</li> <li>Combine equal volumes of incubated reaction mixture and reducing SDS-PAGE sample buffer and boil samples at 100 °C for 3-5 minutes.</li> <li>Load 15 μL (2.5 μg RNase B) per lane on a 4-20% SDS-PAGE gel.</li> <li>Stain gel and analyze for percent deglycosylation using densitometry.</li> </ol>
Final Assay Conditions	Per Reaction:  • rfmPNGaseF: 10 ng  • RNase B: 10 µg
PREPARATION AND ST	FORAGE

### PREPARATION AND STORAGE

Shipping The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below

## Stability & Storage

Use a manual defrost freezer and avoid repeated freeze-thaw cycles.

- 6 months from date of receipt, -20 to -70 °C as supplied
- 3 months, -20 to -70 °C under sterile conditions after opening.

## BACKGROUND

PNGase F, peptide N-glycosidase F from Flavobacterium meningosepticum, catalyzes the hydrolysis of asparagine-linked high mannose, as well as hybrid and complex oligosaccharides from glycoproteins (1). Unlike glycosidases that hydrolyze glycosidic bonds, PNGase F is an amidase that cleaves the β-aspartylglucosamine bond between the innermost GlcNAc of N-glycans and asparagine residues of glycoproteins (2). The enzyme is highly active on various N-glycans except those with the innermost GlcNAc modified with α1-3-linked core fucose, which is commonly found on plant glycoproteins (3). Cleavage with PNGase F will convert the asparagine residue to an aspartic residue, allowing identification of the glycosylation site by mass spectrometry (4). This purified enzyme is compatible with glycan analysis using mass spectrometry.

## References:

- Elder, J.H. and Alexander, S. (1982) Proc. Natl. Acad. Sci. USA 79:4540.
- 2. Maley, F. et al. (1989) Anal. Biochem. 180:195.
- 3. Tarentino, A.L. and Plummer, T.H. (1994) Methods Enzymol 230:44.
- 4. Zhang, H. et al. (2003) Nat. Biotechnol. 21:660.

### PRODUCT SPECIFIC NOTICES

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