

# **Recombinant Human** Adenosine Deaminase 2/CECR1

Catalog Number: 7518-AD

DESCRIPTION	RIPTION	
Source	Chinese Hamster Ovary cell line, CHO-derived human Adenosine Deaminase 2/CECR1 protein lle30-Lys511, with a C-terminal 6-His tag Accession # AAF65941	
N-terminal Sequence Analysis	lle30	
Predicted Molecular Mass	57 kDa	

SPECIFICATIONS	
SDS-PAGE	58-66 kDa, reducing conditions
Activity	Measured by the ability to catalyze the hydrolytic deamination of adenosine to inosine. The specific activity is >14,000 pmol/min/µg, as measured under the described conditions.
Endotoxin Level	<1.0 EU per 1 µg of the protein by the LAL method.
Purity	>95%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.
Formulation	Supplied as a 0.2 µm filtered solution in HEPES, NaCl, Glycerol. See Certificate of Analysis for details.

#### **Activity Assay Protocol** Materials Assay Buffer: 25 mM Sodium phosphate, 1 M NaCl, pH 6.0 Recombinant Human Adenosine Deaminase 2/CECR1 (rhCECR1) (Catalog # 7518-AD) Substrate: Adenosine (Sigma, Catalog # A9251), 10 mM stock in deionized water (incubate 10 minutes at 37 °C to fully solubilize) Stop/detection reagent: 0.2 M Sodium Hydroxide, 15 mM ortho-phthaldehyde (Sigma, Catalog # P0657), 0.1% β-mercaptoethanol F16 Black Maxisorp Plate (Nunc, Catalog # 475515) Fluorescent Plate Reader (Model: SpectraMax Gemini EM by Molecular Devices) or equivalent Assay 1. Dilute rhCECR1 to 0.5 μg/mL in Assay Buffer. Dilute Substrate to 2 mM in Assay Buffer

۷.	Dilute Substrate to 2 mily in Assay buller.
3.	In plate, combine 50 µL dilute rhCECR1 with 50 µL dilute substrate. Include a substrate control containing 50 µL dilute enzyme only.
4.	Incubate reactions at room temperature for 10 minutes.

- 5. Add 100 μL stop/detection reagent to all wells used. Add 50 μL substrate to substrate control wells.
- 6. Incubate for 30 minutes at room temperature in the dark to fully develop.
- 7. Read with excitation and emission wavelengths of 330 nm and 450 nm (top read), respectively, in endpoint mode.
- 8. Specific activity may be determined using the following equation:

Specific Activity (pmol/min/ $\mu$ g) =  $\frac{\text{Adjusted Fluorescence* (RFU) x Conversion Factor*** (pmol/RFU)}}{\text{Adjusted Fluorescence*}}$ Incubation time (min) x amount of enzyme (µg) \*Adjusted for substrate control

\*\*Derived using calibration standard ammonium sulfate (Amresco, Catalog # 0191)

Final Assay	Per Reaction:
Conditions	<ul> <li>rhCECR1: 0.025 μg</li> </ul>
	Adenosine: 1 mM

## PREPARATION AND STORAGE

The product is shipped with dry ice or equivalent. Upon receipt, store it immediately at the temperature recommended below. Shipping

# Stability & Storage

Use a manual defrost freezer and avoid repeated freeze-thaw cycles.

- 6 months from date of receipt, -70 °C as supplied
- 3 months, -70 °C under sterile conditions after opening.

## BACKGROUND

Adenosine deaminase is one of the key enzymes of purine nucleotide metabolism. It catalyses the conversion of adenosine and deoxy-adenosine to inosine and deoxy-inosine, respectively (1, 2). Adenosine Deaminase 2 (ADA2) is also known as CECR1 because it is a candidate gene for cat eye syndrome, a developmental disorder (3), ADA is a secreted protein that is expressed in many tissues, with the highest expression in lymphoblasts, heart, lung, and placenta (4), ADA2 is a member of a family of adenosine deaminase-related growth factors (ADGFs), proteins that are involved in tissue development (4). ADA2 induces the differentiation of monocytes into macrophages and stimulates the proliferation of T helper cells and macrophages by a mechanism independent of its catalytic activity (5). It has been suggested that ADA2 could be a therapeutic target for the control of immune responses in inflammation and cancer (5).

### References:

- 1. Wolfenden, R.V. et al. (1969) Biochemistry 6:2412.
- 2. Lowenstein, J.M. (1972) Physiol. Rev. 52:382.
- 3. Riazi, M.A. et al. (2000) Genomics 64:277.
- Zavialov, A.V. and A. Engstrom (2005) Biochem. J. 391:51.
- 5. Zavialov, A.V. et al. (2010) J. Leucoc. Biol. 88:279.

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