

Recombinant Human β-1,4-Glucuronyltransferase 1/B4GAT1 His-tag

Catalog Number: 6664-GT

DESCRIPTION	
Source	Chinese Hamster Ovary cell line, CHO-derived human beta-1,4-Glucuronyltransferase 1/B4GAT1 protein His37-Cys415, with N-terminal 6-His tag Accession # O43505
Predicted Molecular Mass	43.8 kDa
SPECIFICATIONS	
SDS-PAGE	42-53 kDa under reducing conditions
Activity	Measured by its ability to transfer GIcA from UDP-GIcA to Xylose. The specific actiivity is >55 pmol/min/µg, as measured under the described conditions.
Endotoxin Level	<1.0 EU per 1 µg of the protein by the LAL method.
Purity	>95%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.
Formulation	Supplied as a 0.2 µm filtered solution in Tris and NaCl. See Certificate of Analysis for details.
Activity Assay Protoc	ol ,
Materials	 Glycosyltransferase Activity Kit (Catalog # EA001) Assay Buffer: 25 mM Tris, 10 mM CaCl₂, 10 mM MnCl₂ (supplied in kit), pH 7.5 Recombinant Human β-1,4-Glucuronyltransferase 1/B4GAT1 His-tag (rhB4GAT1) (Catalog # 6664-GT) Donor Substrate: UDP-GlcA (Sigma, Catalog # U5625), 10 mM stock in deionized water Acceptor Substrate: Xylose (V-lab, Catalog # BX53), 100 mM stock in deionized water 96-well Clear Plate (Catalog # DY990) Plate Reader (Model: SpectraMax Plus by Molecular Devices) or equivalent
Assay	 Dilute 1 mM Phosphate Standard provided by the Glycosyltransferase Kit by adding 40 μL of the 1 mM Phosphate Standard to 360 μL of Assay Buffer for a 100 μM stock. This is the first point of the standard curve. Prepare standard curve by performing six one-half serial dilutions of the 100 μM Phosphate stock in Assay Buffer. The standard curve has a range of 0.078 to 5 nmol per well. Prepare reaction mixture containing 0.8 mM UDP-GlcA, 40 mM Xylose, and 4 μg/mL Coupling Phosphatase I in Assay Buffer. Dilute rhB4GAT1 to 20 μg/mL in Assay Buffer. Load 50 μL of each dilution of the standard curve into a plate. Include a curve blank containing 50 μL of Assay Buffer. Load 25 μL of 20 μg/mL rhB4GAT1 into the plate. Include a Control containing 25 μL of Assay Buffer. Add 25 μL of reaction mixture to all wells, excluding the standard curve and curve blank. Seal plate and incubate at 37 °C for 20 minutes. Add 30 μL of the Malachite Green Reagent A to all wells. Mix briefly. Add 30 μL of the Malachite Green Reagent B to all wells. Mix and incubate for 20 minutes at room temperature. Read plate at 620 nm (absorbance) in endpoint mode. Calculate specific activity: Specific Activity (pmol/min/μg) = Phosphate released* (nmol) x (1000 pmol/nmol)
	Incubation time (min) x amount of enzyme (µg) *Derived from the phosphate standard curve using linear or 4-parameter fitting and adjusted for Control.
Final Assay Conditions	Per Reaction: • rhB4GAT1: 0.5 μg

PREPARATION AND STORAGE	
Shipping	The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below.
Stability & Storage	Use a manual defrost freezer and avoid repeated freeze-thaw cycles.
	 6 months from date of receipt, -20 to -70 °C as supplied.
	 3 months, -20 to -70 °C under sterile conditions after opening.

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• Coupling Phosphatase I: 0.1 μg

Xylose: 20 mMUDP-GlcA: 0.4 mM



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BACKGROUND

B4GAT1 was previously described in the literature as B3GNT1 (1). It is now characterized as a β 1,4 glucuronyltransferase that is responsible for the synthesis of a glucuronyl- β 1,4-xylosyl disaccharide found on α -dystroglycan (α - DG), a peripheral membrane protein that binds to several extracellular matrix components (2). Proper glycosylation of α -DG is critical to maintain structural integrity and force transmission between the cytoskeleton and the extracellular matrix for efficient signal transduction. Mutation of B4GAT1 will lead to failure of proper glycosylation of α -DG and loss of receptor binding of α -DG, therefore causes congenital muscular dystrophies (CMDs) (3). The enzymatic activity of recombinant human B4GAT1 was determined using a phosphatase-coupled assay (4) using xylose as acceptor substrate.

References:

- 1. Sasaki, K. et al. (1997) PNAS 94:14294.
- 2. Willer, T. et al. (2014) eLife; 3: e03941.
- 3. Barresi, R. and Campbell, K.P. (2006) J. Cell Sci. 119:199.
- 4. Wu, Z.L. et al. (2011) Glycobiology 21:727.

