

DESCRIPTION

Source	Chinese Hamster Ovary cell line, CHO-derived human Integrin alpha V beta 8 protein			
	Human Integrin α V (Phe31-Val992) Accession # NP_002201.1	His-Pro	GGGSGGGG	Acidic Tail
	Human Integrin β 8 (Glu43-Arg684) Accession # P26012.1	His-Pro	GGGSGGGG	Basic Tail
	N-terminus		C-terminus	
N-terminal Sequence Analysis	Phe31 (α V subunit) & Glu43 (β 8 subunit)			
Predicted Molecular Mass	110.5 kDa (α V subunit), 75.3 kDa (β 8 subunit)			

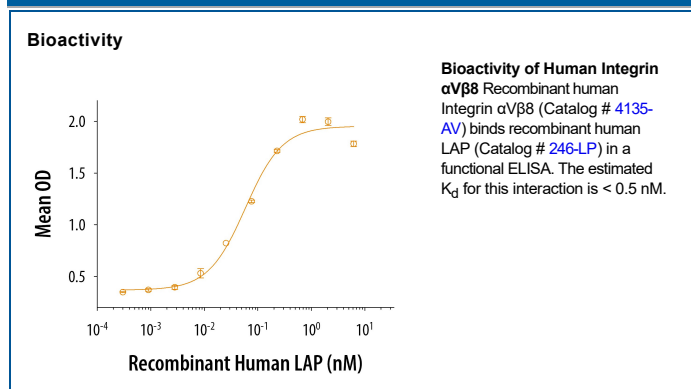
SPECIFICATIONS

SDS-PAGE	130-155 kDa and 85-100 kDa, reducing conditions
Activity	Measured by its binding ability in a functional ELISA. Immobilized Recombinant Human Integrin α V β 8 at 2 μ g/mL can bind recombinant human LAP with an apparent K_d <0.5 nM.
Endotoxin Level	<0.10 EU per 1 μ g of the protein by the LAL method.
Purity	>90%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.
Formulation	Lyophilized from a 0.2 μ m filtered solution in Tris, NaCl and MgCl ₂ . See Certificate of Analysis for details.

PREPARATION AND STORAGE

Reconstitution	Reconstitute at 100 μ g/mL in sterile PBS.
Shipping	The product is shipped at ambient temperature. Upon receipt, store it immediately at the temperature recommended below.
Stability & Storage	<p>Use a manual defrost freezer and avoid repeated freeze-thaw cycles.</p> <ul style="list-style-type: none"> 12 months from date of receipt, -20 to -70 °C as supplied. 2 weeks, 2 to 8 °C under sterile conditions after reconstitution. 3 months, -20 to -70 °C under sterile conditions after reconstitution.

DATA



BACKGROUND

Integrin α V β 8 is one of five α V integrins and the only known β 8 integrin (1-3). The non-covalent heterodimer of 170 kDa α V and ~90 kDa β 8 integrin type I transmembrane glycoprotein subunits is expressed in yolk sac, placenta, brain perivascular astrocytes, Schwann cells, renal glomerular mesangial cells and pulmonary epithelial cells (3-7). Unlike other α V integrins, α V β 8 does not appear to assume different activation states, and the cytoplasmic tail does not connect to the cytoskeleton (3, 8). It binds ligands containing an RGD motif, including vitronectin, fibrin and the latency associated peptide (LAP) of the latent TGF- β complex (7-12). High affinity binding of α V β 8 to LAP allows proteolytic cleavage by MT1-MMP, which releases active TGF- β . This mechanism differs from that of α V β 6, the other α V integrin which can activate TGF- β from latency through non-proteolytic mechanisms (13). Downstream effects of TGF- β activation include control of cell growth and associated vascularization (10-13). Deletion of either α V or β 8 reveals that α V β 8 is required for vascular morphogenesis in the embryonic brain and yolk sac (4, 14, 15). The 962 aa human α V extracellular domain (ECD) shares 92-95% aa sequence identity with mouse, rat and cow α V, while the 642 aa human β 8 ECD shares 92%, 92%, 89%, 87% and 87% aa identity with cow, dog, rabbit, mouse and rat β 8, respectively. The β 8 ECD of β 8 shows low (~35%) aa identity with other integrin β subunits, and the cytoplasmic tail is unlike any other integrin. The α V ECD contains an N-terminal β -propeller structure, followed by domains termed thigh, calf-1 and calf-2 (1). The β 8 ECD contains a vWFA domain, which interacts with the α V β -propeller to form a binding domain. Each subunit has a transmembrane sequence and a short cytoplasmic tail.

References:

1. Hynes, R. O. (2002) *Cell* **110**:673.
2. Suzuki, S. *et al.* (1987) *J. Biol. Chem.* **262**:14080.
3. Moyle, M. *et al.* (1991) *J. Biol. Chem.* **266**:19650.
4. Zhu, J. *et al.* (2002) *Development* **129**:2891.
5. Cambier, S. *et al.* (2000) *Cancer Res.* **60**:7084.
6. Lakhe-Reddy, S. *et al.* (2006) *J. Biol. Chem.* **281**:19688.
7. Chernousov, M. A. and D. J. Carey (2003) *Exp. Cell Res.* **291**:514.
8. Nishimura, S. L. *et al.* (1994) *J. Biol. Chem.* **269**:28708.
9. Milner, R. *et al.* (1999) *J. Cell Sci.* **112**:4271.
10. Cambier, S. *et al.* (2005) *Am. J. Pathol.* **166**:1883.
11. Fjellbirkeland, L. *et al.* (2003) *Am. J. Pathol.* **163**:533.
12. Araya, J. *et al.* (2006) *Am. J. Pathol.* **169**:405.
13. Mu, D. *et al.* (2002) *J. Cell Biol.* **157**:493.
14. Proctor, J. M. *et al.* (2005) *J. Neurosci.* **25**:9940.
15. McCarty, J. H. *et al.* (2005) *Development* **132**:165.