

PRODUCT DESCRIPTION

Type I Collagen is the major structural component of extracellular matrices found in connective tissue and internal organs, but is most prevalent in the dermis, tendons, and bone. It is a 300 kDa molecule composed of two $\alpha_1(I)$ chains and one $\alpha_2(I)$ chain that spontaneously forms a triple helix scaffold at a neutral pH and 37 °C. This phenomenon can be exploited to promote cell attachment, growth, differentiation, migration, and tissue morphogenesis during development.

3-D culture is an innovative approach to modeling the morphological effects of early oncogenesis on three-dimensional microenvironments. When healthy, differentiating cells exhibit a structured, polarized morphology that is critical for cellular formation and function. During carcinoma development, cell cycle controls associated with cellular development, proliferation and death are lost, and as a result, these structures are disrupted. In effect, the morphology of these structures can be used as a measure to study factors in early carcinoma development. The 3-D Culture Matrix Rat Collagen was designed to be used as a gel on which to grow cells or a media additive alone or in concert with other basement membrane components to study cellular growth and differentiation in three dimensions *in vitro*.

INTENDED USE

Cultrex 3-D Culture Matrix Rat Collagen I may be used as a hydrogel or thin coating for tissue culture vessels. It may also be used to supplement customized coatings, hydrogels or medium formulations for cell culture. To provide the most standardized Collagen I for use in 3-D cultures, a special process is employed to provide material at a standard concentration of approximately 5 mg/mL. This material is then incorporated in a 3-D culture to validate efficacy.

PRODUCT SPECIFICATIONS

Concentration	5 mg/mL
Source	Rat tail tendons
Storage Buffer	20 mM Acetic Acid
Stability	Product is stable for a minimum of 3 months if stored at 2-8 °C. See lot specific Certificate of Analysis for expiration date.
Storage	Store at 2-8 °C. Do Not Freeze.

PRECAUTION

When handling bio-hazardous materials such as human cells, safe laboratory procedures should be followed and protective clothing should be worn.

LIMITATIONS

- FOR LABORATORY RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The safety and efficacy of this product in diagnostic or other clinical uses has not been established.
- Results may vary due to variations among tissue/cells derived from different donors or sources.

MATERIAL QUALIFICATIONS

Sterility Testing:

- No bacterial or fungal growth detected after a 14 day culture.
- No mycoplasma contamination detected by PCR.
- Endotoxin concentrations < 20 EU/mL by LAL assay.

Gelling Assay:

- Cultrex 3-D Culture Matrix Rat Collagen I forms a firm gel at neutral pH and 37 °C when diluted to 0.4 mg/mL.

Functional Assays:

- Cell Attachment: Tested for the ability to promote cell attachment and spreading of HT-1080 human fibrosarcoma cells.
- 3-D Culture: Cultrex 3-D Culture Matrix Rat Collagen I promotes attachment and growth of murine endothelial SVEC4-10 cells.

GELLING PROCEDURE

To prevent contamination maintain aseptic techniques in a laminar flow biological hood throughout this procedure. Working with solutions that are pre-chilled at 2-8 °C, and keeping solutions on ice extends the time that Cultrex 3-D Culture Matrix Rat Collagen I will remain in solution after neutralization.

Note: *The recommended working concentration for Cultrex 3-D Culture Matrix Rat Collagen I 1 mg/mL.*

Gelling Procedure using 1X PBS and 1 N NaOH for Neutralization.

1. Place the following on ice:
 - a. Cultrex 3-D Culture Matrix Rat Collagen I (5 mg/mL)
 - b. Sterile 10X Phosphate Buffered Saline (PBS)
 - c. Sterile distilled water
 - d. Sterile 1N NaOH (fresh)
2. Determine the concentration and final volume of Collagen needed for experimentation.
 - a. Volume of Collagen needed =
$$\frac{(\text{Final concentration of Collagen}) \times (\text{Total volume})}{\text{Initial concentration of Collagen}}$$
 - b. Volume of 10X PBS needed =
$$\frac{\text{Total Volume}}{10}$$
 - c. Volume of 1N NaOH needed = [(Volume of Collagen I, step a) x 0.023 mL]
 - d. Volume of distilled water needed = [Total Volume - (sum of volumes from steps a+b+c)]
3. In a sterile tube mix the 10X PBS, and distilled water, and 1N NaOH.
4. Add the Cultrex 3-D Culture Matrix Rat Collagen I to the tube and pipette up and down to mix. **Do not vortex.**
5. Place the diluted Cultrex 3-D Culture Matrix Rat Collagen I into the desired plates or dishes. This solution is stable for up to 1 hour on ice.

Note: *Plates may be centrifuged 300 x g for 10 minutes at 2-8 °C to prevent bubbles from forming in the gel.*
6. Incubate the plate at 37 °C for 1 hour to promote gel formation.

GELLING PROCEDURE *Continued*

Gelling Procedure using 7.5% (w/v) Sodium Bicarbonate for Neutralization.

- Place the following on ice:
 - Cultrex 3-D Culture Matrix Rat Collagen I (5 mg/mL)
 - Sterile 10X PBS
 - Sterile distilled water
 - Sterile 7.5% Sodium Bicarbonate
- Determine the concentration and final volume of Cultrex 3-D Culture Matrix Rat Collagen I (5 mg/mL) needed for experimentation.
 - Volume of Collagen needed =
$$\frac{(\text{Final concentration of Collagen}) \times (\text{Total volume})}{\text{Initial concentration of Collagen I}}$$
 - Volume of 10X PBS needed =
$$\frac{\text{Total Volume}}{10}$$
 - Volume of 7.5% Sodium Bicarbonate needed = [(Volume of Collagen I, step a) x 0.0125 mL]
 - Volume of distilled water needed = [Total Volume - (sum of volumes from steps a+b+c)]
- In a sterile tube mix the 10X PBS, distilled water, and 7.5% Sodium Bicarbonate.
- Add the Cultrex 3-D Culture Matrix Rat Collagen I to the tube and pipette up and down to mix. **Do not vortex.**
- Place the diluted Cultrex 3-D Culture Matrix Rat Collagen I into the desired plates or dishes. This solution is stable for up to 1 hour on ice.

Note: *Plates may be centrifuged 300 x g for 10 minutes at 2-8 °C to prevent bubbles from forming in the gel.*
- Incubate the plate at 37 °C for 1 hour to promote gel formation.

High Concentration Gelling Procedure

- Place Cultrex 3-D Culture Matrix Rat Collagen I (5 mg/mL), 7.5% Sodium Bicarbonate solution, sterile tube, and cell culture plate on ice.
- Add necessary amount of Cultrex 3-D Culture Matrix Rat Collagen I into sterile tube.
- Add 5 µL of 7.5% Sodium Bicarbonate per 0.1 mL of Cultrex 3-D Culture Matrix Rat Collagen I (5 mg/mL).
- Pipette Cultrex 3-D Culture Matrix Rat Collagen I up and down to mix. **Do not vortex.**
- Pipette Cultrex 3-D Culture Matrix Rat Collagen into a cell culture plate.

Note: *Plate may be centrifuged for 300 x g for 10 minutes at 2-8 °C to prevent bubbles from forming in the gel.*
- Incubate the plate at 37 °C for 1 hour to promote gel formation.

DATA EXAMPLE

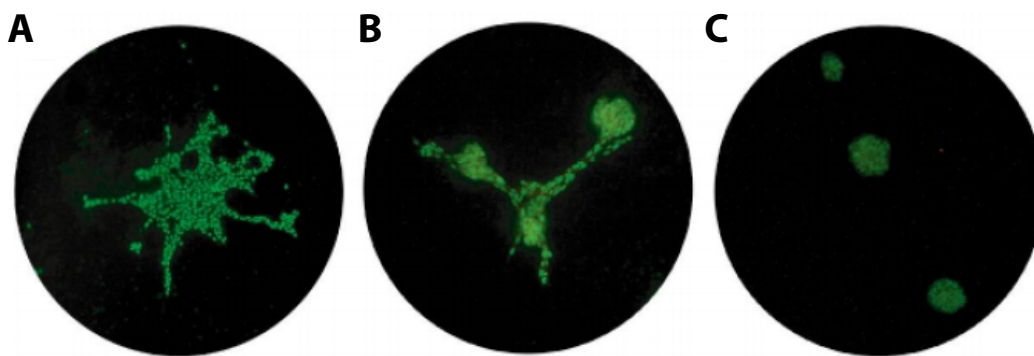


Figure 1: Differentiation of MCF-10A mammary epithelial cells cultured on Cultrex 3-D Culture Matrix Rat Collagen I. Mammary epithelial cells, MCF-10A cultured on 3-D Culture Matrix Collagen I are induced to differentiate with the addition of 3-D Culture Matrix Laminin I at: A) 0 mg/mL, B) 1 mg/mL, and C) 2 mg/mL.

REFERENCES

1. Debnath, J. *et al.* (2003) *Methods* **30**:256.
2. Chen, S. *et al.* (2003) *Stem Cells* **21**:281.
3. Kokenyesi, R. *et al.* (2003) *Gynecol. Oncol.* **89**:60.
4. Kutznetsova, N. *et al.* (1998) *Biochem.* **37**:11888.
5. Kutznetsova, N. and S. Leikin. (1999) *J. Bio. Chem.* **274**:36083.
6. Leikin, S. *et al.* (1994) *Proc. Natl. Acad. Sci. USA* **91**:276.
7. Leikina, E. *et al.* (2002) *Proc. Natl. Acad. Sci. USA* **99**:1314.
8. O' Shaughnessy, T. *et al.* (2003) *Neuroscience Letters* **340**:169.
9. Park, D. *et al.* (2003) *Cancer Letters* **195**:185.
10. Ritty, T. and J. Herzog (2003) *J. Ortho. Res.* **21**:442.
11. Van Oostveldt, K. *et al.* (2002) *J. Dairy Sci. Ass.* **85**:139.