

DESCRIPTION

Source Chinese Hamster Ovary cell line, CHO-derived human MGAT4A protein
Leu93-Asn535, with a C-terminal 6-His tag
Accession # Q9UM21

N-terminal Sequence Analysis Leu93

Predicted Molecular Mass 52 kDa

SPECIFICATIONS

SDS-PAGE 54-60 kDa, under reducing conditions.

Activity Measured by its ability to transfer N-acetyl- α -D-glucosamine from UDP-N-Acetyl- α -D-glucosamine to glycan Cy5-Fuc labeled N2f. Able to convert >85% of the substrate glycan N2f, as measured under the described conditions.

Endotoxin Level <0.10 EU per 1 μ g of the protein by the LAL method.

Purity >90%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.

Formulation Supplied as a 0.2 μ m filtered solution in Tris and NaCl. See Certificate of Analysis for details.

Activity Assay Protocol

- Materials**
- Assay Buffer: 50 mM Tris, 10 mM MnCl₂, pH 7.0
 - Recombinant Human MGAT4A His-tag (rhMGAT4A) (Catalog # 11496-GT)
 - Cy5-Fuc labeled N2f (Cy5-N2f) (Catalog # GL304)
 - UDP-GlcNAc, 50 mM stock in 50% Ethanol
 - 17% SDS-PAGE Gel
 - Gel loading dye
 - Gel Imager with Cy5 fluorescent dye detection capability

- Assay**
1. Dilute rhMGAT4A to 50 μ g/mL with Assay Buffer.
 2. Create a Reaction Mix containing 0.02 μ M Cy5-N2f and 1 mM UDP-GlcNAc in Assay Buffer.
 3. Combine 10 μ L of rhMGAT4A and 10 μ L of Reaction Mix. For a Control, combine 10 μ L of Assay Buffer and 10 μ L of Reaction Mix.
 4. Incubate reaction and control at 37 °C for 60 minutes.
 5. Add gel loading dye to each tube.
 6. Load half the volume of each reaction and control onto a 17% SDS-PAGE gel. Let samples migrate at least 80% down the gel before stopping.
 7. Acquire gel image and determine percent conversion of Cy5-N2f.

- Final Assay Conditions** Per Reaction:
- rhMGAT4A: 0.5 μ g
 - Cy5-N2f: 0.2 pmol
 - UDP-GlcNAc: 0.5 mM

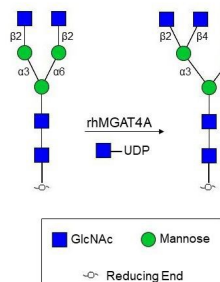
PREPARATION AND STORAGE

Shipping The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below.

- Stability & Storage** Use a manual defrost freezer and avoid repeated freeze-thaw cycles.
- 6 months from date of receipt, -20 to -70 °C as supplied.
 - 3 months, -20 to -70 °C under sterile conditions after opening.

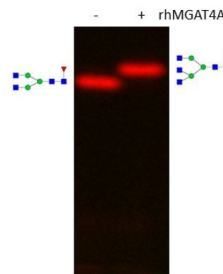
DATA

Enzyme Activity



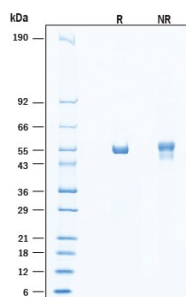
Recombinant Human MGAT4A His-tag Protein Enzyme Activity Diagram. Recombinant Human MGAT4A His-tag Protein (Catalog # 11496-GT) catalyzes a β 1-4GlcNAc linkage to the α 1-3Man arm of N-glycans.

Enzyme Activity



Recombinant Human MGAT4A His-tag Protein Enzyme Activity. Recombinant Human MGAT4A His-tag Protein (Catalog # 11496-GT) activity was measured by its ability to transfer N-acetyl- α -D-glucosamine from UDP-N-acetyl- α -D-glucosamine to glycan Cy5-Fuc labeled N2f (Catalog # GL304). Following treatment, a mobility shift was observed via SDS-PAGE gel.

SDS-PAGE



Recombinant Human MGAT4A His-tag Protein SDS-PAGE. 2 μ g/lane of Recombinant Human MGAT4A His-tag Protein (Catalog # 11496-GT) was resolved with SDS-PAGE under reducing (R) and non-reducing (NR) conditions and visualized by Coomassie® Blue staining, showing bands at 54-60 kDa, under reducing conditions.

BACKGROUND

Recombinant human Alpha-1,3-mannosyl-glycoprotein 4- β -N-acetylglucosaminyltransferase A (MGAT4A), also known as GlcNAc-T IVa, is a metal-dependent, golgi single-pass type II membrane protein that contains an N-terminal transmembrane region, a catalytic domain, and a C-terminal carbohydrate binding module that regulates its catalytic activity (1, 2). MGAT4A is one of several human MGAT enzymes involved in the initiation and synthesis of N-glycans, with each enzyme having unique specificity and preferences (3). MGAT4 has two human isozymes that catalyze formation of the β 1-4 GlcNAc branch in the α 1-3 Mannose arm of N-glycans (4-6). The two isozymes have different expression profiles in tissues and cancer cell lines, with MGAT4A expression within gastrointestinal tissues such as the pancreas (2, 4, 5). Loss of MGAT4A in mouse models has been shown to induce type 2 diabetes (9,10). Expression and activity of MGAT4A is particularly important during differentiation and oncogenesis (7, 8) including within choriocarcinoma, invasive mole, and placental site trophoblastic tumors (11-13). The discovery of MGAT4A modulators may lead to new therapeutics for the treatment of these diseases (10). MGAT4A may also be useful as a tool for the glycoengineering of complex N-glycans. The activity of MGAT4A was demonstrated using a fluorescent gel shift assay.

References:

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