

## DESCRIPTION

**Source** *E. coli*-derived *h. pylori* Fucosyltransferase protein  
Met1-Tyr392, with a C-terminal 6-His tag  
Accession # AAD07710.1

**N-terminal Sequence Analysis** Met1

**Predicted Molecular Mass** 46 kDa

## SPECIFICATIONS

**SDS-PAGE** 39 kDa, under reducing conditions.

**Activity** Measured by its ability to transfer fucose from GDP-fucose to *N*-Acetyllactosamine  
The specific activity is >1500 pmol/min/μg, as measured under the described conditions.

**Endotoxin Level** <1.0 EU per 1 μg of the protein by the LAL method.

**Purity** >95%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.

**Formulation** Supplied as a 0.2 μm filtered solution in Tris and NaCl. See Certificate of Analysis for details.

## Activity Assay Protocol

- Materials**
- Glycosyltransferase Activity Kit (Catalog # EA001)
  - Assay Buffer: 25 mM Tris, 10 mM CaCl<sub>2</sub>, pH 7.0
  - Recombinant *H. pylori* Fucosyltransferase (rHp FUT) (Catalog # 11072-GT)
  - Lactosamine (Dextra Laboratories, Catalog # GN204), 50 mM stock in deionized water
  - GDP-Fucose (Sigma, Catalog # G4401), 1.6 mM stock in deionized water
  - 96-well Clear Plate (Catalog # DY990)
  - Plate Reader (Model: SpectraMax Plus by Molecular Devices) or equivalent

- Assay**
- Dilute 1 mM Phosphate Standard provided by the Glycosyltransferase Activity Kit by adding 40 μL of the 1 mM Phosphate Standard to 360 μL of Assay Buffer for a 100 μM stock. This is the first point of the standard curve.
  - Complete the standard curve by performing six one-half serial dilutions of the 100 μM Phosphate stock in Assay Buffer. The standard curve has a range of 0.078 to 5 nmol per well.
  - Prepare Reaction Mixture containing 2.4 mM Lactosamine, 0.240 mM GDP-Fucose, and 4 μg/mL Coupling Phosphatase 1 (provided in kit) in Assay Buffer.
  - Dilute rHp FUT to 1 μg/mL in Assay Buffer.
  - Load 50 μL of each dilution of the standard curve into a plate. Include a curve blank containing 50 μL of Assay Buffer.
  - Load 25 μL of 1 μg/mL of rHp FUT into the plate. Include a Control containing 25 μL of Assay Buffer.
  - Start the reactions by adding 25 μL of Reaction Mixture to the wells, excluding the standard curve.
  - Incubate sealed plate at 37 °C for 20 minutes.
  - Add 30 μL of the Malachite Green Reagent A to all wells. Mix briefly.
  - Add 100 μL of deionized water to all wells.
  - Add 30 μL of the Malachite Green Reagent B to all wells. Mix and incubate for 20 minutes at room temperature.
  - Read plate at 620 nm (absorbance) in endpoint mode.
  - Calculate specific activity:

$$\text{Specific Activity (pmol/min/μg)} = \frac{\text{Phosphate released* (nmol)} \times (1000 \text{ pmol/nmol})}{\text{Incubation time (min)} \times \text{amount of enzyme (μg)}}$$

\*Derived from the phosphate standard curve using linear or 4-parameter fitting and adjusted for Control.

- Final Assay Conditions**
- Per Reaction:
- rHp FUT: 0.025 μg
  - Coupling Phosphatase 1: 0.1 μg
  - Lactosamine: 1.2 mM
  - GDP-Fucose: 0.12 mM

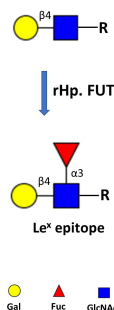
## PREPARATION AND STORAGE

**Shipping** The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below.

- Stability & Storage** Use a manual defrost freezer and avoid repeated freeze-thaw cycles.
- 6 months from date of receipt, -20 to -70 °C as supplied.
  - 3 months, -20 to -70 °C under sterile conditions after opening.

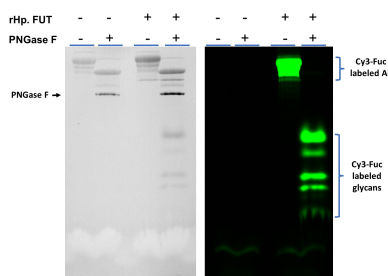
## DATA

#### Enzyme Activity



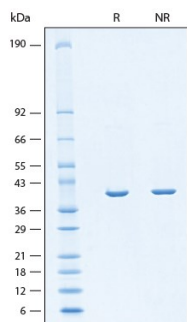
**Recombinant *H. pylori* Fucosyltransferase Protein**  
**Enzyme Activity Diagram.**  
Recombinant *H. pylori* Fucosyltransferase His-tag Protein (Catalog # 11072-GT)  
Enzyme Activity Diagram.

#### Enzyme Activity



**Fluorescent glycan labeling of asialofetuin (AF) with Cy3-Fuc using rHp. FUT.** Asialofetuin (4  $\mu$ g) was incubated with 0.2 nmol of GDP-Cy3-Fucose (Catalog # ES401) with or without rHp. FUT (Catalog # 11072) (1.0  $\mu$ g) in 25  $\mu$ L of 25 mM Tris pH 7.5, 10 mM MnCl<sub>2</sub> at 37 °C for 30 minutes. Following labeling, half of the AF was deglycosylated with PNGase F N-glycan Releasing Kit (Catalog # EA006) and resolved in 15% SDS-PAGE. Left side is the TCE image and right side is the Fluorescent image of the same gel.

#### SDS-PAGE



**Recombinant *H. pylori* Fucosyltransferase Protein**  
**SDS-PAGE.** 2  $\mu$ g/lane of Recombinant *H. pylori* Fucosyltransferase Protein (Catalog # 11072-GT) was resolved with SDS-PAGE under reducing (R) and non-reducing (NR) conditions and visualized by Coomassie® Blue staining, showing bands at 39 kDa.

#### BACKGROUND

Lewis X ( $\text{Le}^{\text{X}}$ ), a fucosylated trisaccharide glycan epitope ( $\text{Gal}\beta 1,4 [\text{Fuc}\alpha 1,3]\text{GlcNAc}\beta$ ) also known as CD15, and sialylated Lewis X ( $\text{sLe}^{\text{X}}$ ) are distributed throughout eukaryotes and are determinants of many functional glycoconjugates that play central roles in numerous physiological and pathological processes (1, 2).  $\text{Le}^{\text{X}}$ -bearing glycans are also found in the infectious bacterium *Helicobacter pylori* to mask the bacterium from the host immune surveillance (3). *H. pylori* is the pathogen that causes peptic ulcers that can further lead to stomach cancer and gastritis. *H. pylori* fucosyltransferase is responsible for generating the  $\text{Le}^{\text{X}}$  glycans, therefore is potentially a drug target for curing peptic ulcers, gastritis and stomach cancer. In molecular terms, *H. pylori* fucosyltransferase is an  $\alpha 1,3$  fucosyltransferase that shares function with human FUT4, FUT5, FUT6, and FUT9 (4). Remarkably, *H. pylori* fucosyltransferase can transfer IgG antibody to the glycocalyx on the surfaces of live cells when the antibody is conjugated to the enzyme's natural donor substrate GDP-Fucose (5). The activity of this enzyme has been measured with a phosphatase coupled method (6).

#### References:

1. Gooi, H.C. *et al.* (1981) *Nature* **292**:156.
2. Phillips, M.L. *et al.* (1990) *Science* **250**:1130.
3. Moran, A.P. *et al.* (1996) *FEMS Immunol Med Microbiol* **16**:105.
4. de Vries, T. *et al.* (2001). *Glycobiology* **11**:119R.
5. Li, J. *et al.* (2018) *ACS Cent. Sci.* **4**:1633.
6. Wu, Z.L. *et al.* (2011) *Glycobiology* **21**:727.