

DESCRIPTION

Source	<i>E. coli</i> -derived <i>f. tularensis</i> Cpf1 protein			
	APKKRKRKVGIHGVPA	<i>F. tularensis</i> Cpf1 (Ser2-Asn1300) Accession # WP_003034647.1	KRPAATKKAGQAKKKKGG	HHHHHH
	N-terminus			C-terminus
N-terminal Sequence Analysis	Ala			
Predicted Molecular Mass	156 kDa			

SPECIFICATIONS

SDS-PAGE	110-135 kDa, under reducing conditions
Activity	Measured by its ability to cleave a targeted DNA substrate. <i>rF. tularensis</i> Cpf1 achieves >80% substrate cleavage, as measured under the described conditions.
Endotoxin Level	<0.10 EU per 1 µg of the protein by the LAL method.
Purity	>90%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.
Formulation	Supplied as a 0.2 µm filtered solution in Tris, NaCl, EDTA, Glycerol and TCEP. See Certificate of Analysis for details.

Activity Assay Protocol

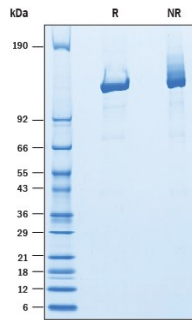
Materials	<ul style="list-style-type: none"> Assay Buffer: 50 mM NaCl, 10 mM Tris-HCl, 10 mM MgCl₂, 100 µg/ml BSA, pH 7.9 Recombinant <i>F. tularensis</i> Cpf1 (<i>rF.t.Cpf1</i>) (Catalog # 10343-C1) DNA Substrate: PBR322 vector (NEB, Catalog # N3033S) digested with EcoRI-HF (NEB, Catalog # R3101S)* Integrated DNA Technologies (IDT) Alt-R Cpf1 crRNA, targeting sequence: TGCCGCCTCGGCGAGCACAT Ultrapure DNase/RNase-Free Distilled Water (Invitrogen, Catalog # 10977015), to prepare Assay Buffer DNA gel TAE Buffer, 25X Liquid Concentrate (VWR, Catalog # 97062-386) Ethidium Bromide, 10 mg/mL (Amresco, Catalog # X328) <p>*Digest was gel purified using gel purification kit and eluted in EB buffer (10 mM Tris-HCl, pH 8.5).</p>
Assay	<ol style="list-style-type: none"> Prepare RNP Complex: <ol style="list-style-type: none"> 200 nM crRNA (2 µL addition from 3 µM stock prepared in Assay Buffer) 0.35 µg <i>rF.t.Cpf1</i> Add Assay Buffer for a final RNP Complex volume of 23 µL Incubate for 5 minutes at 25 °C Mix RNP Complex with 7 µL of 8.6 ng/µL of DNA Substrate (diluted in Assay Buffer, if possible). Incubate for 20 minutes at 37 °C. Incubate for 10 minutes at 65 °C to dissociate enzyme from DNA. Load total reaction with loading dye on a 1.25% agarose gel. Run gel at 140V for 40 minutes. Soak gel in 200 mL of 1X TAE Buffer with 150 µL of 10 mg/mL Ethidium Bromide for 1 hour. Use imaging software to detect and quantify hydrolysis of the DNA substrate.
Final Assay Conditions	<p>Per Reaction:</p> <ul style="list-style-type: none"> <i>rF.t.Cpf1</i>: 0.35 µg DNA Substrate: 60 ng crRNA: 200 nM

PREPARATION AND STORAGE

Shipping	The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below.
Stability & Storage	<p>Use a manual defrost freezer and avoid repeated freeze-thaw cycles.</p> <ul style="list-style-type: none"> 6 months from date of receipt, -20 to -70 °C as supplied. 3 months, -20 to -70 °C under sterile conditions after opening.

DATA

SDS-PAGE



2 µg/lane of Recombinant Human *F. tularensis* Cpt1 Protein (Catalog # 10343-C1) was resolved with SDS-PAGE under reducing (R) and non-reducing (NR) conditions and visualized by Coomassie® Blue staining, showing bands at 110-135 kDa.

BACKGROUND

Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR)-associated endonuclease from *Prevotella* and *Francisella* 1, Cpf1, also known as Cas12a, is a 1200-1500 amino-acids long monomeric protein that belongs to the CRISPR/Cas system (1, 2), an adaptive immune system of prokaryotes that has now become a powerful tool for genome editing (3). CRISPR/Cpf1 belongs to the class II (type 5) of the CRISPR/Cas system that is defined by a single-subunit effector (4). Cpf1 has recently emerged as an alternative for Cas9, due to its distinct features (2, 5) such as the ability to target T-rich motifs, no need for trans-activating crRNA, inducing a staggered double-strand break and potential for both RNA processing and DNA nuclease activity. In addition, Cpf1 is able to process more structured pre-CRISPR/RNA(crRNA) molecules into mature crRNAs (6) which allows the possibility to use both mature or pre-crRNA for genome editing purposes(7). All these features make the CRISPR-Cpf1 system a valuable genome-engineering tool (8). CRISPR-Cpf1(Cas12a) has been successfully used to edit genomes in mammalian cells (2), plants (9), mice (10), *Drosophila* (11) and recently zebrafish and *Xenopus* (7). Two Cpf1 orthologs have been commonly used for genome editing in different organisms: AsCpf1 and LbCpf1, which are derived from *Acidaminococcus* sp. BV3L6 and *Lachnospiraceae* bacterium ND2006, respectively (8). The attached nuclear localization signals (NLSs) on the chimeric protein ensures nuclear compartmentalization in cells during gene editing (12).

References:

1. Jinek, M. *et al.* (2012) *Science* **337**:816.
2. Zetsche, B. *et al.* (2015) *Cell* **163**:759.
3. Sandler J.D. and Joung J.K. (2014) *Nat Biotechnol.* **32**:347.
4. Koonin E.V. *et al.* (2017) *Curr. Opin. Microbiol.* **37**:67.
5. Bayat H. *et al.* (2018) *Curr. Microbiol.* **75**:107.
6. Fonfara, I. *et al.* (2016) *Nature* **532**:517.
7. Moreno-Mateo, M.A. *et al.* (2017) *Nat. Commun* **8**:2024.
8. Joung K.J. *et al.* (2016) *Nat. Biotechnol.* **34**:869.
9. Kim, H *et al.* (2017) *Nat. Commun.* **8**:14406.
10. Kim, Y. *et al.* (2016) *Nat. Biotechnol.* **34**:808.
11. Port, F. and Bullock, S.L. (2016) *Nat Methods* **13**:852.
12. Cong, L. *et al.* (2013) *Science* **339**:819.