

**DESCRIPTION**

<b>Source</b>	Mouse myeloma cell line, NS0-derived Ile21-Leu410, with a C-terminal 10-His tag Accession # Q3UCD9
<b>N-terminal Sequence Analysis</b>	Ile21
<b>Structure / Form</b>	Pro form
<b>Predicted Molecular Mass</b>	44 kDa

**SPECIFICATIONS**

<b>SDS-PAGE</b>	52 kDa, reducing conditions
<b>Activity</b>	Measured by its ability to cleave the fluorogenic peptide substrate, Mca-PLGL-Dpa-AR-NH <sub>2</sub> (Catalog # ES001). The specific activity is >600 pmol/min/μg, as measured under described conditions.
<b>Endotoxin Level</b>	<1.0 EU per 1 μg of the protein by the LAL method.
<b>Purity</b>	>95%, by SDS-PAGE under reducing conditions and visualized by silver stain.
<b>Formulation</b>	Lyophilized from a 0.2 μm filtered solution in MES and NaCl. See Certificate of Analysis for details.

**Activity Assay Protocol**

<b>Materials</b>	<ul style="list-style-type: none"> <li>● Assay Buffer: 0.1 M Sodium Acetate, 0.2 M NaCl, pH 3.5</li> <li>● Recombinant Mouse Cathepsin D (rmCathepsin D) (Catalog # 1029-AS)</li> <li>● Substrate: MCA-Pro-Leu-Gly-Leu-Dpa-Ala-Arg-NH<sub>2</sub> (Catalog # ES001)</li> <li>● F16 Black Maxisorp Plate (Nunc, Catalog # 475515)</li> <li>● Fluorescent Plate Reader (Model: SpectraMax Gemini EM by Molecular Devices) or equivalent</li> </ul>
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<b>Assay</b>	<ol style="list-style-type: none"> <li>1. Dilute rmCathepsin D to 20 μg/mL in Assay Buffer.</li> <li>2. Incubate at RT for 10 minutes.</li> <li>3. Dilute activated rmCathepsin D to 0.4 ng/μL in Assay Buffer.</li> <li>4. Dilute Substrate to 20 μM in Assay Buffer.</li> <li>5. Load 50 μL of the 0.4 ng/μL rmCathepsin D in a plate, and start the reaction by adding 50 μL of 20 μM Substrate. Include a Substrate Blank containing 50 μL Assay Buffer and 50 μL of 20 μM Substrate.</li> <li>6. Read at excitation and emission wavelengths of 320 nm and 405 nm (top read), respectively in kinetic mode for 5 minutes.</li> <li>7. Calculate specific activity:</li> </ol> $\text{Specific Activity (pmol/min/}\mu\text{g)} = \frac{\text{Adjusted } V_{\text{max}}^* \text{ (RFU/min)} \times \text{Conversion Factor}^{**} \text{ (pmol/RFU)}}{\text{amount of enzyme (}\mu\text{g)}}$ <p>*Adjusted for Substrate Blank **Derived using calibration standard MCA-Pro-Leu-OH (Bachem, Catalog # M-1975)</p>
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<b>Final Assay Conditions</b>	Per Well: <ul style="list-style-type: none"> <li>● rmCathepsin D: 0.020 μg</li> <li>● Substrate: 10 μM</li> </ul>
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**PREPARATION AND STORAGE**

<b>Reconstitution</b>	Reconstitute at 100 μg/mL in sterile, deionized water.
<b>Shipping</b>	The product is shipped with polar packs. Upon receipt, store it immediately at the temperature recommended below.
<b>Stability &amp; Storage</b>	Use a manual defrost freezer and avoid repeated freeze-thaw cycles. <ul style="list-style-type: none"> <li>● 6 months from date of receipt, -20 to -70 °C as supplied.</li> <li>● 3 months, -20 to -70 °C under sterile conditions after reconstitution.</li> </ul>

**BACKGROUND**

Cathepsin D is a lysosomal aspartic protease of the pepsin family (4). Mouse Cathepsin D is synthesized as a precursor protein, consisting of a signal peptide (residues 1-20), a propeptide (residues 21-64), and a mature chain (residues 65-410) (1-3). It is expressed in most cells and overexpressed in breast cancer cells (5). It is a major enzyme in protein degradation in lysosomes, and also involved in the presentation of antigenic peptides. Mice deficient in this enzyme showed a progressive atrophy of the intestinal mucosa, a massive destruction of lymphoid organs, and a profound neuronal ceroid lipofucinos, indicating that Cathepsin D is essential for proteolysis of proteins regulating cell growth and tissue homeostasis (6). Cathepsin D secreted from human prostate carcinoma cells is responsible for the generation of angiostatin, a potent endogeneous inhibitor of angiogenesis (6).

**References:**

1. Diedrich, *et al.* (1990) Nucl. Acid Res. **18**:7184.
2. Grusby, *et al.* (1990) Nucl. Acid Res. **18**:4008.
3. Hetman, *et al.* (1994) DNA Cell Biol. **13**:419.
4. Conner (2004) in *Handbook of Proteolytic Enzymes* (Barrett, *et al.* eds) Elsevier Academic Press, San Diego, p. 43.
5. Rochefort, *et al.* (2000) Clin. Chim. Acta. **291**:157.
6. Tsukuba, *et al.* (2000) Mol. Cells **10**:601.