# PRODUCT INFORMATION & MANUAL

## Human EGF Valukine™ ELISA

## VAL111

For the quantitative determination of natural and
recombinant human EGF concentrations human EGF

For research use only.
Not for diagnostic or therapeutic procedures.

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Novus kits are guaranteed for 3 months from date of receipt

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# TABLE OF CONTENTS

# I. BACKGROUND

EGF (Epidermal Growth Factor; also Urogastrone) precursor is a 185 kDa group 1 member of the EGF family of growth factors (1-4). Group 1 members are molecules that bind to, and activate, the EGF receptor (EGF R). All EGF family members are synthesized as type I transmembrane (TM) proteins that are proteolytically cleaved to generate soluble forms. Human EGF is a fragment of a 1185 amino acid (aa) proform that contains a 1010 aa extracellular region, a 21 aa TM segment, and a 153 aa cytoplasmic domain. The proform extracellular domain (ECD) has three principal structural modules. There are nine class B LDL R repeats, one von Willebrand Factor A domain, and nine EGF-like repeats, the most membrane-proximal of which constitutes the mature 53 aa EGF molecule (aa's # 971-1023 of the preproprecursor) (5, 6). The transmembrane 185 kDa EGF proform undergoes proteolytic processing to generate multiple isoforms. Cleavage by ADAM10 releases a 160-170 kDa isoform (aa 21-1023) that is found in most body fluids (7-10). This is accompanied by the appearance of numerous 40-100 kDa fragments that may represent proteolytic degradation products (10). The process that generates the 6 kDa mature EGF molecule is unclear. It may arise internally, or be generated on the cell surface through the action of membrane-bound serine proteases that act on either the solubilized, 160 kDa proform, or a 70 kDa processed form of the 160 kDa proform (11, 12).

Notably, and in addition to mature EGF, both the 185 kDa TM, and proteolytically cleaved (but unprocessed) circulating, 160 kDa proform have bioactivity (7, 13, 14). The activity in both cases is attributed to the sole EGF peptide embedded in the precursor. None of the accompanying EGF-like motifs have activity on the EGF R (15). There are four potential alternative splice forms for the gene encoding EGF, none of which affect the mature EGF sequence. Two are in the ECD and show deletions of aa 913-953 and aa 314-355, respectively. Two others are in the cytoplasmic region and contain substitutions of 12 aa and 17 aa for aa 1125-1207 and aa 1136-1207, respectively. Mature human EGF is 70%, 70%, and 85% aa identical to mouse, rat and porcine EGF, respectively. Cells known to express EGF include platelets (16), cerebral neurons, astrocytes, and cerebellar Purkinje cells (3), cells of the Brunner (duodenum) and submandibular glands (17), nonpigmented ciliary epithelium (18), and cells of the anterior pituitary (19). EGF has a number of diverse physiological effects. A full appreciation of its activity is complicated by the fact that it operates through the EGF receptor, which is utilized by other EGF family members, heterodimerizes with other EGF R family members, and associates with other transmembrane proteins such as the PDGF Rβ and HGF receptor (16, 20, 21). In any event, EGF is proposed to affect both fetal and adult tissues. In the fetus, EGF influences thymocyte growth and differentiation at the double negative-to-double positive stage (22). It also seems to drive neuroglia production at the expense of neuron formation (3), and promote epithelization (23). Finally, it inhibits adipocyte maturation, thus increasing preadipocyte numbers (23). In the adult, EGF plays a role in mammary gland lactogenesis (24). It also causes fibroblast mitosis, ECM dissociation, and migration, general effects often associated growth factor activity (25).

The ligand-binding receptor for EGF is the EGF receptor (also known as HER1 and ErbB1) (16). Although uncertainty exists as to the exact mechanism for receptor activation, it is now suggested that one EGF molecule binds to one receptor molecule at two distinct sites. This forces a conformational change in the receptor that allows for its association with a second EGF-EGF R complex (26). This dimerization forms a functional EGF receptor. It is also known that ErbB2 heterodimerizes with EGF R, but ErbB2 does not itself bind EGF. This may be due to the fact that ErbB2 exists naturally in a form that will form a dimer, but contains a ligand-binding site that is inaccessible to ligands. Thus, it waits for an activated partner (EGF-EGF R) before it forms a functional EGF receptor (26). ErbB2:ErbB2 homodimers are precluded from forming due to an inherent electrostatic repulsion. EGF is also suggested to participate in ErbB3:ErbB2 heterodimer formation at high concentrations (27). The significance of this is unknown. Alternative splice forms of EGF R exist in tumor cells, and may contribute to either tumorigenesis or to sensitivity to EGF R inhibitors (28).

# II. OVERVIEW

## A. PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for EGF has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any EGF present is bound by the immobilized antibody. After washing away any unbound substances, an enzyme-linked polyclonal antibody specific for EGF is added to the wells. Following a wash to remove any unbound antibody-enzyme reagent, a substrate solution is added to the wells and color develops in proportion to the amount of EGF bound in the initial step. The color development is stopped and the intensity of the color is measured.

## B. LIMITATIONS OF THE PROCEDURE

- **FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.**
- This kit is suitable for cell culture supernate and serum.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples generate values higher than the highest standard, dilute the samples with Diluent and repeat the assay.
- Any variation in operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

# III. ADVANTAGES

## A. PRECISION

**Intra-assay Precision** (Precision within an assay)

Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

**Inter-assay Precision** (Precision between assays)

Three samples of know concentration were tested in forty separate assays to assess inter-assay precision. Assay were performed by at least three technicians using two lots of components.

| | Intra-assay Precision | | | Inter-assay Precision | | |
|---|---|---|---|---|---|---|
| Sample | 1 | 2 | 3 | 1 | 2 | 3 |
| N | 20 | 20 | 20 | 40 | 40 | 40 |
| Mean (pg/mL) | 17.8 | 53.1 | 97.8 | 18.4 | 52.1 | 90.3 |
| Standard Deviation | 0.519 | 1.12 | 2.08 | 1.02 | 1.53 | 3.74 |
| CV% | 2.9 | 2.1 | 2.1 | 5.5 | 2.9 | 4.1 |

## B. RECOVERY

The recovery of human EGF spiked to different levels throughout the range of the assay was evaluated. The recovery ranged from 96-103% with an average of 99%.

## C. SENSITIVITY

The minimum detectable dose (MDD) of EGF is typically 0.089-0.740 pg/mL. The mean MDD was 0.266 pg/mL.

The MDD was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

## D. CALIBRATION

This immunoassay is calibrated against highly purified *E. coli*-expressed recombinant human EGF produced at R&D Systems®.

## E. LINEARITY

To assess the linearity of the assay, different samples were containing or spiked with high concentrations of EGF and diluted with Diluent 1× to produce samples with values within the dynamic range of the assay.

| Dilution | Average % of Expected | Range (%) |
|---|---|---|
| 1:2 | 102 | 100 – 104 |
| 1:4 | 105 | 102 – 108 |
| 1:8 | 107 | 102 – 111 |
| 1:16 | 107 | 100 – 113 |

## F. SAMPLE VALUES

**Cell Culture Supernates -** Human peripheral blood mononuclear cells ($1\times10^6$ cells/mL) were cultured in RPMI supplemented with 10% fetal calf serum. Cell were cultured and stimulated with 10 µg/mL PHA for 6 days. An aliquot of the cell culture supernate was removed, assayed for levels of human EGF, and measured 5.4 pg/mL.

**Serum -** Four serum samples were evaluated for the presence of EGF in this assay. All samples measured ranged from 384.5 to 443.0 pg/mL with an average of 415.4 pg/mL.

## G. SPECIFICITY

This assay recognizes both natural and recombinant human EGF. The factors listed below were prepared at 50 ng/mL in Diluent 1x and assayed for cross-reactivity. Preparations of the following factors at 50 ng/mL in a mid-range rhEGF control were assayed for interference. No significant cross-reactivity or interference was observed.

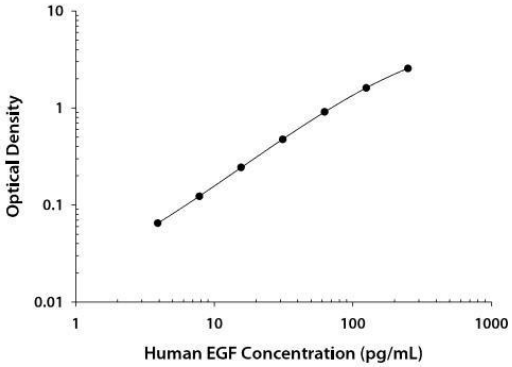
| **Recombinant Human** | | **Recombinant Mouse** | |
|---|---|---|---|
| Amphiregulin | HB-EGF | Amphiregulin | Epigen |
| Betacellulin | MFG-E8 | Betacellulin | Epiregulin |
| Cripto-1 | NRG1/HRG1 | Cripto | ErbB3 |
| EGF R | NRG1 Isoform SMDF | EGF | ErbB4 |
| Epigen | NRG1-α/HRG1-α EGF Domain | pro-EGF | MFG-E8 |
| Epiregulin | NRG1-HRG1-β1 EGF Domain | EGF R | |
| ErbB2 | NRG1-β1/HRG1-1β1 ECD | | |
| ErbB3 | TGF-α | | |
| ErbB4 | TNF-α | | |

Recombinant rat EGF cross-reacts approximately 1.0%, and recombinant human pro-EGF cross-reacts approximately 1.3% in this assay.

# IV. EXPERIMENT

## EXAMPLE STANDARD

The standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.




| (pg/mL) | O.D. | Average | Corrected |
|---|---|---|---|
| 0 | 0.010<br>0.012 | 0.011 | — |
| 3.91 | 0.073<br>0.079 | 0.076 | 0.065 |
| 7.81 | 0.132<br>0.135 | 0.134 | 0.123 |
| 15.6 | 0.247<br>0.262 | 0.255 | 0.244 |
| 31.3 | 0.474<br>0.496 | 0.485 | 0.474 |
| 62.5 | 0.918<br>0.926 | 0.922 | 0.911 |
| 125 | 1.612<br>1.627 | 1.620 | 1.609 |
| 250 | 2.537<br>2.599 | 2.568 | 2.557 |

# V. KIT COMPONENTS AND STORAGE

## A. MATERIALS PROVIDED

| Parts | Description | Amount |
|---|---|---|
| EGF Microplate | 96 well polystyrene microplate (12 strips of 8 wells) coated with a mouse monoclonal antibody against human EGF | 1 plate |
| EGF Conjugate | a solution of polyclonal antibody against EGF conjugated to horseradish peroxidase | 1 vial |
| EGF Standard | recombinant human EGF in a buffered protein base; lyophilized | 2 vials |
| Calibrator Diluent (5×) | a 5× concentrated buffered protein base | 1 vial |
| Wash Buffer concentrate (25×) | a 25× concentrated solution of buffered surfactant | 1 vial |
| Color Reagent A | stabilized hydrogen peroxide | 1 vial |
| Color Reagent B | stabilized chromogen (tetramethylbenzidine) | 1 vial |
| Stop Solution | 2 N sulfuric acid | 1 vial |
| Plate Covers | adhesive strip | 3 strips |

## B. STORAGE

<table>
<tr><td>Unopened Kit</td><td colspan="2">Store at 2-8°C not use past kit expiration date.</td></tr>
<tr><td rowspan="8">Opened/ Reconstituted Reagents</td><td>Diluted Wash Solution</td><td rowspan="6">May be stored for up to 1 month at 2-8°C.*</td></tr>
<tr><td>Stop Solution</td></tr>
<tr><td>Diluent 1×</td></tr>
<tr><td>Conjugate</td></tr>
<tr><td>Unmixed Color Reagent A</td></tr>
<tr><td>Unmixed Color Reagent B</td></tr>
<tr><td>Standard</td><td>Use a new standard for each assay. Discard after use.</td></tr>
<tr><td>Microplate Wells</td><td>Return unused wells to the foil pouch containing the desiccant pack, reseal along entire edge of zip-seal. May be stored for up to 1 month at 2-8°C.*</td></tr>
</table>

* Provided this is within the expiration date of the kit.

## C. OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- Pipettes and pipette tips.
- Deionized or distilled water.
- Polypropylene tubes for dilution of standards and samples.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinder.

## D. PRECAUTION

- EGF is detectable in saliva. Take precautionary measures to prevent contamination of kit reagents while running this assay.
- Some components in this kit contain ProClin which may cause an allergic skin reaction. Avoid breathing mist.
- Color Reagent B may cause skin, eye and respiratory irritation. Avoid breathing fumes. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.
- The Stop Solution provided with this kit is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

# VI. PREPARATION

## A. SAMPLE COLLECTION AND STORAGE

**Cell Culture Supernates** - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤-20°C. Avoid repeated freeze-thaw cycles. Samples may require dilution with Diluent 1×.

**Serum -** Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤-20°C. Avoid repeated freeze-thaw cycles.

## B. SAMPLE PREPARATION

Serum samples require a 5-fold dilution. A suggested 5-fold dilution is 40 µL of sample + 160 µL of Diluent (1×).

## C. REAGENT PREPARATION

**Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.**

***Note:*** *High concentrations of EGF are found in saliva. It is recommended that a face mask and gloves be used to protect kit reagents from contamination.*

**Wash Solution** - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate (25×) into deionized or distilled water to prepare 500 mL of Wash Buffer.

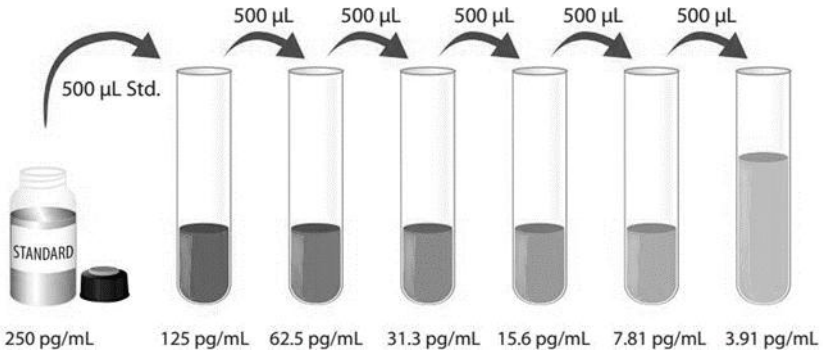
**Substrate Solution** - Color Reagent A and B should be mixed together in equal volumes within 15 minutes of use. Protect from light. 200 µL of the resultant mixture is required per well.

**Diluent 1×** - Add 20 mL of Calibrator Diluent (5×) into 80 mL of deionized or distilled water to prepare 100 mL of Diluent 1×.

**EGF Standard - Refer to the vial label for reconstitution volume*.** This reconstitution produces a stock solution of 250 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

*if you have any question, please seek help from our Technical Support.

**Use polypropylene tubes.** Pipette 500 µL of the Diluent 1× into each tube. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The undiluted standard serves as the high standard (250 pg/mL). The Diluent 1× serves as the zero standard (0 pg/mL).




## D. TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- It is recommended that the samples be pipetted within 15 minutes.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Substrate Solution should remain colorless until added to the plate. Keep Substrate Solution protected from light. Substrate Solution should change from colorless to gradations of blue.
- Stop Solution should be added to the plate in the same order as the Substrate Solution. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.

## VII.ASSAY PROCEDURE

***Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.***

***Note:*** *High concentrations of EGF are found in saliva. It is recommended that a face mask and gloves be used to protect kit reagents from contamination.*

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 200 µL of Standard, sample, or control per well. Cover with the adhesive strip provided. Incubate for 2 hours at room temperature. A plate layout is provided for a record of standards and samples assayed.
4. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Solution (400 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 200 µL of EGF Conjugate to each well. Cover with a new adhesive strip. Incubate for 1 hours at room temperature.
6. Repeat the aspiration/wash as in step 4.
7. Add 200 µL of Substrate Solution to each well. Incubate for 20 minutes at room temperature. **Protect from light**.
8. Add 50 µL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well within 30 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.
10. **CALCULATION OF RESULTS:** Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the EGF concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

## VIII. REFERENCES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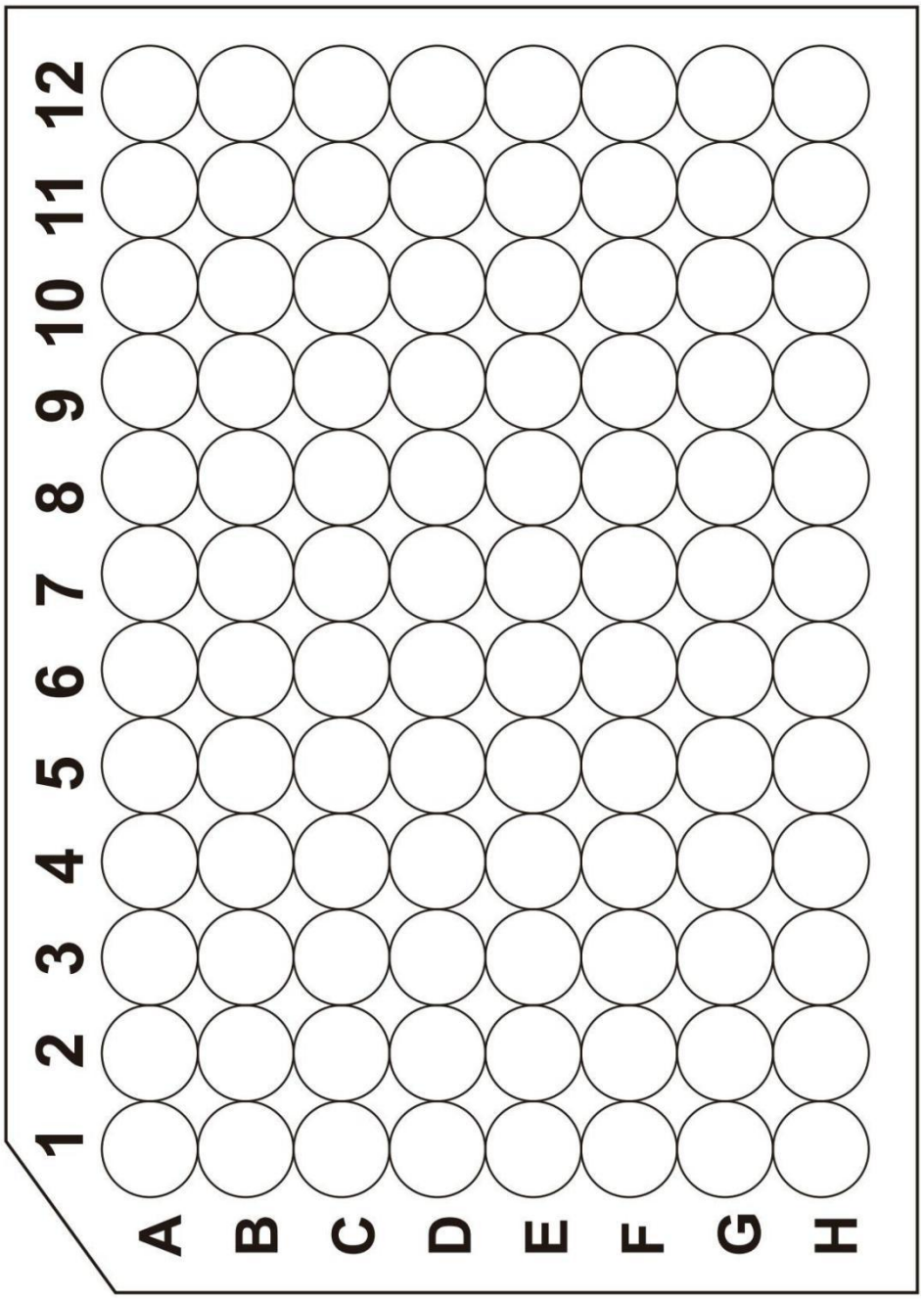
## PLATE LAYOUT

Use this plate layout to record standards and samples assayed.



# 产品信息及操作手册

## 人 EGF Valukine™ ELISA 试剂盒

## 目录号：VAL111

适用于定量检测天然和重组人表皮生长因子 EGF 的含量

科研专用，不可用于临床诊断

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Novus 试剂盒确保在你收货日期 3 个月内有效

# 目录

# I. 背景

EGF（表皮生长因子，又称 Urogastrone）前体是生长因子 EGF 家族的Group I 成员，分子量为185 kDa（1-4）。Group I成员是结合并活化表皮生长因子受体（EGF R）的分子。EGF 家族成员被合成为I型跨膜（TM）蛋白，且由蛋白水解生成可溶形式。人表皮生长因子（EGF）是一个长为1185个氨基酸前体分子形式的一个片段，包含1010个氨基酸细胞外区域、21个氨基酸 TM 区域和153个氨基酸胞质域。这个前体的胞外域（ECD）有三个主要的结构模块。其中包括 9个BLDLR 重复、1个血管性血友病因子 A 区域和 9个类EGF 重复，并在近膜区包含了成熟 EGF 分子的 53个氨基酸的结构（为前体形式的第971 位至1023 位氨基酸）（5, 6）。这个跨膜185 kDa的亚型（氨基酸21-1023），存在于大多数体液中（7-10）。这个过程同时产生了大量40-100 kDa 的蛋白片段，可能为蛋白水解降解产物（10）。6 kDa 成熟EGF 亚型的产生过程尚不清楚。它可能源自细胞内部，也可能源于细胞表面，通过细胞表面的膜结合丝氨酸酶水解可溶性的160 kDa 前体或这个前体水解以后生产的 70 kDa 的分子亚型（11, 12）而来。

值得注意的是，成熟形式的EGF、185 kDa 跨膜蛋白、已被水解但尚未加工的循环型分子和 160 kDa 前体分子都具有生物学活性（7, 13, 14）。而这些生物学活性主要是因为有EGF肽段嵌入在这些前体分子中。其它类型EGF的修饰都不具备结合EGF受体的活性（15）。编码EGF的基因在表达EGF 蛋白过程中存在 4 种可能的剪切体，但都不影响成熟EGF的序列。其中两种剪切体发生在胞外区序列，分别删除了第913位至953位的氨基酸和第314至355位的氨基酸。另外两种剪切体则涉及EGF的胞内区域，分别以12个氨基酸替换了第1125位至 1207位的氨基酸和以17个氨基酸替换了第1136-1207 位的氨基酸。成熟的人 EGF 与小鼠、大鼠和猪的EGF 的同源性分别是 70%、70%和 85%。已知表达EGF的细胞包括血小板（16）、大脑神经元、星状胶质细胞和小脑浦肯野细胞（3）、布鲁纳细胞（十二指肠）和颌下腺细胞（17）、不着色的纤毛上皮（18）和垂体前叶细胞（19）。EGF有许多不同的生理作用。目前广泛认可的EGF功能主要基于EGF受体-配体的结合而实现。而EGF受体也能和其它 EGF 家族的蛋白结合，进而形成异源多聚体并与其它 EGF 受体家族成员二聚化，或者和其它跨膜蛋白如PDGF R　和HGF 受体产生关联（16, 20, 21）。在任何一种情况下，EGF都可以对胎儿和成人组织产生作用。在胎儿，EGF 影响双阴性-双阳性阶段的胸腺细胞生长和分化（22）。在神经元形成过程中，EGF似乎也可以刺激神经胶质神经元的生成（3）并促进其上皮化（23）。最后，它能抑制脂肪细胞的成熟，从而增加前脂肪细胞数量。在成人，EGF 可以在乳腺的乳生成过程中发挥作用（24），也可使纤维细胞有丝分裂，ECM离解和迁移，广泛地影响生长因子相关的各种作用（25）。

EGF 受体即表皮生长因子受体（也称为HER1 和ErbB1）（16），其激活机制尚不明确。目前的研究认为每一个EGF分子和一个EGF 受体分子有两个结合位点，这会迫使受体产生构象变化，而和第二个 EGF-EGF R 复合物结合（26）。这种二聚体即具有实际功能的EGF受体。ErbB2 也可以和EGF受体形成异源二聚体，但不能和EGF结合。这可能是因为ErbB2天然以二聚体的形式存在，掩盖了其配体结合的位点，而使配体无法与其结合。因此，它需

要结合一个已经激活的EGF-EGF R 复合体才能形成一个功能性的EGF受体（26）。而由于内在的静电斥力作用，ErbB2 自身无法形成同源二聚体。 EGF 也只有在高浓度的环境下也能和ErbB3：ErbB2 形成异源多聚体。这个过程的重要性，目前仍是未知的。EGF R的选择性剪切亚型存在于肿瘤细胞中，并可能参与肿瘤发生或使细胞对EGF 受体抑制剂敏感（28）。

# II. 概述

## A. 检测原理

本实验采用双抗体夹心 ELISA 法。抗人EGF 单抗包被于微孔板上，样品和标准品中的 EGF 会与固定在板上的抗体结合，游离的成分被洗去；加入辣根过氧化酶标记的抗人EGF 多抗，未结合的抗体被洗去；加入底物溶液（显色剂），溶液颜色与结合的目标蛋白成正比；加入终止液；用酶标仪测定吸光度。

## B. 检测局限

- **仅供科研使用，不可用于体外诊断；**
- 该试剂盒适用于细胞培养上清样本和血清样本；
- 请在试剂盒有效期内使用；
- 不同试剂盒及不同批号试剂盒的组分不能混用；
- 样本值若大于标准曲线的最高值，应将样本用稀释剂（1×）稀释后重新检测； 若细胞培养上清液样本需分布稀释，除最后一步用稀释剂稀释外，其它中间稀释可采用细胞培养基；
- 检测结果的不同可由多种因素引起，包括实验人员的操作、移液器的使用方式、洗板技术、反应时间或温度、试剂盒的储存等。

# III. 优势

## A. 精确度

**板内精确度**（同一板内不同孔间的精确度）

已知浓度的三个样本，在同一板内分别检测 20 次，以确定板内精确度。

**板间精确度**（不同板之间的精确度）

已知浓度的三个样本，在不同板间分别检测 40 次，以确定板间精确度。

| | 板内精确度 | | | 板间精确度 | | |
|---|---|---|---|---|---|---|
| 样本 | 1 | 2 | 3 | 1 | 2 | 3 |
| 数量 | 20 | 20 | 20 | 40 | 40 | 40 |
| 平均值 (pg/mL) | 17.8 | 53.1 | 97.8 | 18.4 | 52.1 | 90.3 |
| 标准差 | 0.519 | 1.12 | 2.08 | 1.02 | 1.53 | 3.74 |
| CV% | 2.9 | 2.1 | 2.1 | 5.5 | 2.9 | 4.1 |

## B. 回收率

在细胞培养基样本中掺入检测范围内不同水平的人EGF，测定其回收率。回收率范围在 96-103%，平均回收率在 99%。

## C. 灵敏度

人 EGF 的最低可测剂量（MDD）一般在 0.089-0.740 pg/mL。

MDD 是根据20 个重复的零标准品孔的吸光度值的平均值加两倍标准差计算得到的相对应浓度。

## D. 校正

此ELISA 试剂盒经由R&D Systems ®生产的大肠杆菌表达的高纯度重组人EGF 蛋白校正。

## E. 线性

不同的样本中含有或掺入高浓度的人EGF，然后用稀释剂（1×）将样本稀释到检测范围内，测定其线性。

| 稀释倍数 | 平均值/期待值（%） | 范围 (%) |
|---|---|---|
| 1:2 | 102 | 100 – 104 |
| 1:4 | 105 | 102 – 108 |
| 1:8 | 107 | 103 – 111 |
| 1:16 | 107 | 100 – 113 |

## F. 样本预值

**细胞上清样本 -** 人的外周血单核细胞（1×$10^6$ 细胞/mL）培养于含有 10%胎牛血清的 RPMI1640 培养基中，细胞培养基还含有 2 mM L-谷氨酰胺、50 µM β-巯基乙醇、100 U/mL 青霉素, 100 µg/mL 链霉素，另加 10 µg/mL PHA 刺激细胞，培养 6 天。取细胞上清液测定 EGF 含量，结果为 5.4 pg/mL。

**血清样本 -** 使用本试剂盒检测了 4 份人血清样本中EGF 的水平。4份样本的检测值在384.5 – 443.0 pg/mL之间，平均值为415.4 pg/mL。

## G. 特异性

此ELISA 法可检测天然及重组人EGF 蛋白。将以下因子用稀释剂（1×）配置成 50 ng/mL 的浓度来检测与人 EGF 的交叉反应。将 50 ng/mL 的干扰因子掺入中间范围的重组人 EGF 对照品中，来检测对人EGF 的干扰。没有观察到明显的交叉反应或干扰。

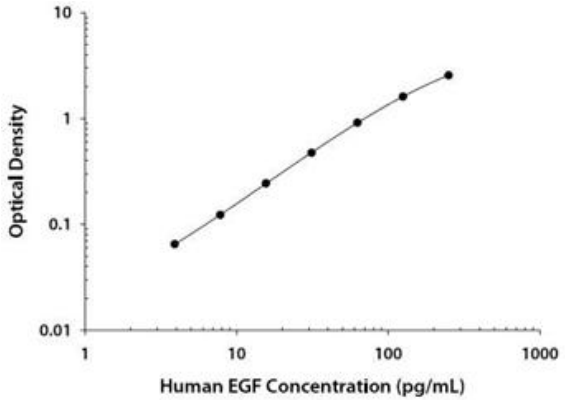
| 重组人蛋白 | | 重组小鼠蛋白 | |
|---|---|---|---|
| Amphiregulin | HB-EGF | Amphiregulin | Epigen |
| Betacellulin | MFG-E8 | Betacellulin | Epiregulin |
| Cripto-1 | NRG1/HRG1 | Cripto | ErbB3 |
| EGF R | NRG1 Isoform SMDF | EGF | ErbB4 |
| Epigen | NRG1-α/HRG1-α EGF Domain | pro-EGF | MFG-E8 |
| Epiregulin | NRG1-β1/HRG1-β1 EGF Domain | EGF R | |
| ErbB2 | NRG1-β1/HRG1-β1ECD | | |
| ErbB3 | TGF-α | | |
| ErbB4 | TNF-α | | |

此方法分析检测大鼠 EGF 交叉反应约 1.0%，与重组人 Pro-EGF 交叉反应约 1.3%。

# IV. 实验标准

## 标准曲线实例

该标准曲线数据仅供参考，每次实验应绘制其对应的标准曲线。




| (pg/mL) | O.D. | Average | Corrected |
|---|---|---|---|
| 0 | 0.010<br>0.012 | 0.011 | — |
| 3.91 | 0.073<br>0.079 | 0.076 | 0.065 |
| 7.81 | 0.132<br>0.135 | 0.134 | 0.123 |
| 15.6 | 0.247<br>0.262 | 0.255 | 0.244 |
| 31.3 | 0.474<br>0.496 | 0.485 | 0.474 |
| 62.5 | 0.918<br>0.926 | 0.922 | 0.911 |
| 125 | 1.612<br>1.627 | 1.620 | 1.609 |
| 250 | 2.537<br>2.599 | 2.568 | 2.557 |

# V. 试剂盒组成及储存

## A. 试剂盒组成

| 组成 | 描述 | 规格 |
|---|---|---|
| EGF Microplate | 包被抗体的 96 孔聚苯乙烯板，8 孔×12 条 | 1 块板 |
| EGF Conjugate | 酶标检测EGF 抗体 | 1 瓶 |
| EGF Standard | 标准品（冻干粉） | 2 瓶 |
| Calibrator Diluent （5×） | 浓缩稀释剂（5×） | 1 瓶 |
| Wash Buffer Concentrate（25×） | 浓缩洗涤缓冲液（25×） | 1 瓶 |
| Color Reagent A | 显色液A | 1 瓶 |
| Color Reagent B | 显色液B | 1 瓶 |
| Stop Solution | 终止液 | 1 瓶 |
| Plate Covers | 封板胶纸 | 3 张 |

## B. 试剂盒储存

<table>
<tr><td>未开封试剂盒</td><td colspan="2">2-8℃储存；请在试剂盒有效期内使用</td></tr>
<tr><td rowspan="8">已打开，稀释或重溶的试剂</td><td>洗涤缓冲液（1×）</td><td rowspan="6">2-8℃储存，最多 30 天*</td></tr>
<tr><td>终止液</td></tr>
<tr><td>稀释剂 1×</td></tr>
<tr><td>酶标检测抗体</td></tr>
<tr><td>显色剂 A</td></tr>
<tr><td>显色剂 B</td></tr>
<tr><td>标准品</td><td>现用现配；使用后弃掉。</td></tr>
<tr><td>包被的微孔板条</td><td>将未用的板条放回带有干燥剂的铝箔袋内，密封；2-8℃储存，最多 30 天*</td></tr>
</table>

*必须在试剂盒有效期内

## C. 实验所需自备试验器材

- 酶标仪（可测量 450 nm 检测波长的吸收值及 540 nm 或 570 nm 校正波长的吸收值）
- 高精度加液器及一次性吸头
- 蒸馏水或去离子水
- 洗瓶（喷瓶）、多通道洗板器或自动洗板机
- 500 mL 量筒

## D. 注意事项

试剂盒中的终止液是酸性溶液，使用时请做好眼镜、手、面部及衣服的防护。

# VI. 实验前准备

## A. 样品收集及储存

以下样本收集和保存为常规收集和保存方法，样本稳定性未经测试。

**细胞培养上清：**离心去除颗粒物。样本可以立即分析，或分装后保存于<-20℃。避免反复冻融。

**血清样本：**用血清分离管 (SST) 分离血清。使血样室温凝集 30 分钟，然后1000x g 离心15分钟。吸取血清样本之后即刻用于检测，或者分装，-20℃贮存备用。避免反复冻融。

## B. 样本准备工作

血清样本需要用稀释剂（1×）5倍稀释后进行检测，即40 µL血清+160 µL稀释剂（1×）。

## C. 检测前准备工作

**使用前请将所有试剂和样本放置于室温，建议所有的实验样本和标准品做复孔检测**

***注：****唾液中含有高浓度的EGF，为避免污染，实验时请带口罩、手套。*

**洗涤液：**从冰箱中取出的浓缩洗涤液可能有结晶，属于正常现象；放置室温，轻摇混匀，待结晶完全溶解后再配制洗涤液。可将 20 mL 浓缩洗涤液用蒸馏水或去离子水稀释配置成 500 mL 工作浓缩的洗涤液。未用完的放回 4℃。

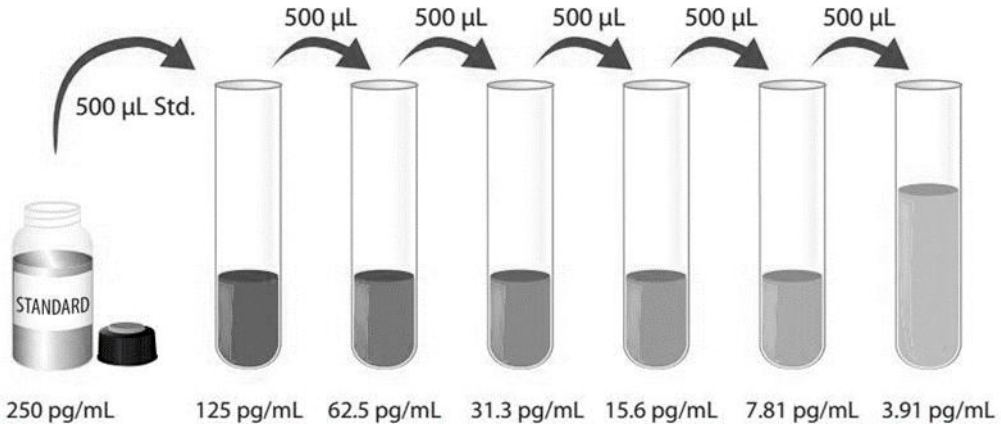
**显色剂：**按当次试验所需要用量将显色剂 A 和显色剂B 等体积混合，避光；在使用前 15 分钟内准备，仅供当日使用；每孔需 200 µL。

**稀释剂（1×）：**可将 20 mL 浓缩稀释剂用 80 mL 蒸馏水或去离子水稀释配置成 100 mL 工作浓度的稀释剂。

**标准品：**依照标准品标签上注明的重溶体积重溶冻干标准品，得到浓度为 250 pg/mL 标准品母液。轻轻震摇至少 15 分钟，其充分溶解。

*如有疑问，请咨询我们的技术支持。

使用聚丙烯试管作为标准品稀释管。每个稀释管中加入 500 µL 稀释剂（1×）。将标准品母液参照下图做系列稀释，每管须充分混匀后再移液到下一管。没有稀释的标准品母液可用作标准曲线最高点（250 pg/mL），稀释剂（1×）可用作标准曲线零点（0 pg/mL）。




## D. 技术小提示

- 当混合或重溶蛋白液时，尽量避免起沫；
- 为了避免交叉污染，配置不同浓度标准品、上样、加不同试剂都需要更换枪头。另外不同试剂请分别使用不同的移液槽；
- 建议 15 分钟内完成一块板的上样；
- 每次孵育时，正确使用封板胶纸可保证结果的准确性；
- 混合后的显色底物在上板前应为无色，请避光保存；加入微孔板后，将由无色变成不同深度的蓝色；
- 终止液上板顺序应同显色底物上板顺序一致；加入终止液后，孔内颜色由蓝变黄；若孔内有绿色，则表明孔内液体未混匀请充分混合。

## VII. 操作步骤

**使用前请将所有试剂和样本放置于室温，建议所有的实验样本和标准品做复孔检测**

1. 按照上一节的说明，准备好所有需要的试剂和标准品；
2. 从已平衡至室温的密封袋中取出微孔板，未用的板条请放回铝箔袋内，重新封口；
3. 分别将不同浓度标准品，实验样本或者质控品加入相应孔中，每孔 200 µL。用封板胶纸封住反应孔，室温孵育 2 小时。说明书提供了一张 96 孔模板图，可用于记录标准品和试验样本的板内位置；
4. 将板内液体吸去，使用洗瓶、多通道洗板器或自动洗板机洗板。每孔加洗涤液 400 µL，然后将板内洗涤液吸去。重复操作 4 次。每次洗板尽量吸去残留液体会有助于得到好的实验结果。最后一次洗板结束，请将板内所有液体吸干或将板倒置，在吸水纸拍干所有残留液体；
5. 在每个微孔内加入 200 µL 酶标检测抗体。用封板胶纸封住反应孔，室温孵育 1 小时；
6. 重复第 4 步洗板操作；
7. 在每个微孔内加入 200 µL 显色底物，室温孵育 20 分钟。**注意避光；**
8. 在每个微孔内加入 50 µL 终止液，孔内溶液颜色会从蓝色变为黄色。如果溶液颜色变为绿色或者颜色变化不一致，请轻拍微孔板，使溶液混合均匀；
9. 加入终止液后 30 分钟内，使用酶标仪测量 450 nm 的吸光度值，设定 540 nm 或 570nm 作为校正波长。如果没有使用双波长校正，结果准确度可能会受影响；
10. **计算结果：**将每个标准品和样品的校正吸光度值 (OD450-OD540/OD570)、复孔读数取平均值，然后减去平均零标准品 OD 值。使用计算机软件作四参数逻辑 (4-PL) 曲线拟合创建标准曲线。另一种方法是，可以通过绘制标准品浓度做对数与相应 OD 值对数生成曲线，并通过回归分析确定最佳拟合线。这个过程可生成一个足够使用但不太精确的数据拟合。若样本经过稀释，计算浓度时应乘以稀释倍数。

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## 96 孔模板图

请使用 96 孔模板图来记录标准品及样本在板内的位置