



PRODUCT INFORMATION & MANUAL

Human FGF-19 Valukine™ ELISA

Catalog Number: VAL148

For the quantitative determination of natural and recombinant
Human FGF-19 concentrations

For research use only.
Not for diagnostic or therapeutic procedures.

Bio-Techne China Co. Ltd

P: +86 (21) 52380373 P: 8009881270 F: +86 (21) 52381001
info.cn@bio-techne.com

Please refer to the kit label for expiry date.
Novus kits are guaranteed for 3 months from date of receipt

Version 202409.3

TABLE OF CONTENTS

I. BACKGROUND	2
II. OVERVIEW	3
III. ADVANTAGES	4
IV. EXPERIMENT	6
V. KIT COMPONENTS AND STORAGE	7
VI. PREPARATION	10
VII. ASSAY PROCEDURE	12
VIII. REFERENCES	14

I. BACKGROUND

The Fibroblast Growth Factor (FGF) family consists of at least 22 highly conserved proteins and are found in many species ranging from *C. elegans* and *Drosophila* to humans (1). FGFs are heparin-binding growth factors with a core 120 amino acid (aa) FGF domain that allows for a common tertiary structure. In general, FGFs are expressed during embryonic development and in restricted adult tissues. They act on cells of mesodermal and neuroectodermal origin to regulate diverse physiologic functions including angiogenesis, cell growth, pattern formation, metabolic regulation, cell migration, neurotrophic effects, and tissue repair (2). The activities of the FGF family are mediated by four receptors, FGF R1 through FGF R4 (2). The receptors are transmembrane proteins with a tyrosine kinase domain and are thought to act as classical receptor tyrosine kinases. FGF R5/FGF RL1 has also been described, but lacks the kinase domain and signaling capability (3).

Human FGF-19 cDNA predicts a 251 aa precursor protein with a 22 aa signal peptide and a 229 aa secreted mature protein with no potential N-linked glycosylation sites (4, 5). It shares approximately 50% aa sequence identity with mouse and rat FGF-15 and 62% aa sequence identity with chicken FGF-19 (4-6). Unlike most FGFs, which bind to and activate more than one FGF receptor, FGF-19 appears to bind only FGF R4 (5). Receptor binding is capable of activating MAP Kinase and inducing the Prolactin promoter (7). FGF-19 has two putative disulfide bonds, one of which is conserved in the FGF family (8). Uniquely, it also has an extended loop structure that may affect heparan sulfate binding (8).

The functional roles of FGF-19 continue to be elucidated and include developmental, metabolic, and tumorigenic activities. During chick embryogenesis, FGF-19 has been shown to act synergistically with Wnt-8c to initiate inner ear development (6). However, the mouse FGF-19 ortholog, FGF-15, is not required for this activity (9). FGF-19 is also expressed in the embryonic chicken eye where it may play a role in lens development (10). Questions regarding the functions of human FGF-19 have been addressed using ectopic expression. Transgenic over-expression of human FGF-19 in mice results in tumor formation, increased energy expenditure, decreased adiposity, and a resistance to weight gain in response to a high fat diet (11, 12). Similar affects have been observed in mice treated intravenously with recombinant FGF-19 (13).

II. OVERVIEW

A. PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for human FGF-19 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any human FGF-19 present is bound by the immobilized antibody. A biotinylated detection antibody specific for human FGF-19 is pipetted into the wells. After washing away any unbound substances, Streptavidin-HRP is pipetted into the wells. Following a wash to remove any unbound reagent, TMB substrate (Chromogenic agent) is added to the wells and color develops in proportion to the amount of human FGF-19 bound in the initial step. The color development is stopped, and the intensity of the color is measured.

B. LIMITATIONS OF THE PROCEDURE

- **FOR RESEARCH USE ONLY NOT FOR USE IN DIAGNOSTIC PROCEDURES.**
- This kit is suitable for cell culture supernate and human serum.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples generate values higher than the highest standard, dilute the samples with Calibrator Diluent (1×) and repeat the assay.
- Any variation in operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

III. ADVANTAGES

A. PRECISION

Intra-assay Precision (Precision within an assay)

Three samples were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

Three samples were tested in twenty separate assays to assess inter-assay precision.

	Intra-assay Precision			Inter-assay Precision		
Sample	1	2	3	1	2	3
Mean (pg/mL)	87.5	200.2	566.9	88.3	201.6	569.5
Standard Deviation	2.2	4.3	14.6	2.4	4.5	13.4
CV%	2.5	2.1	2.6	2.7	2.3	2.4

B. RECOVERY

The recovery of human FGF-19 spiked to different levels throughout the range of the assay in cell culture media was evaluated. The recovery ranged from 102.4 to 121.3% with an average of 112.6%.

The recovery of Human FGF-19 spiked to different levels throughout the range of the assay in human serum was evaluated. The recovery ranged from 101.5 to 113.5% with an average of 105.8%.

C. SENSITIVITY

The minimum detectable dose (MDD) of human FGF-19 is typically less than 3.86 pg/mL.

The MDD was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

D. CALIBRATION

This immunoassay is calibrated against highly purified *E.coli*-expressed recombinant human FGF-19 produced at R&D Systems.

E. LINEARITY

To assess the linearity of the assay, different samples were containing or spiked with high concentrations of human FGF-19 and diluted with Calibrator Diluent (1×) to produce samples with values within the dynamic range of the assay.

Dilution	Average % of Expected	Range (%)
1:2	103.6	86.7-106.7
1:4	101.0	82.3-111.1
1:8	99.1	80.0-113.3
1:16	95.5	81.4-106.3

F. SAMPLE VALUES

Human serum - Five human serum samples were evaluated for the presence of human FGF-19 in this assay. All samples measured ranged from 54.8 to 119.1 pg/mL with an average of 89.1 pg/mL.

G. SPECIFICITY

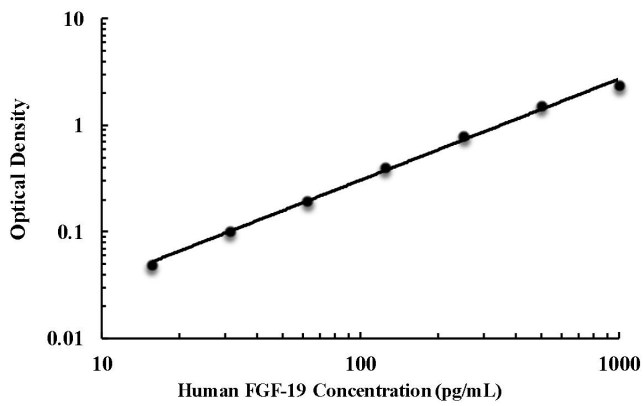
The following factors prepared at 50 ng/mL were assayed and exhibited no cross-reactivity or interference.

Recombinant human	Recombinant mouse
FGF R1 α /Fc Chimera	FGF-15
FGF R1 β /Fc Chimera	
FGF R2 α /Fc Chimera	
FGF R2 β /Fc Chimera	
FGF R3/Fc Chimera	
FGF R4/Fc Chimera	
FGF R5/Fc Chimera	

IV. EXPERIMENT

EXAMPLE STANDARD

The standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.



pg/mL	OD	Average	Corrected
0	0.015 0.015	0.015	—
15.6	0.065 0.063	0.064	0.049
31.3	0.118 0.113	0.116	0.101
62.5	0.209 0.212	0.211	0.196
125	0.423 0.400	0.412	0.397
250	0.820 0.799	0.810	0.795
500	1.507 1.535	1.521	1.506
1000	2.354 2.413	2.384	2.369

V. KIT COMPONENTS AND STORAGE

A. MATERIALS PROVIDED

Parts	Description	Size
Human FGF-19 Microplate	96 well polystyrene microplate (12 strips of 8 wells) coated with an antibody against human FGF-19.	1 plate
Human FGF-19 Standard	Recombinant human FGF-19 in a buffered protein base; lyophilized. Refer to the vial label for reconstitution volume.	2 vials
Human FGF-19 Detection Antibody	Biotinylated human FGF-19 antibody, lyophilized. Refer to the vial label for reconstitution volume.	1 vial
Calibrator Diluent (4×)	A 4× concentrated buffered diluent used to dilute standard and samples.	1 vial
Streptavidin-HRP A (200×)	200× Streptavidin conjugated to horseradish peroxidase.	1 vial
Reagent Diluent (10×)	A 10× concentrated buffered protein base used to dilute detection antibody and HRP.	1 vial
Wash Buffer Concentrate (25×)	A 25× concentrated solution of buffered surfactant with preservatives.	1 vial
TMB Substrate	TMB ELISA Substrate Solution/TMB Substrate Solution	1 vial
Stop Solution	2 N sulfuric acid.	1 vial
Plate Sealers	Adhesive strip.	3 strips

B. STORAGE

Unopened Kit	Store at 2-8 °C. Do not use past kit expiration date.	
Opened/ Reconstituted Reagents	Streptavidin-HRP A	May be stored for up to 1 month at 2-8 °C.*
	Wash Buffer (1×)	
	TMB Substrate	
	Stop Solution	
	Standard	Prepare fresh for each assay. Standards may be stored for up to 1 month at -20°C *.
	Detection Antibody	Aliquot and store for up to 1 month at -20 °C in a manual defrost freezer. *
	Reagent Diluent (10×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Reagent Diluent (1×). Prepare fresh for each assay.
	Calibrator Diluent (4×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Calibrator Diluent (1×). Prepare fresh for each assay.
Microplate Wells	Return unused wells to the foil pouch containing the desiccant pack, reseal along entire edge of zip-seal. May be stored for up to 1 month at 2-8 °C.*	

* Provided this is within the expiration date of the kit.

C. OTHER SUPPLIES REQUIRED

- ◆ Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- ◆ Pipettes and pipette tips.
- ◆ Deionized or distilled water.
- ◆ Squirt bottle, manifold dispenser, or automated microplate washer.
- ◆ Horizontal orbital microplate shaker capable of maintaining a speed of 500±50 rpm.
- ◆ 500 mL graduated cylinder.

D. PRECAUTION

- ◆ Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.
- ◆ The Stop Solution provided with this kit is an acid solution. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.

VI. PREPARATION

A. SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. Samples may require dilution with Calibrator Diluent (1 \times).

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 \times g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. Samples may require dilution with Calibrator Diluent (1 \times).

B. SAMPLE PREPARATION

Human serum samples recommend a 2-fold dilution. A suggested 2-fold dilution is 100 μL of sample + 100 μL of Calibrator Diluent (1 \times). Optimal dilutions should be determined by the end user.

C. REAGENT PREPARATION

Note: *Bring all reagents to room temperature before use.*

Wash Buffer (1 \times)- If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate (25 \times) into deionized or distilled water to prepare 500 mL of Wash Buffer (1 \times).

Calibrator Diluent (1 \times) - Use deionized or distilled water to prepare Calibrator Diluent (1 \times).

Reagent Diluent (1 \times) - Use deionized or distilled water to prepare Reagent Diluent (1 \times).

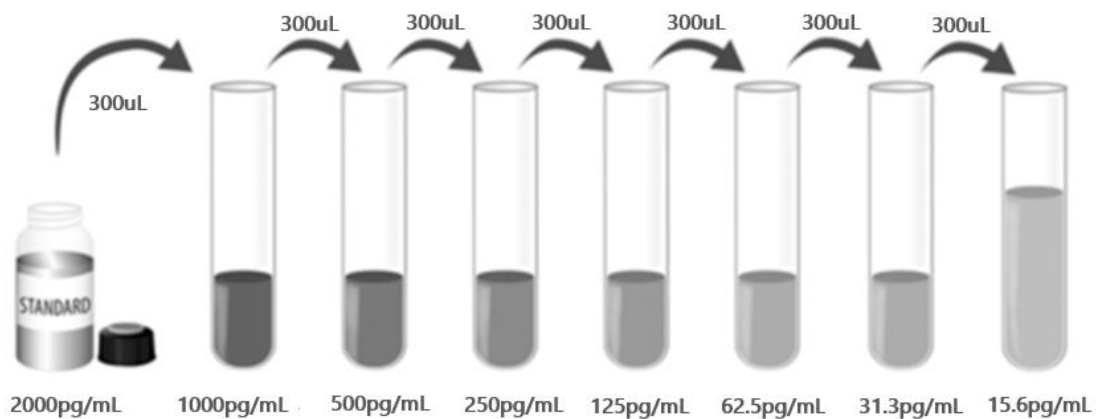
Detection Antibody (1 \times) - **Centrifuge briefly before opening. Reconstitution volume refer to vial label to prepare Detection Antibody (100 \times)**. Allow the Detection Antibody to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Aliquot and store if needed. Dilute to Detection Antibody (1 \times) with Reagent Diluent (1 \times). Prepare at least 15 minutes prior to use.

Streptavidin-HRP A (1 \times) - **Centrifuge briefly before opening.** Dilute to the working concentration specified on the vial label using Reagent Diluent (1 \times).

Human FGF-19 Standard - Centrifuge briefly before opening. Refer to the vial label for the reconstitution volume*. This reconstitution produces a stock solution of 2000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

*If you have any question, please seek help from our Technical Support.

Pipette 300 μ L of the appropriate Calibrator Diluent (1 \times) into each tube. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 1000 pg/mL standard serves as the high standard. The Calibrator Diluent (1 \times) serves as the zero standard (0 pg/mL).



D. TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- It is recommended that the samples be pipetted within 15 minutes.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- TMB Substrate should remain colorless until added to the plate. Keep TMB Substrate protected from light. TMB Substrate should change from colorless to gradations of blue.
- Stop Solution should be added to the plate in the same order as the TMB Substrate. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the TMB Substrate.

VII. ASSAY PROCEDURE

Note: Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 100 μ L of standard and prepared sample per well. Then add 50 μ L of the Detection Antibody (1 \times) diluted in Reagent Diluent (1 \times), to each well. Cover with the adhesive strip provided. **Incubate for 2 hours at room temperature on a horizontal orbital microplate shaker set at 500 \pm 50 rpm.** A plate layout is provided for a record of standards and samples assayed. (Samples may require dilution. See Sample Preparation section.)
4. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (400 μ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 μ L of the working dilution of Streptavidin-HRP A to each well. Cover the plate and **incubate for 20 minutes at room temperature on a horizontal orbital microplate shaker set at 500 \pm 50 rpm. Avoid placing the plate in direct light.**
6. Repeat the aspiration/wash as in step 4.
7. Add 100 μ L of TMB Substrate to each well. **Incubate for 10 minutes at room temperature on a horizontal orbital microplate shaker set at 500 \pm 50rpm. Avoid placing the plate in direct light.**
8. Add 50 μ L of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well within 10 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in

the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

10. **CALCULATION OF RESULTS**

Average the duplicate readings for each standard and sample and subtract the average zero standard optical density (O.D.). Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the human FGF-19 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

VIII. REFERENCES

1. Ornitz, D.M. and N. Itoh (2001) *Genome Biol.* **2**(3):1-12.
2. Eswarakumar, V.P. *et al.* (2005) *Cytokine Growth Factor Rev.* **16**:139.
3. Kim, I. *et al.* (2001) *Biochim. Biophys. Acta* **1518**:152.
4. Nishimura, T. *et al.* (1999) *Biochim. Biophys. Acta* **1444**:148.
5. Xie, M.H. *et al.* (1999) *Cytokine* **11**:729.
6. Ladher, R.K. *et al.* (2000) *Science* **290**:1965.
7. Yu, S. *et al.* (2002) *Am. J. Physiol. Endocrinol. Metab.* **283**:E490.
8. Harmer, N.J. *et al.* (2004) *Biochemistry* **43**:629.
9. Wright, T.J. *et al.* (2004) *Dev. Biol.* **269**:264.
10. Kurose, H. *et al.* (2005) *Dev. Growth Differ.* **47**:213.
11. Nicholes, K. *et al.* (2002) *Am. J. Pathol.* **160**:2295.
12. Tomlinson, E. *et al.* (2002) *Endocrinology* **143**:1741.
13. Fu, L. *et al.* (2004) *Endocrinology* **145**:2594.

PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

1								
2								
3								
4								
5								
6								
7								
8								
9								
10								
11								
12								
	A	B	C	D	E	F	G	H



产品信息及操作手册

人 FGF-19 Valukine™ ELISA 试剂盒

目录号: VAL148

适用于定量检测天然和重组人 FGF-19 的浓度

科研专用, 不可用于临床诊断

Bio-Techne China Co. Ltd

P: +86 (21) 52380373 P: 8009881270 F: +86 (21) 52381001

info.cn@bio-techne.com

有效期详见试剂盒包装标签

Novus 试剂盒确保在你收货日期 3 个月内有效

版本号 202409.3

目录

I. 背景	18
II. 概述	19
III. 优势	20
IV. 实验	22
V. 试剂盒组成及储存	23
VI. 实验前准备	25
VII. 操作步骤	27
VIII. 参考文献	28

I. 背景

成纤维细胞生长因子（FGF）家族至少包含22种高度保守的蛋白质，在线虫、果蝇和人类等许多物种中均有发现(1)。FGF是一种肝素结合生长因子，其核心120个氨基酸(aa)构成的FGF结构域，具有普通的三级结构。通常，FGFs在胚胎发育过程和受限性成人组织中表达。它们作用于中胚层和神经外胚层来源的细胞，以调节多种生理功能，包括血管生成，细胞生长，图式形成，代谢调节，细胞迁移，神经营养作用和组织修复(2)。FGF家族的活性由四种受体介导，即FGF R1至FGF R4(2)。这些受体是具有酪氨酸激酶结构域的跨膜蛋白，被认为是典型的酪氨酸激酶受体。FGF R5 / FGF RL1也有报道，但缺乏激酶结构域和信号传导能力(3)。

人FGF-19 cDNA序列预测了一个251 aa的前体蛋白，带有一个22 aa的信号肽和一个229 aa的分泌型成熟蛋白，且没有潜在的N-连接糖基化位点(4,5)。它与小鼠和大鼠FGF-15的氨基酸序列同源性约为50%，与鸡FGF-19的氨基酸序列同源性为62%(4-6)。与大多数结合并激活多个FGF受体的FGF不同，FGF-19似乎仅结合FGF R4(5)。受体结合能够激活MAP激酶并诱导催乳素启动子(7)。FGF-19有两个推测的二硫键，其中一个在FGF家族中是保守的(8)。特别的是，它还有一个扩展的环状结构二硫键，可能会影响硫酸乙酰肝素的结合(8)。

FGF-19的功能作用仍有待阐明，包括发育、代谢和致瘤活性方面。在鸡胚胎发生过程中，FGF-19已被证明会与Wnt-8c协同作用，启动内耳发育(6)。然而，小鼠的此活性不需要FGF-19的同系物FGF-15(9)。FGF-19也在鸡胚胎眼中表达，可能在晶状体发育中发挥作用(10)。关于人FGF-19功能的问题已经通过异位表达来验证。小鼠中人FGF-19的转基因过表达导致肿瘤形成，能量消耗增加，肥胖减少，以及对高脂肪饮食致使体重增加有所抵抗(11,12)。在静脉注射重组FGF-19的小鼠中也观察到类似的影响(13)。

II. 概述

A. 检测原理

本实验采用双抗体夹心ELISA法。抗人FGF-19抗体包被于微孔板上，样品和标准品中的人FGF-19会与固定在板上的抗体结合，接着加入生物素化的抗人FGF-19检测抗体进行孵育，洗涤去除未结合的物质后，加入链霉亲和素标记的辣根过氧化物酶 (Streptavidin-HRP) 孵育。洗涤未结合试剂后，加入TMB底物溶液（显色剂）。溶液颜色与结合的目标蛋白成正比；加入终止液；用酶标仪测定吸光度。

B. 检测局限

- ◆ 仅供科研使用，不可用于体外诊断；
- ◆ 该试剂盒适用于细胞培养上清样本和人血清样本；
- ◆ 请在试剂盒有效期内使用；
- ◆ 不同试剂盒及不同批号试剂盒的组分不能混用；
- ◆ 样本值若大于标准曲线的最高值，应将样本用标准品稀释液（1×）稀释后重新检测；
- ◆ 检测结果的不同可由多种因素引起，包括实验人员的操作、移液器的使用方式、洗板技术、反应时间或温度、试剂盒的效期等。

III. 优势

A. 精确度

板内精确度（同一板内不同孔间的精确度）

已知浓度的三个样本，在同一板内分别检测20次，以确定板内精确度。

板间精确度（不同板之间的精确度）

已知浓度的三个样本，在不同板中分别检测20次，以确定板间精确度。

样本	板内精确度			板间精确度		
	1	2	3	1	2	3
平均值 (pg/mL)	87.5	200.2	566.9	88.3	201.6	569.5
标准差	2.2	4.3	14.6	2.4	4.5	13.4
CV%	2.5	2.1	2.6	2.7	2.3	2.4

B. 回收率

在细胞培养基样本中掺入检测范围内不同水平的人FGF-19，测定其回收率。回收率范围在 102.4-121.3%，平均回收率在112.6%。

在人血清样本中掺入检测范围内不同水平的人FGF-19，测定其回收率。回收率范围在 101.5-113.5%，平均回收率在105.8%。

C. 灵敏度

人FGF-19的最低可测剂量（MDD）一般小于3.86 pg/mL。

MDD是根据20个重复的零标准品孔的吸光度值的平均值加两倍标准差计算得到的相对应浓度。

D. 校正

此ELISA试剂盒经由R&D Systems生产的*E.coli*表达的高纯度重组人FGF-19蛋白所校正。

E. 线性

不同的样本中含有或掺入高浓度的人FGF-19，然后用标准品稀释液（1×）将样本稀释到检测范围内，测定其线性。

稀释倍数	平均值/期待值 (%)	范围 (%)
1:2	103.6	86.7-106.7
1:4	101.0	82.3-111.1
1:8	99.1	80.0-113.3
1:16	95.5	81.4-106.3

F. 样本预值

人血清样本 - 使用本试剂盒检测了5份人血清样本中人FGF-19的水平。所有样本检测值在54.8-119.1 pg/mL之间，平均值为89.1 pg/mL。

G. 特异性

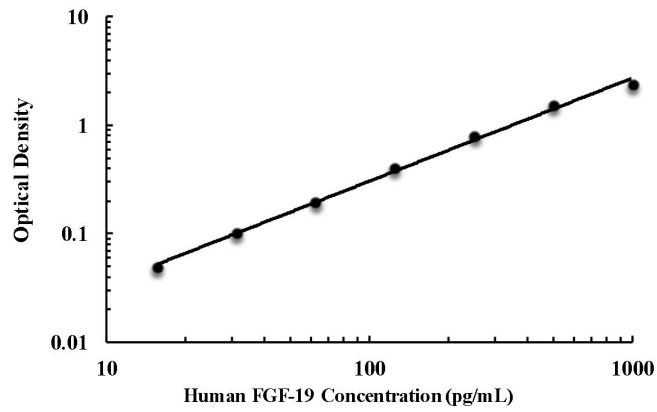
将以下因子配制成50 ng/mL的浓度来检测没有观察到明显的交叉反应或干扰。

Recombinant human	Recombinant mouse
FGF R1 α /Fc Chimera	FGF-15
FGF R1 β /Fc Chimera	
FGF R2 α /Fc Chimera	
FGF R2 β /Fc Chimera	
FGF R3/Fc Chimera	
FGF R4/Fc Chimera	
FGF R5/Fc Chimera	

IV. 实验

标准曲线实例

该标准曲线数据仅供参考，每次实验应绘制其对应的标准曲线。



pg/mL	OD	Average Corrected	
0	0.015 0.015	0.015	—
15.6	0.065 0.063	0.064	0.049
31.3	0.118 0.113	0.116	0.101
62.5	0.209 0.212	0.211	0.196
125	0.423 0.400	0.412	0.397
250	0.820 0.799	0.810	0.795
500	1.507 1.535	1.521	1.506
1000	2.354 2.413	2.384	2.369

V. 试剂盒组成及储存

A. 试剂盒组成

组成	描述	规格
Human FGF-19 Microplate	包被抗人 FGF-19 抗体的 96 孔聚苯乙烯板，8 孔×12 条	1 块板
Human FGF-19 Standard	人 FGF-19 标准品（冻干粉），参考瓶身标签进行重溶	2 瓶
Human FGF-19 Detection antibody	生物素化的抗人 FGF-19 检测抗体，冻干粉，参考瓶身标签进行重溶	1 瓶
Streptavidin-HRP A (200×)	200×浓缩的链霉亲和素标记的 HRP	1 瓶
Reagent Diluent (10×)	浓缩的试剂稀释液（10×）用于稀释检测抗体和 HRP	1 瓶
Calibrator Diluent (4×)	浓缩的标准品稀释液（4×）用于稀释标准品和样本	1 瓶
Wash Buffer Concentrate (25×)	浓缩洗涤缓冲液（25×）	1 瓶
TMB Substrate	TMB ELISA 底物溶液/TMB 底物溶液	1 瓶
Stop Solution	终止液	1 瓶
Plate Sealers	封板膜	3 张

B. 试剂盒储存

未开封试剂盒	2-8℃储存；请在试剂盒有效期内使用	
已打开，稀释或重溶的试剂	链霉亲和素-HRP A	2-8℃储存，最多 30 天*
	洗涤液（1×）	
	TMB 底物溶液	
	终止液	
	标准品	使用时新鲜配制* 标准品-20℃储存，最多 30 天*
	检测抗体	分装，-20℃储存，最多 30 天*
	标准品稀释液（4×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×标准品稀释液，多余的丢弃
	试剂稀释液（10×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×试剂稀释液，多余的丢弃
包被的微孔板条	将未用的板条放回带有干燥剂的铝箔袋内，密封：2-8℃储存，最多 30 天*	

*必须在试剂盒有效期内

C. 实验所需自备试验器材

- ◆ 酶标仪（可测量450 nm检测波长的吸收值及540 nm或570 nm校正波长的吸收值）
- ◆ 高精度加液器及一次性吸头
- ◆ 蒸馏水或去离子水
- ◆ 洗瓶（喷瓶）、多通道洗板器或自动洗板机
- ◆ 振荡器（速度可调至500±50 rpm）
- ◆ 500 mL量筒

D. 注意事项

- ◆ 试剂盒中的一些组分含有防腐剂，可能引起皮肤过敏反应，避免吸入。
- ◆ 试剂盒中的终止液是酸性溶液，使用时请做好眼睛、手、面部及衣服的保护。使用后请彻底洗手。

VI. 实验前准备

A. 样品收集及储存

细胞培养上清液：颗粒物应离心去除；立刻检测样本。样本收集后若不及时检测，需按一次使用量分装，冻存于 $\leq -20^{\circ}\text{C}$ 冰箱内，避免反复冻融。样本可能需要用标准品稀释液（1×）稀释。

血清样本：用血清分离管(SST)分离血清。使血样室温凝集30分钟，然后 $1000 \times \text{g}$ 离心15分钟。吸取血清样本之后即刻用于检测，或者分装， $\leq -20^{\circ}\text{C}$ 贮存备用。避免反复冻融。样本可能需要用标准品稀释液（1×）稀释。

B. 样本准备工作

人血清样本建议用标准品稀释液（1×）2倍稀释后进行检测，取 $100 \mu\text{L}$ 样本加到 $100 \mu\text{L}$ 标准品稀释液（1×）中，充分混匀，即制备成2倍稀释的样本。最佳稀释度应由最终用户确定。

C. 检测前准备工作

使用前请将所有试剂放置于室温

洗涤液（1×）：从冰箱中取出的浓缩洗涤液可能有结晶，属于正常现象；放置室温，轻摇混匀，待结晶完全溶解后再配制洗涤液。可将 20 mL 浓缩洗涤液（25×）用蒸馏水或去离子水稀释配制成 500 mL 工作浓度的洗涤液（1×）。

标准品稀释液（1×）：使用蒸馏水或去离子水稀释配制成标准品稀释液（1×）。

试剂稀释液（1×）：使用蒸馏水或去离子水稀释配制成试剂稀释液（1×）。

检测抗体（1×）：开盖前请瞬时离心。参考检测抗体瓶标签重溶冻干粉，制备检测抗体（100×）。轻轻震摇至少15分钟，其充分溶解。如有需要分装保存。用试剂稀释液（1×）稀释至检测抗体（1×），至少在使用前15分钟准备。

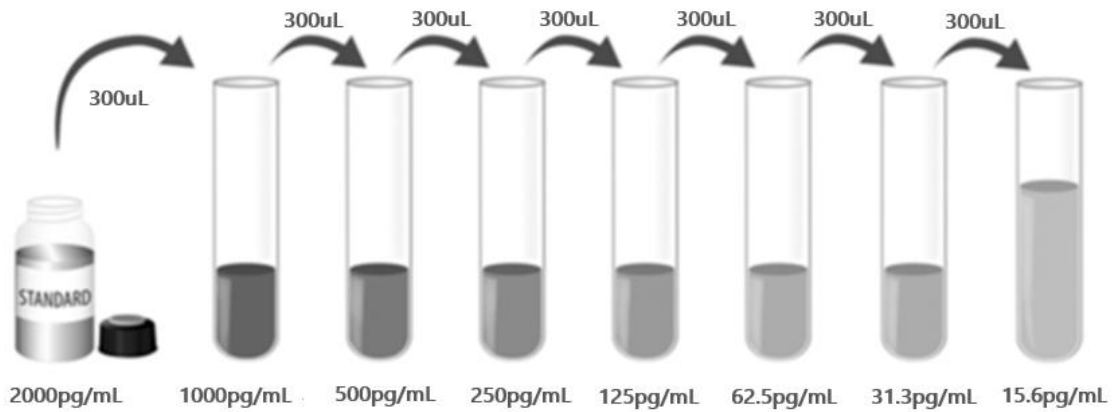
链霉亲和素-HRP A（1×）：开盖前请瞬时离心。用试剂稀释液（1×）将链霉亲和素-HRP A（200×）稀释至工作浓度。

人FGF-19标准品：开盖前请瞬时离心。冻干标准品的重溶体积请参考瓶身标签，得到浓度为 2000 pg/mL 标准品母液。轻轻振摇至少15分钟，使其充分溶解。

*如有疑问，请咨询我们的技术支持。

向各稀释管中加入 $300 \mu\text{L}$ 标准品稀释液（1×）。将标准品母液参照下图做系列稀释，每管须充分混匀后再移液到下一管。 1000 pg/mL 管作标准曲线最高点，标准品稀释液（1×）

可用作标准品零点（0 pg/mL）。



D. 技术小提示

- ◆ 当混合或重溶蛋白液时，尽量避免起沫；
- ◆ 为了避免交叉污染，配制不同浓度标准品、上样、加不同试剂都需要更换枪头。另外不同试剂请分别使用不同的移液槽；
- ◆ 建议15分钟内完成一块板的上样；
- ◆ 每次孵育时，正确使用封板膜可保证结果的准确性；
- ◆ TMB底物溶液在上板前应为无色，请避光保存；加入微孔板后，将由无色变成不同深度的蓝色；
- ◆ 终止液上板顺序应同TMB底物溶液上板顺序一致；加入终止液后，孔内颜色由蓝变黄；若孔内有绿色，则表明孔内液体未混匀，请充分混合。

VII. 操作步骤

使用前请将所有试剂和样本放置于室温，建议所有的实验样本和标准品做复孔检测

1. 按照上一节的说明，准备好所有需要的试剂和标准品；
2. 从已平衡至室温的密封袋中取出微孔板，未用的板条请放回铝箔袋内，重新封口；
3. 分别将不同浓度标准品和实验样本加入相应孔中，每孔100 μL 。然后每个微孔内加入50 μL 配制好的检测抗体（1 \times ），用封板膜封住反应孔，**室温500 \pm 50 rpm水平振荡孵育2小时**。说明书提供了一张96孔模板图，可用于记录标准品和试验样本的板内位置；（样本需要稀释，详情参见样本制备部分。）
4. 将板内液体吸去，使用洗瓶、多通道洗板器或自动洗板机洗板。每孔加洗涤液400 μL ，然后将板内洗涤液吸去。重复操作3次，共洗4次。每次洗板尽量吸去残留液体会有助于得到好的实验结果。最后一次洗板结束，请将板内所有液体吸干或将板倒置，在吸水纸拍干所有残留液体；
5. 在每个微孔内加入100 μL 稀释好的链霉亲和素-HRP A工作液。用封板膜封住反应孔，**室温500 \pm 50 rpm水平振荡孵育20分钟，注意避光**；
6. 重复第4步洗板操作；
7. 在每个微孔内加入100 μL TMB底物溶液，**室温500 \pm 50 rpm水平振荡孵育10分钟，注意避光**；
8. 在每个微孔内加入50 μL 终止液，请轻拍微孔板，使溶液混合均匀。孔内溶液颜色会从蓝色变为黄色；
9. 加入终止液后10分钟内，使用酶标仪测量450 nm的吸光度值，设定540 nm或570 nm作为校正波长。如果波长校正不可用，以450 nm的读数减去540 nm或570 nm的读数。这种减法将校正酶标板上的光学缺陷。没有校正而直接在450 nm处进行的读数可能会更高且更不准确；
10. **计算结果**：将每个标准品和样品的复孔吸光值取平均值，然后减去零标准品平均OD值（O.D.），使用计算机软件作四参数逻辑（4-PL）曲线拟合创建标准曲线。另一替代方法是，通过绘制y轴上每个标准品的平均吸光值与x轴上的浓度来构建标准曲线，并通过图上的点绘制最佳拟合曲线。数据可以通过绘制人FGF-19浓度的对数与O.D.的对数来线性化，并且最佳拟合线可以通过回归分析来确定。该程序将产生足够但不太精确的数据拟合。

如果样品被稀释，从标准曲线读取的浓度必须乘以稀释倍数。

VIII. 参考文献

1. Ornitz, D.M. and N. Itoh (2001) *Genome Biol.* **2**(3):1-12.
2. Eswarakumar, V.P. *et al.* (2005) *Cytokine Growth Factor Rev.* **16**:139.
3. Kim, I. *et al.* (2001) *Biochim. Biophys. Acta* **1518**:152.
4. Nishimura, T. *et al.* (1999) *Biochim. Biophys. Acta* **1444**:148.
5. Xie, M.H. *et al.* (1999) *Cytokine* **11**:729.
6. Ladher, R.K. *et al.* (2000) *Science* **290**:1965.
7. Yu, S. *et al.* (2002) *Am. J. Physiol. Endocrinol. Metab.* **283**:E490.
8. Harmer, N.J. *et al.* (2004) *Biochemistry* **43**:629.
9. Wright, T.J. *et al.* (2004) *Dev. Biol.* **269**:264.
10. Kurose, H. *et al.* (2005) *Dev. Growth Differ.* **47**:213.
11. Nicholes, K. *et al.* (2002) *Am. J. Pathol.* **160**:2295.
12. Tomlinson, E. *et al.* (2002) *Endocrinology* **143**:1741.
13. Fu, L. *et al.* (2004) *Endocrinology* **145**:2594.

96 孔模板图

请使用 96 孔模板图来记录标准品及样本在板内的位置

